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Air pollution and the fetal origin of disease: A systematic review of the
molecular signatures of air pollution exposure in human placenta

Leen J. Luyten^{1,2}, Nelly D. Saenen¹, Bram G. Janssen¹, Karen Vrijens¹, Michelle Plusquin,¹ Harry A.
Roels^{1,3}, Florence Debacq-Chainiaux², Tim S. Nawrot^{1,4}

¹Centre for Environmental Sciences, Hasselt University, Hasselt, Belgium;

²Unité de Recherche en Biologie Cellulaire (URBC) - Namur Research Institute for Life Sciences (Narilis),
University of Namur, Belgium

³Louvain Centre for Toxicology and Applied Pharmacology, Université catholique de Louvain, Brussels,
Belgium

⁴Department of Public Health & Primary Care, Occupational and Environmental Medicine, Leuven
University (KULeuven), Leuven, Belgium.

Correspondence to Prof. Dr. T.S. Nawrot, Centre for Environmental Sciences, Hasselt University,
Agoralaan Building D, 3590 Diepenbeek, Belgium. Telephone: 32-11-268382. Fax: 32-11-268299. E-
mail: tim.nawrot@uhasselt.be

Running title: Placental -omics and air pollution

ABSTRACT

Background Fetal development is a crucial window of susceptibility in which exposure-related alterations can be induced on the molecular level, leading to potential changes in metabolism and development. The placenta serves as a gatekeeper between mother and fetus, and is in contact with environmental stressors throughout pregnancy. This makes the placenta as a temporary organ an informative non-invasive matrix suitable to investigate omics-related aberrations in association with *in utero* exposures such as ambient air pollution.

Objectives To summarize and discuss the current evidence and define the gaps of knowledge concerning human placental -omics markers in association with prenatal exposure to ambient air pollution.

Methods Two investigators independently searched the PubMed, ScienceDirect, and Scopus databases to identify all studies published until January 2017 with an emphasis on epidemiological research on prenatal exposure to ambient air pollution and the effect on placental -omics signatures.

Results From the initial 386 articles, 25 were retained following an *a priori* set inclusion and exclusion criteria. We identified eleven studies on the genome, two on the transcriptome, five on the epigenome, five on the proteome category, one study with both genomic and proteomic topics, and one study with both genomic and transcriptomic topics. Six studies discussed the triple relationship between exposure to air pollution during pregnancy, the associated placental -omics marker(s), and the potential effect on disease development later in life. So far, no metabolomic or exposomic data discussing associations between the placenta and prenatal exposure to air pollution have been published.

Conclusions Integration of placental biomarkers in an environmental epidemiological context enables researchers to address fundamental questions essential in unraveling the fetal origin of disease and helps to better define the pregnancy exposome of air pollution.

Keywords: placenta, air pollution, child development, -omics, Barker hypothesis

47 **ABBREVIATIONS**

48	3-NTp:	3-nitrotyrosine
49	8-oxodG:	(8-oxo-2'-deoxyguanosine)
50	AHH:	Aryl hydrocarbon hydroxylase
51	BC:	Black carbon
52	BDNF:	Brain-derived neurotrophic factor
53	CI:	Confidence interval
54	CYP1A1:	Cytochrome P450 1A1
55	ECOD:	7-ethoxycoumarin O-deethylase
56	GST:	Glutathione S-transferase
57	GSTM1:	Glutathione S-transferase M1
58	LEP:	Leptin
59	miRNA:	MicroRNA
60	MT:	Metallothionein
61	mtDNA:	Mitochondrial DNA
62	NAT2:	N-acetyl transferase 2
63	NO ₂ :	Nitrogen dioxide
64	PAH:	Polycyclic aromatic hydrocarbon
65	PECO:	Population, Exposure, Comparator, and Outcome elements
66	PM:	Particulate matter
67	PM _{2.5} :	Particulate matter with a diameter smaller than 2.5 µm
68	PM ₁₀ :	Particulate matter with a diameter smaller than 10 µm
69	SO ₂ :	Sulfur dioxide
70	SYN1:	Synapsin 1

1. INTRODUCTION

Both genetic and environmental factors contribute to a multitude of complex diseases, while the precise environmental causes and early pathophysiological mechanisms of these diseases remain poorly understood (Ellis *et al.* 2014). The development of diseases can find its origin in every stage of human life. However, the distinct time windows, i.e. pregnancy, infancy, adolescence, adulthood, and old age are characterized by differences in age-specific susceptibilities (Cohen Hubal *et al.* 2008). During the last decade, a major public health concern has focused on the pregnancy period during which the exposure to harmful substances should be avoided to give the newborn the chance to start life as healthy as possible (Sun *et al.* 2016).

Over the entire intrauterine period, the placenta plays a crucial role for growth, development, and survival of the fetus (Burton *et al.* 2016). After the syncytiotrophoblast cells of the blastocyst have invaded the uterine wall, the placenta starts to grow with the formation of chorionic villi, which constitute the fetal side of this temporary organ (Figure 1). One of the first functions of placental cells is to suppress the maternal immune system in such a way that the developing embryo is not rejected (Nugent and Bale 2015). In later stages of pregnancy, the placenta develops a wide spectrum of functions to ensure proper fetal growth. It is endowed with an important transport function mediating the transfer of oxygen, nutritional components, growth factors, and hormones from mother to child, while carbon dioxide and other waste substances are transferred in the opposite direction (Levkovitz *et al.* 2013). This may occur by means of simple diffusion, (energy driven) transporter proteins, and endo- or exocytosis within complex matrices of different cell types, such as trophoblasts, amniotic cells, endothelium lining of the placental blood vessels, decidual cells, Hofbauer cells, and mesenchymal cells (Burton *et al.* 2016).

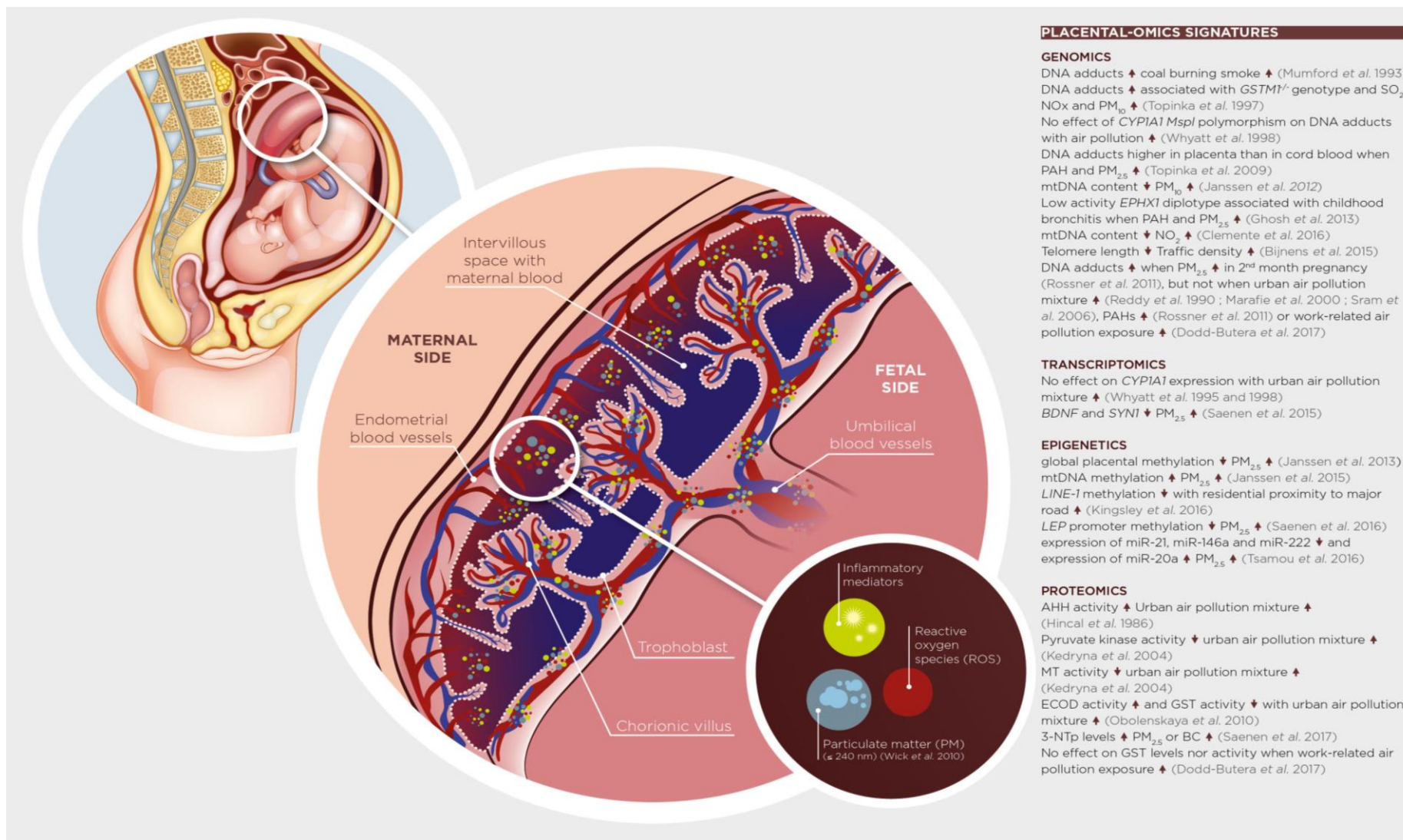


Figure 1: Placental migration of direct (particulate matter) and indirect (reactive oxygen species and inflammatory mediators) potential effectors of exposure to air pollution during pregnancy. The column on the right summarizes the -omics characteristics (genomics, transcriptomics, epigenetics, and proteomics) as described in this systematic review in association with exposure to *in utero* ambient air pollution.

In this way, the placenta comes in contact with, contains and interacts with the substances to which both mother and fetus are exposed to during the timespan of the entire pregnancy. In addition, the placenta itself is an important endocrine organ regulating the production of hormones such as progesterone, human chorionic gonadotrophin (hCG), and human placental lactogen (hPL), to ensure the continuation of pregnancy and to acquire the appropriate maternal responses to optimize the development of the fetus (Burton *et al.* 2016; Nugent and Bale 2015). Furthermore, within the fetoplacental unit, a great number of signals are sent from the placenta to the fetus - and *vice versa* - to regulate developmental processes (Dötsch *et al.* 2010). Such signals can also elicit the appropriate reactions to various environmental exposures. Together, all these properties make the placenta an essential organ for the regulation of fetal development. Indeed, placental dysfunction has been linked to for example the occurrence of preeclampsia and adverse birth outcomes such as intrauterine growth restriction (Cha and Kim 2010).

Intrauterine exposure to pollutants can lead to altered metabolic functions that may be detrimental for fetal development. For example, the embryonic brain has a great plasticity and its development depends on, and can be influenced by, various environmental factors (Buss *et al.* 2013). The etiology of diseases in adulthood may have a fetal origin and may be attributed to the effects of adverse environmental exposures *in utero*. This causality concept is known as the Barker hypothesis or the Developmental Origins of Health and Disease (DOHaD). Professor David Barker was the first to recognize this potential link when he became concerned about the association between malnutrition during pregnancy and the development of coronary heart disease in adult life (Barker 1995). Since then, many implications of this hypothesis have been reported (Deng *et al.* 2016, 2017; Lu *et al.* 2017). Adverse environmental exposures during pregnancy already identified in this context are active and passive cigarette smoke (Mund *et al.* 2013), and exposure to ambient air pollution [including nitrogen dioxide (NO₂) (Ballester *et al.* 2010), polycyclic aromatic hydrocarbons (PAH) (Jedrychowski *et al.* 2015), and particulate matter (PM) (Rappazzo *et al.* 2014)]. Particles with a diameter smaller than 500 nm are known to pass the placental barrier during the gestational period, while particles with a diameter

smaller than 240 nm are even able to reach the fetal bloodstream (Wick *et al.* 2010) (Figure 1), possibly affecting the newborn's metabolism before birth.

Various reviews have already described the associations between prenatal ambient air pollution exposure and birth outcomes such as prematurity and birth weight (Lamichhane *et al.* 2015; Shah and Balkhair 2011). However, none of these reviews described the placenta as an intermediate matrix having the potential to express distinct biological (-omics) signatures associated with prenatal exposure to ambient air pollution. Hence, the goal of this systematic review is to provide a structured overview and an evaluation of the current knowledge on the potential of placental tissue as a non-invasive biological matrix for the study of molecular -omics signatures that are associated with *in utero* exposure to ambient air pollution and are probably useful as early-life markers of disease development later in life. With this systematic review we aim to identify signatures in the -omics fields that already have been well addressed and those of which a substantial gap of knowledge still remains in the scope of epidemiological research involving the placenta as a tissue to identify sentinel biological effects of air pollution exposure during pregnancy.

2. MATERIALS AND METHODS

The goal of this systematic review was to provide an answer to the question: “Which -omics biomarkers have been analyzed in human placental tissue used as a non-invasive matrix in epidemiological research in association with prenatal exposure to air pollution in the context of disease development later in life?”. The PECO elements that can be deduced from this question were used to determine the selection criteria to search and structure the articles for the synthesis of this review. These PECO elements are:

- “Population”: human. In this article we focused on research conducted in an epidemiological context, thus not including research on human cell lines.
- “Exposure”: ambient air pollution [including particulate matter with particles smaller than 2.5 μm (PM_{2.5}), particulate matter with particles smaller than 10 μm (PM₁₀), ultrafine particles, black carbon (BC), derivatives of nitrogen oxide (NO_x), and polycyclic aromatic hydrocarbons (PAHs)]. We defined ambient air pollution as a mixture of indoor and outdoor pollutants, in both solid and gaseous form, and we excluded direct (maternal) or indirect (environmental) exposure to tobacco smoke from this concept.
- “Comparator”: in this review were included both studies in which comparisons are made between groups exposed to either a higher or a lower concentration of air pollution, as well as studies with a continuous exposure scale.
- “Outcomes”: placental -omics biomarkers and, if discussed, disease development or the development of adverse birth outcomes.

This systematic review was constructed according to existing guidelines on the structure of systematic reviews and maps (Bates *et al.* 2007). An online database search was performed in January 2017, according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines (<http://www.prisma-statement.org>) to identify articles that are dealing with the scope of this review, without any limitation set on the publication date. Two investigators (LJL and NDS) were

appointed to conduct the literature search, because of their expertise on the effects of air pollution in the placenta. These investigators read all papers, extracted, and archived the relevant information independently. The level of consensus between LIL and NDS was determined by performing a Cohen's kappa analysis. Any remaining discrepancies were resolved by consensus. The exploration was conducted on PubMed (<http://www.ncbi.nlm.nih.gov/pubmed>), Scopus (<http://www.scopus.com/>), and ScienceDirect (<http://www.sciencedirect.com>). Only English MeSH-terms were used to form the search strings. First, a search was conducted with the key terms "placenta" and "air pollution". Next, additional searches were performed by replacing these terms with related search queries (for a list of all used queries see Supplemental Tables S1 and S2). Additionally, since we were interested in the link between -omics in the placenta and the development of disease, we replaced the air pollution-related MeSH-terms with the MeSH-terms "fetal origin adult disease", "barker hypothesis", "barker hypothesis fetal" and "barker hypothesis fetal origins" in the identification phase (see Supplemental Table S1 and S2). Only primary research was included in this paper: in case a review article was found in the literature search, the list of references in this review was checked manually to determine if additional articles could be identified that met the inclusion criteria of this systematic review. If a full text could not be obtained, a request was sent via ResearchGate (<https://www.researchgate.net/>) or via the website of the journal in which the article was published. In search for potential additional information from grey literature, we used a popular search engine (<http://www.google.com>), and accessed the OpenGrey (<http://www.opengrey.eu>), and Cochrane Library (<http://onlinelibrary.wiley.com/cochranelibrary/>) websites. First of all we read the abstract of all papers that were found from the identification procedure and excluded the research articles on animals or human cell lines, since we wanted to put the emphasis solely on epidemiological research. The comparison of differences in placental -omics signatures between different (animal) models is beyond the scope of this systematic review. We also excluded comments on other research articles and the papers not written in English to avoid potential misinterpretation of the results due to incorrect translation. Subsequently, we examined the full text of the remaining articles and excluded

188 those studying exclusively the effect of maternal active and/or passive smoking during pregnancy on
189 placental -omics signatures or fetal health. These articles were excluded because air pollution is a
190 complex mixture that takes into account the effects of various sources, such as traffic- and industry-
191 related pollution, while research on smoking only focusses on the effects of tobacco use. Additionally,
192 research articles that did not consider the measurement of -omics markers in the placenta were not
193 included, because this review specifically focusses on the effects of air pollution exposure during
194 pregnancy on the -omics biomolecular signatures of the placenta. For the remaining articles that were
195 included in this systematic review, the content was examined in detail with a great focus on (i) the
196 placental -omics marker(s) studied and the techniques used to measure them, (ii) the characteristics
197 of prenatal exposure to ambient air pollution in association with the placental -omics marker(s), and
198 (iii) whether the authors mentioned any association with disease development later in life. Finally, a
199 descriptive analysis of these articles was made and a summary of the current knowledge has been
200 provided based on the different -omics fields (genomics, epigenetics, transcriptomics, proteomics and
201 metabolomics). In this way, existing gaps of knowledge in this research field could be established.

3. RESULTS

Using the initial MeSH-terms “placenta” and “air pollution”, 118 articles could be identified (Figure 2).

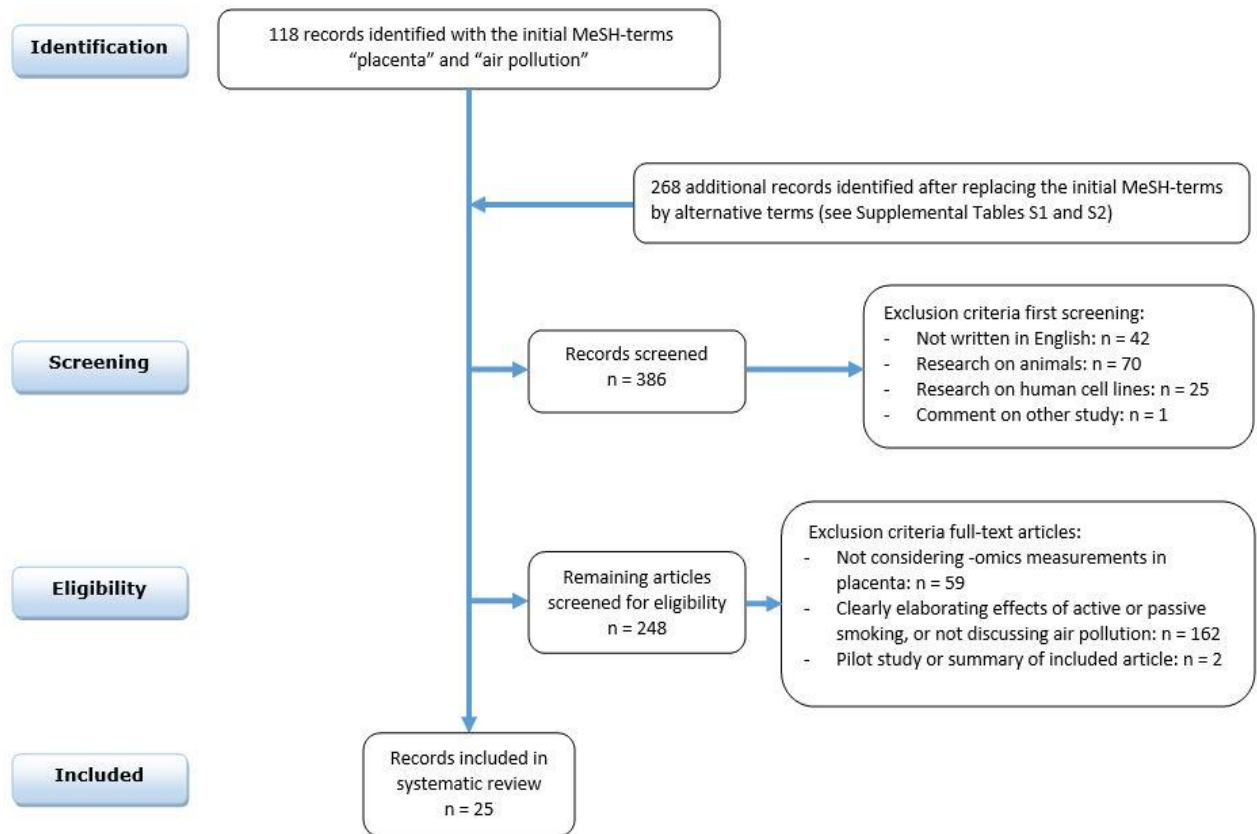


Figure 2: Flowchart of the selection protocol according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines. From the 386 initially screened articles, 25 were included in this systematic review.

Replacing the MeSH-terms by alternative terms (see Supplemental Tables S1 and S2), 268 additional records could be added to the list. No new articles were identified from reference lists of other reviews and no additional information could be retrieved from grey literature. From the total of 386 articles, 42 were excluded because they were not written in English. One study was excluded since it was a comment on another research article. The abstracts of the remaining articles were scanned for eligibility based on whether they pertained to epidemiological research. We excluded 70 animal studies and 25 studies using human cell lines. Of the remaining 248 articles, 59 were excluded because they did not report -omics measurements in human placental tissue and 162 were not included since

the article only elaborated on the effects of active or passive maternal smoking and not on concomitant effects of exposure to ambient air pollution during the gestational period. One study of Topinka *et al.* (1997a) was considered a pilot study of one of the remaining articles of these authors (Topinka *et al.* 1997b), and one article of Sram *et al.* (1999) summarized the latter study, so the results of these three studies were discussed simultaneously. The inter-rater variability as determined by the Cohen's kappa analysis was 0.98 (95% confidence interval: 0.96 – 0.99), which can be regarded as a value indicating an almost perfect agreement between LIL and NDS.

Twenty-five studies (Supplemental Table S3 and Figure 1) met all the selection criteria. The publication dates of the articles ranged from August 1990 to September 2016. Six articles discussed the triple relationship involving *in utero* air pollution exposure leading to molecular changes in placental tissue, with a direct or indirect descriptive link to adverse birth outcomes and/or the development of (chronic) diseases (Clemente *et al.* 2016; Ghosh *et al.* 2013; Hincal 1986; Kingsley *et al.* 2016; Rossner *et al.* 2011; Sram *et al.* 2006). Five out of these six studies investigated a change in birth weight as an adverse outcome, three out of these five also looked at growth restriction (Hincal 1986; Kingsley *et al.* 2016; Rossner *et al.* 2011), and one article studied prematurity of the neonate as an additional detrimental birth outcome (Sram *et al.* 2006). Only one of these five studies investigated air pollution exposure during pregnancy, while looking at the associations with placental -omics markers and the development of a disease outcome later in life, namely childhood bronchitis (Ghosh *et al.* 2013).

All 25 studies were observational, conducted in an epidemiological context, and used the placenta as a biological matrix to study molecular effects of prenatal ambient air pollution exposure. Among these studies, all categories of -omics markers were covered with exception of the placental metabolome and exposome. We identified eleven studies on the genome, five on the epigenome, two on the transcriptome, five on the proteome, one study with both genomic and transcriptomic topics, and one study covered topics on both genomics and proteomics (Supplemental Figure 1). The 25

included research articles showed a bottom-up approach for all -omics categories , focusing on specific preselected targets and their association with prenatal exposure to ambient air pollution.

Twelve of the 25 articles discussed the effects of PM air pollution on placental –omics (Supplemental Figure 2). More specifically, three studies investigated PM₁₀ (one study in combination with other forms of air pollution, namely PAHs, SO₂ and NO_x), while nine studies investigated PM_{2.5}. Exposure to PM_{2.5} was often studied in combination with other air pollution components, such as PAHs (three studies) and black carbon (one study). Other forms of ambient air pollution were discussed separately as well, such as NO_x (one study), and PAHs (one study). Seven articles analyzed a comparison of two groups of participants, based on their exposure to urban air pollution. Finally, four articles used proxies for air pollution exposure, such as the distance of the residence to a major road, residential traffic density, work-related air pollution exposure in maquiladoras (factories at the border between Mexico and the USA), and smoke from residential coal burning as a heating source.

4. DISCUSSION

4.1. Placental tissue in epidemiological research: advantages and disadvantages

All 25 studies that were selected for discussion in this review used placental tissue as a biological matrix for epidemiological research purposes. This temporary organ has the advantage that it can serve to evaluate biological outcomes of environmental exposures simultaneously in tissue with both maternal and fetal origin. Moreover, the sampling of placental tissue requires no invasive procedure, avoiding unnecessary potential damage to the fetus. The placenta shows to be a crucial tissue to study certain developmental processes, since it provides the necessary molecules for these mechanisms. In mice it has been shown that this organ produces serotonin at the earliest phases of pregnancy, which is an important factor in the development of the fetal central nervous system (Bonnin *et al.* 2011). Five studies discussed in this review made a link between biomolecular characteristics of the placenta and health conditions that could interfere with human development later in life, more specifically a

decrease in birth weight (Clemente *et al.* 2016; Hincal 1986; Rossner *et al.* 2011; Sram *et al.* 2006), fetal growth restriction (Hincal 1986; Rossner *et al.* 2011) or the development of bronchitis in early childhood (Ghosh *et al.* 2013). This shows that the placenta has the potential to serve as a tissue to study the link between prenatal exposures and the effects on the (mal-)development of children in early life. Apart from the different functions of umbilical cord blood and the placenta during pregnancy, several molecular differences between both matrices have been identified such as different turnover rates of mitochondrial DNA (mtDNA) (Janssen *et al.* 2012). In contrast to cord blood, which can encompass the effects of environmental exposures on the short term, the placenta can reflect the cumulative effect of prenatal exposures over the pregnancy period. In the context of the evaluation of exposure conditions on fetal development, biomolecular measurements in placental samples can be particularly useful since it has been suggested that changes in the placenta could be involved in the epigenetic regulation of fetal development, possibly to a slightly greater extent than in cord blood (Nomura *et al.* 2014).

A disadvantage of using placental tissue for research purposes is that obtaining representative sample aliquots is challenging as the placenta is composed of a heterogeneous mix of cells, blood vessels, chorionic villi, and membranes. Therefore, standardization of placental sampling is of great importance to account for the complexity of this tissue. Moreover, the sampling procedures carried out in several studies and cohorts using different protocols could introduce variability in the observed results and the conclusions drawn from this research. When comparing the sampling methods of the 25 studies included in this review, differences were identified in terms of sampling position on the placenta, the placental layers which were sampled, and the size of the tissue samples [ranging from 1-2 cm³ (Janssen *et al.* 2012) to 50 g (Obolenskaya *et al.* 2010)]. In the context of relatively large numbers of samples or subjects under investigation in epidemiological studies and the related costs for molecular measurements, an additional disadvantage is that it is not always feasible to analyze multiple samples from the same placenta. Observational studies may consider pooling several biopsies of one placenta to further reduce sample variability. Suggestions for a more standardized protocol

have already been made by Burton *et al.* (2014), with regard to speed of sampling, aliquoting and preservation of the tissue to ensure sufficient quality of the DNA, RNA, and proteins for further analyses. These authors advice to use a standardised grid to sample each placenta at minimal four different sites, take samples of 1-2cm³, and divide these biopsies into smaller aliquots according to your -omics field of interest, and quickly snap freeze the samples after rinsing them in phosphate-buffered saline (PBS) at 4°C (Burton *et al.* 2014).

4.2. Placental -omics signatures of prenatal air pollution exposure

At delivery, the placenta is a representative source of the morphological, functional, biological, and molecular information that has been accumulated during gestation. Therefore, it is a suitable matrix for postnatal investigation of potential associations between molecular (-omics) signatures and prenatal environmental influences. Several biomolecular characteristics related to diverse toxicological exposures have already been investigated in placental tissue. Not only direct DNA damage, but also changes in -omics (genomics, epigenetics, transcriptomics, proteomics, metabolomics and exposomics) signatures can occur due to hazardous environmental exposures such as ambient air pollution (Table 1). These alterations may possibly provide early effect predictors for human health risk due to *in utero* environmental exposures (Fowler 2012). In this context, characteristic biomolecular signatures measured in humans may be considered biomarkers - which can be a chemical or its metabolite - biomolecules, or the product of an interaction between a substance and a target molecule or cell (World Health Organization 2010). The measurement of placental -omics markers can provide useful insights on gestational exposure effects, susceptibility, and disease risk of the neonate (Fowler 2012; Ryan *et al.* 2012). Despite the fact that several changes in -omics fields have been characterized in placental tissue in association with air pollution exposure, two fields - metabolomics (discussed below) and exposomics - could not be sufficiently covered in the context of this systematic review because of the lack of studies on these topics.

319 **Table 1.** -Omics categories and placental markers analyzed in association with exposure to ambient air pollution during the gestational period

-Omics category	Placental markers
Genomics	<ul style="list-style-type: none"> - Telomere length (Bijnens <i>et al.</i> 2015) - Mitochondrial DNA content (Clemente <i>et al.</i> 2016; Janssen <i>et al.</i> 2012) - Presence of the low activity <i>EPHX1</i> (His/His) diplotype (Ghosh <i>et al.</i> 2013) - Presence of the <i>CYP1A1</i> MspI polymorphism (Whyatt <i>et al.</i> 1998) - DNA adduct levels (Dodd-Butera <i>et al.</i> 2016; Marafie <i>et al.</i> 2000; Mumford <i>et al.</i> 1993; Reddy <i>et al.</i> 1990; Rossner <i>et al.</i> 2011; Sram <i>et al.</i> 2006; Topinka <i>et al.</i> 1997, 2009; Whyatt <i>et al.</i> 1998)
Epigenetics	<ul style="list-style-type: none"> - Global DNA methylation level (Janssen <i>et al.</i> 2013) - LINE-1 and AluYb8 DNA methylation levels (Kingsley <i>et al.</i> 2016) - Mitochondrial DNA methylation level (Janssen <i>et al.</i> 2015) - <i>LEP</i> promoter methylation (Saenen <i>et al.</i> 2017) - Levels of miR-21, miR-146a, miR-222, and miR-20a (Tsamou <i>et al.</i> 2016)
Transcriptomics	<ul style="list-style-type: none"> - Expression levels of <ul style="list-style-type: none"> - <i>BDNF</i> (Saenen <i>et al.</i> 2015) - <i>SYN1</i> (Saenen <i>et al.</i> 2015) - <i>CYP1A1</i> (Whyatt <i>et al.</i> 1995, 1998)
Proteomics	<ul style="list-style-type: none"> - 3-NTp level (Saenen <i>et al.</i> 2016) - Amount of metallothionein (Sorkun <i>et al.</i> 2007) - GST level (Dodd-Butera <i>et al.</i> 2016) - Activity of <ul style="list-style-type: none"> - AHH (Hincal 1986) - Pyruvate kinase (Kedryna <i>et al.</i> 2004) - GST (Dodd-Butera <i>et al.</i> 2016; Obolenskaya <i>et al.</i> 2010) - ECOD (Obolenskaya <i>et al.</i> 2010)
Metabolomics	/

320

321 Abbreviations: 3-NTp, 3-nitrotyrosine; AHH, Aryl hydrocarbon hydroxylase; *BDNF*, Brain-derived neurotrophic factor; *CYP1A1*, Cytochrome

322 (*CYP*) P450 1A1; ECOD, 7-ethoxycoumarin O-deethylase; *EPHX1*, Epoxide hydrolase 1; GST, Glutathione S-transferase; His, Histidine; *LEP*, Leptin;

323 miR, MicroRNA; *SYN1*, Synapsin 1

The field of exposomics encompasses all the environmental exposures for an organism during its lifetime (Wild 2012). Placental exposomics have for example been studied in mothers known to be obese or diabetic at the moment of gestation (Lewis *et al.* 2013). In case of investigating the effects of *in utero* exposure to ambient air pollution one study can be cited which measured asbestos fibers as a part of the exposome in the placentas of stillborn babies (Haque *et al.* 1992). Several intermediate markers including telomere length and microRNA (miRNA) expression patterns have been studied as a proxy-effect of ambient air pollution exposure on exposomics (Martens and Nawrot 2016; Vrijens *et al.* 2015). However, the full placental exposome regarding environmental air pollution exposure is a complex entity of which the parts still need to be assembled.

4.2.1. Genomics (Table 2)

Direct DNA damage and damage through DNA adducts were two of the first placental markers used to evaluate the health significance of genomic insults through prenatal ambient air pollution exposure. As early as 1990, ³²P-postlabeling was performed in placental tissue to study the extent of DNA damage that could be inflicted by exposure to PAHs during pregnancy (Reddy *et al.* 1990). Ten years later, a similar study was published on DNA adducts in placental samples using two different techniques, i.e. nuclease P1 and butanol extraction enhancement prior to ³²P-postlabeling (Marafie *et al.* 2000). Both studies came to the same conclusion: the levels of placental DNA adducts did not differ significantly between women exposed to airborne PAHs by either residential wood combustion (Reddy *et al.* 1990) or pollution from oil well fires (Marafie *et al.* 2000) compared with non-exposed women. In a recent study, lack of association was also found between placental PAH-adducts and exposure to work-related air pollution at the US-Mexican border (Dodd-Butera *et al.* 2016). Mumford *et al.* (1993) came to the opposite conclusion in a study on placental DNA-adduct levels and PAH exposure during pregnancy: the adduct levels increased when mothers were exposed to smoke of coal burning during pregnancy, however, these results lack statistical confirmation. Furthermore, a study on ambient PM_{2.5} and PM₁₀ air pollution also did not show an association between placental DNA-adduct levels and exposure to air pollution (Sram *et al.* 2006). Hence, the consistent negative results from these

independent studies may point to a molecular effect other than the formation of DNA adducts in the placenta associated with maternal air pollution exposure. Topinka *et al.* (2009) compared placental adduct levels with those in cord blood following *in utero* exposure to PAHs and PM_{2.5}, and showed that the total level of DNA adducts was significantly higher in cord blood compared to placenta. Other studies in cord blood also showed positive relationships between DNA adduct levels and exposure to air pollution (Pedersen *et al.* 2009), which indicates that these hazardous airborne substances could affect DNA adduct levels in other tissues than the placenta.

Fetuses are able to adapt their mitochondrial structure and metabolism when the supply of nutrients is limited or compromised. Mitochondria are the biochemical power plants of cells providing energy through the production of adenosine-5'-triphosphate (ATP) via oxidative phosphorylation. These intracellular organelles contain multiple copies of circular DNA - mitochondrial DNA (mtDNA) - of approximately 16 kb in length which are vulnerable to reactive oxygen species (ROS) because of close proximity to the electron transport chain and inefficient DNA repair capacity (Linnane *et al.* 1989). The estimated mutation rate of mtDNA is 5-15 times higher compared to nuclear DNA (Payne *et al.* 2013). Changes in placental mtDNA content may represent a biological effect along the path linking air pollution to adverse effects on the unborn. In placental tissue of 174 mother-newborn pairs of the Belgian birth cohort ENVIRONAGE, an inverse association was found between third trimester PM₁₀ (and NO₂) exposure and placental mtDNA content (-17.4%, 95% CI: -31.8 to -0.1%, for an increment of 10 µg/m³ in PM₁₀ exposure; p = 0.05) (Janssen *et al.* 2012).

A similar inverse association was found in the Spanish INMA birth cohort between placental mtDNA content and gestational exposure to traffic-related NO₂ air pollution (-4.9%, 95% CI: -7.9 to -1.8% for an increment of 10 µg/m³ in NO₂ exposure; p = 0.003) (Clemente *et al.* 2016). The discrepancy in effect-size can be explained by the very dynamic nature of placental mtDNA.

373 **Table 2.** Studies describing the associations between prenatal ambient air pollution exposure and changes in placental genomic markers

Author	Study population	Increase in analyzed air pollution component (average \pm standard deviation if available)	Effect on placental -omics marker
Reddy <i>et al.</i> (1990)	4 non-smoking women exposed to wood smoke during pregnancy and 5 non-exposed controls from Massachusetts	Urban air pollution	No significant differences in DNA-adduct levels between exposed and non-exposed mothers.
Mumford <i>et al.</i> (1993)	38 placental samples from Xuan Wei (China) exposed to coal combustion smoke during pregnancy and 19 samples from controls living in Beijing, using natural gas as heating source.	Smoke from coal combustion	Total DNA-adducts detected in 52% of placentas of exposure group compared to 5.3% of the samples of the control group (no statistics performed)
Topinka <i>et al.</i> (1997)	158 mothers (113 non-smokers and 45 smokers) in two districts of the Czech Republic with different exposure levels of air pollution	Average monthly concentrations of SO ₂ , NO _x , PAH and PM ₁₀ from January 1994 until January 1995	Increased levels of DNA-adducts in samples of the highly exposed regions compared to the lower exposed regions in placentas with the <i>GSTM1</i> null genotype ($p = 0.029$) No effect of NAT2 genotype on DNA adduct levels in correlation with air pollution exposure
Whyatt <i>et al.</i> (1998)	70 subjects from Krakow with higher levels of air pollution and 90 subjects from Limanowa, a less polluted city in Poland	Average annual concentration of 37 $\mu\text{g}/\text{m}^3$ of ambient respirable particles in least exposed group and 78 $\mu\text{g}/\text{m}^3$ in the most exposed group, in the year prior to delivery (particle size not defined)	No significant associations between PAH-adduct levels, presence of the <i>CYP1A1 MspI</i> polymorphism and exposure to air pollution.
Marafie <i>et al.</i> (2000)	40 mothers exposed to oil well fires and 180 non-exposed Kuwaiti mothers	Urban air pollution	No significantly different levels of DNA-adducts between mothers of different exposure groups
Sram <i>et al.</i> (2006)	199 subjects born between 1994 and 1995 for DNA-adduct analyses and 1013 subjects born between 2000 and 2002 for genotyping. All samples collected in two districts of the Czech Republic with different levels of air pollution	Urban air pollution	No significant associations between placental DNA-adduct levels and birth weight, and no effects of air pollution on birth weight or DNA-adduct levels identified.
Topinka <i>et al.</i> (2009)	Placentas from 79 individuals born in 2007 and 2008 in Prague (Czech Republic)	B[a]P, PAHs and PM _{2.5} levels (no mean values provided)	Total DNA-adduct levels are significantly higher in cord blood compared to placental tissue ($p < 0.001$)
Rossner <i>et al.</i> (2011)	891 subjects born between 1994 and 1999 in two districts of the Czech Republic with either high or low levels of air pollution exposure	Average concentrations of PAHs and PM _{2.5} for each month of pregnancy	No significant associations between 8-oxodG-adduct levels and PAH levels, but a significant increase in 8-oxodG-adduct levels with increased PM _{2.5} exposure in second month of pregnancy (OR = 1.68, 95% CI: 1.28 to 2.19; $p < 0.001$)

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375 Abbreviations: 8-oxodG, (8-oxo-2'-deoxyguanosine); B[a]P, Benzo[a]pyrene; CI, Confidence interval; *EPHX1*, Epoxide hydrolase 1; *GSTM1*, Glutathione S-transferase M1; mtDNA, Mitochondrial
376 DNA; NAT2, N-acetyl transferase 2; NO₂, Nitrogen dioxide; NO_x, Nitrogen oxides; OR, Odds ratio; PAH, Polycyclic aromatic hydrocarbon; PM_{2.5}, Particulate matter with a diameter smaller than
377 2.5 μm ; PM₁₀, Particulate matter with a diameter smaller than 10 μm ; RTL, Relative telomere length; SO₂, Sulfur dioxide.

378 **Table 2. (continued)**

Author	Study population	Increase in analyzed air pollution component (average \pm standard deviation if available)	Effect on placental -omics marker
Janssen <i>et al.</i> (2012)	174 individuals from the ENVIRONAGE cohort (Belgium)	10 $\mu\text{g}/\text{m}^3$ increase in PM_{10} ($22.7 \pm 3.7 \mu\text{g}/\text{m}^3$ for entire pregnancy)	16.1% decrease in mtDNA content in association with exposure during the last month of pregnancy (95% CI: -25.2 to -6.0% , $p = 0.003$)
Ghosh <i>et al.</i> (2013)	$n = 793$ randomly selected from children born between 1994 and 1998 in two districts of the Czech Republic	100 ng/m^3 increase in PAH ($63.4 \text{ ng}/\text{m}^3 \pm 51.5 \text{ ng}/\text{m}^3$) and a 25 $\mu\text{g}/\text{m}^3$ increase in $\text{PM}_{2.5}$ ($22.8 \mu\text{g}/\text{m}^3 \pm 11.9 \mu\text{g}/\text{m}^3$) for the entire pregnancy period	Significantly higher effect of both PAH (OR = 1.5, (95% CI: 1.2 to 1.9) and $\text{PM}_{2.5}$ (OR = 1.5) exposure on the development of childhood bronchitis, associated with the low activity <i>EPHX1</i> (His/His) diplotype
Bijnens <i>et al.</i> (2015)	$n = 211$ twins of the East Flanders Prospective Twin Study (Belgium)	Doubling of the residential distance to a major road and doubling in traffic density, as proxy for maternal traffic/air pollution exposure	5.3% increase of RTL with every doubling of the residential distance to a major road (95% CI: 1.9 to 8.9%; $p=0.003$) and a decrease in RTL with 4.0% for every doubling in traffic density (95% CI: -7.6 to -0.2 , $p = 0.04$)
Clemente <i>et al.</i> (2016)	$n = 376$ (INMA cohort, Spain) and $n = 550$ (ENVIRONAGE cohort, Belgium)	10 $\mu\text{g}/\text{m}^3$ increase in NO_2 ($25.5 \pm 11.4 \mu\text{g}/\text{m}^3$ in INMA cohort and $21.1 \pm 4.2 \mu\text{g}/\text{m}^3$ in ENVIRONAGE cohort respectively for the entire pregnancy period)	1) ENVIRONAGE cohort: 11.1% decrease in mtDNA for the second trimester (95% CI: -19.9 to -1.24% ; $p = 0.03$) and 13.5% decrease in mtDNA for the third trimester of pregnancy (95% CI: -20.1 to -6.4% ; $p = 0.003$). 2) INMA cohort: decrease in mtDNA content for the first (-4.1% , 95% CI: -7.1 to -1.1% ; $p = 0.007$), second (-5.0% , 95% CI: -8.0 to -2.0% ; $p = 0.002$) and third (-4.9% , 95% CI: -7.9 to -1.8% ; $p = 0.003$) trimester, and for the entire pregnancy (-5.5% , 95% CI: -8.8 to -2.1% , $p = 0.002$)
Dodd-Butera <i>et al.</i> (2016)	$n = 54$ from Tijuana, Mexico	Work-related PAH air pollution exposure from working in maquiladora factories	No significant differences in DNA-adduct levels

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380 Abbreviations: 8-oxodG, (8-oxo-2'-deoxyguanosine); B[a]P, Benzo[a]pyrene; CI, Confidence interval; *EPHX1*, Epoxide hydrolase 1; *GSTM1*, Glutathione S-transferase M1; mtDNA, Mitochondrial
381 DNA; NAT2, N-acetyl transferase 2; NO_2 , Nitrogen dioxide; NO_x , Nitrogen oxides; OR, Odds ratio; PAH, Polycyclic aromatic hydrocarbon; $\text{PM}_{2.5}$, Particulate matter with a diameter smaller than
382 2.5 μm ; PM_{10} , Particulate matter with a diameter smaller than 10 μm ; RTL, Relative telomere length; SO_2 , Sulfur dioxide.

It is known that mtDNA fluctuates under the influence of age, ethnicity, and tissue investigated, but most importantly depends on oxidative stress level, cellular antioxidant capacity, type of environmental factor, and dose of exposure (Castegna *et al.* 2015; Meyer *et al.* 2013). Further research on this topic is essential, since alterations in placental mitochondrial function or capacity of the placenta may influence fetal energy provision and development (Mayeur *et al.* 2014).

Telomere length predicts life span early in life (Heidinger *et al.* 2012) and captures the history of inflammatory and oxidative stress effects of exposure to environmental stressors (Martens and Nawrot 2016). For example, exposure-related oxidative stress and inflammation are known to contribute to telomere shortening (Zhang *et al.* 2013). Bijmens *et al.* (2015) investigated changes in placental telomere length in twins in correlation with traffic-related exposure. In this study, three indicators of exposure were assessed, i.e. the distance from the residential address of the mother to the nearest major road, traffic density within a 200 m buffer from the residence, and residential greenness. The authors concluded that placental telomere length was longer in association with a doubling of the residential distance to a major road (5.3%, 95% CI: 1.9 to 8.9%; $p = 0.003$), and shorter with a doubling in traffic density (-4.0%, 95% CI: -7.6 to -0.2%; $p = 0.04$).

Other genomic factors of susceptibility in the context of health and disease are DNA polymorphisms. Specific polymorphisms can cause an alteration in the metabolic capacity of cells as to the degradation and/or elimination of toxic substances, such as particle-bound chemicals derived from tobacco smoke or ambient air pollution. In turn, these metabolic changes could entail a new risk of disease development (Bartsch *et al.* 2000). Research has focused specifically on the associations between *in utero* exposure to air pollution and placental genotypes related to detoxification mechanisms. One of the most important actors in this process is cytochrome P450 1A1 (*CYP1A1*), which is expressed in various tissue types throughout the body including the placenta, and fulfils both a detoxifying and a bioactivating role. This enzyme can bioactivate pro-carcinogenic substances such as PAHs to form adducts with DNA in tissues of both the mother and child during pregnancy (Stejskalova and Pavek 2011). Whyatt *et al.* (1998) focused on *in utero* air pollution exposure and its effects on

changes of placental *CYP1A1*. In the genomic category, the authors investigated the association between the homozygous (*MspI*^{+/+}) or heterozygous (*MspI*^{+/-}) presence of the *CYP1A1* *MspI* polymorphism in placental tissue between smokers and non-smokers within areas heavily or less polluted with PAHs. An association between the placental presence of this polymorphism and the formation of DNA adducts due to PAH air pollution could not be demonstrated (Whyatt *et al.* 1998). In addition, Sram *et al.* (2006) studied the association between *CYP1A1* polymorphisms and PAH levels in association with birth weight, but a significant effect of these placental polymorphisms on birth weight following maternal PAH exposure was not found. PM_{2.5} and PM₁₀ levels were also measured in this study, but no effects of these air pollution components were mentioned. Glutathione S-transferase M1 (*GSTM1*) and N-acetyl transferase 2 (*NAT2*) are two other enzymes involved in the detoxification system of cells. Studies conducted on the genotypes of these two enzymes in human placental tissue showed that both genes interact with ROS, but only the null phenotype *GSTM1*^{-/-}, unlike *GSTM1*^{+/-} or *GSTM1*^{+/+}, was correlated with maternal exposure to SO₂, NO_x and PM₁₀ during pregnancy (Topinka *et al.* 1997a, 1997b). The placenta did not only prove to be a useful tissue for genomic analyses of *GST* polymorphisms in connection with air pollution exposure, but also for studying the proteomic level of placental GST activity (Obolenskaya *et al.* 2010) (see proteomics section).

4.2.2. Epigenetics (Table 3)

The most commonly characterized epigenetic marking process is DNA methylation, which involves the addition of a methyl group to the carbon-5 position of cytosine residues of the dinucleotide CpG. DNA methylation undergoes critical modification during early *in utero* life. After fertilization and prior to implantation, DNA methylation patterns are largely erased but are re-established by *de novo* DNA methyltransferases (DNMTs) in the blastocyst stage (Jirtle and Skinner 2007). These waves of epigenetic reprogramming likely make early embryonic development a critical period during which nutritional, environmental, and metabolic factors affect the developmental establishment of epigenetic regulation (Kelsey and Michels 2012). The placenta exhibits a different methylation profile compared to fetal somatic tissue (Magda Price *et al.* 2012) which is probably

needed to generate cells with a broad developmental potential and the correct initiation of embryonic gene expression. Indeed, the placenta shows considerable developmental plasticity which is important for adaptation to fetal and maternal signals including hormonal and environmental exposures or other responses to *in utero* conditions (Hogg *et al.* 2012). Hence, the placenta contains information on DNA methylation patterns revealing the environmental impact to which the fetus has been exposed during gestation.

An expanding body of evidence suggests that exposures to hazardous environmental factors are important determinants for altered DNA methylation-related programming during early life. These alterations can persist throughout the course of life, thereby leading to pathological conditions in adulthood. Recently, Vaiserman (2015) summarized clinical and epidemiological evidence in support of epigenetic factors that may mediate the link between early-life exposures and long-term health outcomes. Changes in DNA methylation patterns of placental tissue have been disclosed in association with adverse maternal exposures such as alcohol and tobacco smoke (Wilhelm-Benartzi *et al.* 2012), however only recently placental epigenetic signatures have been identified in association with exposure to ambient air pollution. Janssen *et al.* (2013) were the first to investigate the association between PM_{2.5} exposure during pregnancy and the global DNA methylation levels in placental tissue. For the entire pregnancy period they found that an increase of 5 µg/m³ in PM_{2.5} exposure correlated with a relative decrease of 2.2% in global placental DNA methylation (95% CI: -3.7 to -0.7%; p = 0.004). These findings have been replicated by Kingsley *et al.* (2016). The authors showed that pregnant mothers living closer to major roads, as a marker of traffic-related air pollution, had lower levels of placental DNA methylation in LINE-1 (-0.82%, 95% CI: -1.57 to -0.07; p = 0.03) but not AluYb8 repetitive elements, which can be regarded as surrogate markers of global DNA methylation.

457 **Table 3.** Studies describing the associations between prenatal ambient air pollution exposure and changes in placental epigenetic markers

Author	Study population	Increase in analyzed air pollution component (average \pm standard deviation if available)	Effect on placental -omics marker
Janssen <i>et al.</i> (2013)	240 samples from the ENVIRONAGE birth cohort (Belgium)	5 $\mu\text{g}/\text{m}^3$ PM _{2.5} increase (17.4 \pm 3.6 $\mu\text{g}/\text{m}^3$ for entire pregnancy)	Decrease in global DNA methylation for whole pregnancy (-2.2%, 95% CI: -3.7 to -0.7%; p = 0.004), first trimester (-2.4%, 95% CI: -3.6 to -1.2%; p = 0.0001) and second trimester of pregnancy (-1.5%, 95% CI: -2.7 to -0.4%; p = 0.01)
Janssen <i>et al.</i> (2015)	381 mother-newborn pairs from the ENVIRONAGE birth cohort (Belgium)	3 $\mu\text{g}/\text{m}^3$ (IQR) increase in PM _{2.5} (16.7 \pm 2.3 $\mu\text{g}/\text{m}^3$)	Increased mtDNA methylation levels (0.5%, 95 % CI: 0.2 to 2.2%; p < 0.05) and decrease of mtDNA content with 15.6% (95% CI: -23.9 to -6.4%; p < 0.05)
Kingsley <i>et al.</i> (2016)	471 mother-infant pairs from the RICHS cohort (Rhode Island, USA)	Proximity of the residential distance to a major road as proxy for air pollution exposure	0.82% decrease in mean LINE-1 methylation levels (95% CI: -1.57 to -0.07; p = 0.03)
Tsamou <i>et al.</i> (2016)	210 mother-child pairs from the ENVIRONAGE cohort (Belgium)	5 $\mu\text{g}/\text{m}^3$ increase in PM _{2.5} (16.38 \pm 5.29 $\mu\text{g}/\text{m}^3$ for the first trimester of pregnancy and 16.74 \pm 5.82 $\mu\text{g}/\text{m}^3$ for the second trimester of pregnancy)	Decreased expression of miR-21 (-33.7%, 95% CI: -53.2 to -6.2%; p = 0.02), miR-146a (-30.9%, 95% CI: -48.0 to -8.1%; p = 0.012) and miR-222 (-25.4%, 95% CI: -43.0 to -2.4%; p = 0.034) for the second trimester of pregnancy and an increased expression of miR-20a (+70.9%, 95% CI: 16.7 to 150.3%; p = 0.007) and miR-21 (+73.7%, 95% CI: 11.7 to 170.1%; p = 0.015) in the first trimester.
Saenen <i>et al.</i> (2017)	361 samples from the ENVIRONAGE birth cohort (Belgium)	7.5 $\mu\text{g}/\text{m}^3$ (IQR) increase in PM _{2.5} (15.5 \pm 4.9 $\mu\text{g}/\text{m}^3$ for the second trimester)	A 1.4% decrease in <i>LEP</i> promoter methylation for the second trimester of pregnancy (95% CI: -2.7 to -0.2%; p = 0.02)

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459 Abbreviations: CI, Confidence interval; IQR, Interquartile range; *LEP*, Leptin; miR, Micro RNA; mtDNA, Mitochondrial DNA; PM_{2.5}, Particulate matter with a diameter smaller than 2.5 μm .

Another interesting finding of the study of Janssen *et al.* (2013) was that the early gestational stage from fertilization up to and including implantation - a critical period for methylation reprogramming - is likely to represent a highly sensitive window for the effects of PM_{2.5} exposure on placental DNA methylation as measured at birth. The health implications of these findings should be further investigated, since it has been shown that overall hypomethylation patterns in the placenta could be an indication of an increased risk to birth defects such as spina bifida (Zhang *et al.* 2015). Furthermore associations have been found between hypomethylation of specific promoters and adverse birth conditions such as low birth weight (Rumbajan *et al.* 2016).

Recently, attention has been drawn to the methylation pattern of a specific gene in the placenta, namely the promoter region of the leptin (*LEP*) gene. Leptin is an important hormone during pregnancy, since it plays a crucial role in fetal growth and development through its function in energy metabolism (Walsh *et al.* 2014). An interquartile range increment (IQR) of PM_{2.5} exposure (7.5 µg/m³) was associated with a 1.4% decrease in placental methylation of the *LEP* promoter region (95 % CI: -2.7 to -0.2%; p = 0.02) (Saenen *et al.* 2017). In previous research, a decrease in *LEP* methylation has been associated with gestational syndromes such as pre-eclampsia (Hogg *et al.* 2013) and impaired glucose tolerance (Bouchard *et al.* 2010). The intricate connection between *LEP* methylation, PM exposure, and disease phenotype should be explored more in depth by studying potential ailments in childhood that may arise from these placental changes.

Besides the nuclear genome, the mitochondrial genome can undergo epigenetic modifications as well. For example, maternal emotional stress during pregnancy has shown to alter gene expression patterns in placental mitochondria, which can eventually affect the temperamental development of the child in early life (Lambertini *et al.* 2015). DNA methylation in specific regions of the mitochondrial genome has been shown to substantially mediate the association between PM_{2.5} exposure during gestation and placental mtDNA content which could reflect signs of mitophagy and mitochondrial death (Janssen *et al.* 2015). However, the epigenetic changes in mtDNA patterns linked to air pollution exposure have not yet been studied in the context of developmental outcomes of the newborn.

Therefore, further exploration of mitochondrial gene expression regulation by DNA methylation is of paramount importance to unravel these potentially important relationships.

A type of epigenetic mark that has not yet been investigated to a great extent in the context of prenatal air pollution exposure is microRNA (miRNA) (Vrijens *et al.* 2015). MiRNAs are endogenous, single-stranded, short non-coding RNA sequences (approximately 22 nucleotides) that regulate gene expression at the post-transcriptional level. Different cell types have both common and unique miRNA expression patterns, which can be influenced by developmental and pathologic states. The human placenta expresses a distinct subset of miRNAs, but although the functions of these placental epigenetic marks are largely unknown, recent research has revealed a functional role for miRNAs in placental biology (Gu *et al.* 2013). The presence of placental miRNAs in the maternal circulation is interesting as it could lead to the discovery of biomarkers of placental dysfunction or pregnancy-related disease (Miura *et al.* 2016). Only one study has described changes in placental miRNA expression in association with prenatal air pollution exposure. A relative decrease in the placental expression of miR-21 (-33.7%, 95% CI: -53.2 to -6.2%; $p = 0.022$), miR-146a (-30.9%, 95% CI: -48.0 to -8.1%; $p = 0.012$), and miR-222 (-25.4%, 95% CI: -43.0 to -2.4%; $p = 0.034$) was found in association with an increase of $5 \mu\text{g}/\text{m}^3$ $\text{PM}_{2.5}$ during the second trimester of pregnancy, whereas a positive association was described between first trimester $\text{PM}_{2.5}$ air pollution exposure and the expression of placental miR-20a (+70.9%, 95% CI: 16.7 to 150.3%; $p = 0.007$) and miR-21 (+73.7%, 95% CI: 11.7 to 170.1%; $p = 0.015$) (Tsamou *et al.* 2016). A common target of these miRNAs is the tumor suppressor phosphatase and tensin homolog (*PTEN*) which also showed an altered expression in association with $\text{PM}_{2.5}$ exposure (+59.6% per $5 \mu\text{g}/\text{m}^3$ increment, 95% CI: 26.9 to 100.7%; $p < 0.0001$) and, as expected, an inverse correlation with the levels of these miRNAs, since increasing levels of miRNAs are known to block the expression of their associated targets (Tsamou *et al.* 2016).

4.2.3. Transcriptomics (Table 4)

The area of placental transcriptomics has been poorly addressed in association with exposure to air pollution during pregnancy. One possible explanation for this is the difficulty to obtain placental

tissue aliquots with a sufficiently high RNA quality for whole transcriptome analyses, since RNA starts degrading quickly after sampling (Gallego Romero *et al.* 2014). However, various reviews have discussed the transcriptome of the placenta in the context of negative birth outcomes (Cox *et al.* 2015; Eidem *et al.* 2015). Recent research has shown that the placenta contains several distinct gene expression patterns compared with other tissues in the human body, for example when the number of splice variants and the expression levels of regulators involved in splicing are concerned (Kim *et al.* 2012). Quantitative PCR, micro-array analysis, and RNA sequencing have proven to be indispensable for the analysis of exposure effects on transcriptomic alterations, potentially leading to perturbation of developmental and biological mechanisms. Apart from their work in the genomic field, Whyatt *et al.* (1995, 1998) also investigated the effects of prenatal exposure to air pollution on the placental expression of *CYP1A1*. However, in both studies significant results concerning this gene could not be established, which was in accordance with the absence of an association in the genomic field of their research on *CYP1A1* (see section 4.2.1. Genomics).

Saenen *et al.* (2015) investigated the placental expression levels of ten genes in the brain-derived neurotrophic factor (*BDNF*) pathway in connection with PM air pollution. A 5 µg/m³ increase in residential PM_{2.5} exposure of the mother during the first trimester of pregnancy was associated with a 15.9% decrease in expression of placental *BDNF* (95% CI: -28.7 to -3.2%, p = 0.015), and with a 24.3% decrease in *synapsin 1* (*SYN1*) expression (95% CI: -42.8 to -5.8%, p = 0.011) which is affected by the actions of *BDNF*. Proper functioning of this pathway in the placenta is crucial for normal fetal development, since altered *BDNF* expression in this tissue has been associated with negative birth outcomes such as fetal growth restriction (Mayeur *et al.* 2010).

533 **Table 4.** Studies describing the associations between prenatal ambient air pollution exposure and changes in placental transcriptomic markers

Author	Study population	Increase in analyzed air pollution component (average \pm standard deviation if available)	Effect on placental -omics marker
Whyatt <i>et al.</i> (1995)	70 subjects from a city with higher levels of air pollution (Krakow) and 90 subjects from a less polluted city (Limanowa) in Poland	Average of 80 $\mu\text{g}/\text{m}^3$ PM_{10} in highly polluted area (range, 23.4-154.2 $\mu\text{g}/\text{m}^3$), no data available for Limanowa	No significant difference in <i>CYP1A1</i> mRNA levels between low and high polluted area ($r = -0.4$; $p = 0.14$).
Whyatt <i>et al.</i> (1998)	70 subjects from Krakow with higher levels of air pollution and 90 subjects from Limanowa, a less polluted city in Poland	Average annual concentration of 37 $\mu\text{g}/\text{m}^3$ of ambient respirable particles in least exposed group and 78 $\mu\text{g}/\text{m}^3$ in the most exposed group, in the year prior to delivery (particle size not defined)	PAH-adduct levels not significantly associated with <i>CYP1A1</i> mRNA ($r = -0.10$; $p = 0.2$)
Saenen <i>et al.</i> (2015)	90 randomly selected mother-child pairs from the ENVIRONAGE birth cohort (Belgium)	5 $\mu\text{g}/\text{m}^3$ increase in $\text{PM}_{2.5}$ (15.4 \pm 5.4 $\mu\text{g}/\text{m}^3$ for the first trimester, 17.6 \pm 7.0 $\mu\text{g}/\text{m}^3$ for the second trimester and 18.7 \pm 6.0 $\mu\text{g}/\text{m}^3$ for the third trimester of pregnancy)	15.9% decrease in placental <i>BDNF</i> expression (95% CI: -28.7 to -3.2%; $p = 0.015$) and a 24.3% lower expression of <i>SYN1</i> (95% CI: -42.8 to -5.8%, $p = 0.011$)

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535 Abbreviations: *BDNF*, Brain-derived neurotrophic factor; CI, Confidence interval; *CYP1A1*, Cytochrome (*CYP*) P450 1A1; PAH, Polycyclic aromatic hydrocarbon; $\text{PM}_{2.5}$, Particulate matter with a
536 diameter smaller than 2.5 μm ; PM_{10} , Particulate matter with a diameter smaller than 10 μm , *SYN1*, Synapsin 1.

4.2.4. Proteomics (Table 5)

Although proteins are the end products of the transcription of genomic sequences, there is no linear relationship between a genome and its resulting proteome because of alternative splicing and the production of non-functional proteins (Pandey and Mann 2000). Therefore, it is essential that proteomics signatures are studied as a separate -omics field which can complement the finding in other -omics categories (Pandey and Mann 2000). Proteomic characteristics have been investigated in placental tissue in association with maternal tobacco smoking habits (Machaalani *et al.* 2014) and other exposures to toxicants during pregnancy. Considerable focus has been put on proteins that play an essential role in the detoxification system of the cells. The activity of placental aryl hydrocarbon hydroxylase (AHH), which is the most important metabolizer of PAHs, was significantly higher in placentas obtained from mothers who lived in an environment exposed to urban air pollution compared with the control group (Hincal 1986). Two other cellular detoxification indicators, glutathione S-transferase (GST) and 7-ethoxycoumarin O-deethylase (ECOD), showed opposite associations with increasing air pollution levels: ECOD activity significantly increased with increasing ambient air pollution (related to industry and traffic-exhaust), while the GST activity decreased under the same conditions (Obolenskaya *et al.* 2010). Work-related exposure to another source of PAH air pollution during pregnancy, such as in the “maquiladoras” at the US-Mexican border, had no effect on the placental GST level or activity (Dodd-Butera *et al.* 2016). Detoxification processes are important for normal cellular functioning, but the maintenance of the delicate redox balance of the cell is a crucial factor as well. In controlling oxidative stress, metallothionein (MT) is an important protein for fixation and transport of metals. In a study of Sorkun *et al.* (2007), a significant increase in the amount of placental MT was observed in regions with higher levels of air pollution exposure. Another area of interest in placental proteomics addresses the energy system of the cell.

560 **Table 5.** Studies describing the associations between prenatal ambient air pollution exposure and changes in placental proteomic markers

Author	Study population	Increase in analyzed air pollution component (average \pm standard deviation if available)	Effect on placental -omics marker
Hincal (1986)	152 mother-child pairs from residential Ankara and 125 mother-child pairs from the more rural areas surrounding Ankara	Urban air pollution	AHH activity was significantly higher in placental tissue of women living in Ankara compared to women of the rural areas ($p < 0.001$)
Kedryna <i>et al.</i> (2004)	15 women from Chorzow and Krakow (polluted areas) and 8 women from the Bieszczady Mountains (less polluted area) in Poland	Urban air pollution	Significant decrease in pyruvate kinase activity in more polluted areas ($p < 0.001$)
Sorkun <i>et al.</i> (2007)	Samples of 92 mothers: 33 smokers, 29 exposed to air pollution and 30 non-smokers residential in a rural area with lower levels of air pollution	Urban air pollution	Higher levels of metallothionein in group exposed to air pollution, compared to mothers living in rural area ($p = 0.013$)
Obolenskaya <i>et al.</i> (2010)	143 mothers who gave birth between 1991-1999 in polluted areas of Ukraine and Belarus, and a less polluted area in the east of Poland.	Urban air pollution	Significantly lower GST activity ($r_s = -0.27$; $p = 0.05$) and higher ECOD activity ($p < 0.05$) in highly polluted areas compared to the lower polluted areas
Saenen <i>et al.</i> (2016)	330 mother-child pairs from the ENVIRONAGE birth cohort (Belgium)	3.5 $\mu\text{g}/\text{m}^3$ (IQR) increase in $\text{PM}_{2.5}$ ($16.1 \pm 2.4 \mu\text{g}/\text{m}^3$ for whole pregnancy) and 0.36 $\mu\text{g}/\text{m}^3$ (IQR) increase in BC ($0.97 \pm 0.28 \mu\text{g}/\text{m}^3$ for entire pregnancy)	35.0% increase in 3-NTp levels for increased $\text{PM}_{2.5}$ levels (95% CI: 13.9 to 60.0%; $p = 0.0006$) and 13.9% increase in 3-NTp levels for increased BC levels (95% CI: -0.2 to 29.9%; $p = 0.05$) during the entire pregnancy period
Dodd-Butera <i>et al.</i> (2016)	$n = 54$ from Tijuana, Mexico	Work-related PAH air pollution exposure from working in maquiladora factories	No significant difference in GST level ($p = 0.243$) or GST activity ($p = 0.965$) between women working in maquiladoras and women from non-exposed area

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562 Abbreviations: 3-NTp, 3-nitrotyrosine; AHH, Aryl hydrocarbon hydroxylase; BC, Black carbon; CI, Confidence interval; *CYP1A1*(*2A), Cytochrome (*CYP*) P450 1A1 (2A); ECOD, 7-ethoxycoumarin
563 O-deethylase; *EPHX1*, Epoxide hydrolase 1; GST, Glutathione S-transferase; IQR, Interquartile range; PAH, Polycyclic aromatic hydrocarbon; $\text{PM}_{2.5}$, Particulate matter with a diameter smaller
564 than 2.5 μm .

Pyruvate kinase, an essential enzyme in the process of glycolysis, showed a significant increase of activity in placental tissue of women who lived in more polluted areas (Kedryna *et al.* 2004). The level of this protein was also increased in placental tissue of preeclampsia pregnancies (Bahr *et al.* 2014), a condition characterized by excessive inflammatory reactions in the placenta. This parallels prenatal exposure to air pollution, as inflammation is the most likely mode of action triggered by ambient air pollution (Janssen *et al.* 2012). Because glycolysis is a very important metabolic pathway of energy production in the placenta (Bloxxam 1985), more attention is needed as to the effects of air pollution on this critical pathway during gestation.

Not only the intact proteins themselves, but also the products of protein degradation or modification can be measured as biomarkers of placental damage caused by detrimental influences during pregnancy. Protein damage can be caused by processes such as oxidative stress and inflammation. In this context, the tyrosine groups of proteins can be modified into 3-nitrotyrosine (3-NTp) by peroxynitrite, which is an intermediate of oxidative or nitrosative stress. Recently, a positive association was found between placental 3-NTp levels and both PM_{2.5} exposure (+35.0% for a 3.5 µg/cm³ increment in PM_{2.5}, 95% CI: 13.9 to 60.0%; *p* < 0.0006) and BC exposure (+13.9% for a 0.36 µg/m³ increment in BC, 95% CI: -0.2 to 29.9%; *p* = 0.05) (Saenen *et al.* 2016), which is in line with recent studies on mice. These animals showed increased placental 3-NTp levels which correlated with exposure to air pollution-related diesel exhaust (Weldy *et al.* 2014).

4.2.5. Metabolomics

Metabolomics is a research area that deals with changes in small metabolites (lipids, amino acids, sugars, etc.) as a consequence of altered metabolism by internal or external influences (Tzoulaki *et al.* 2014). This -omics study area has been broadly addressed within several research topics concerning placental tissue. The placental metabolome of complicated pregnancies has been compared with that of normal pregnancies to investigate the molecules associated with adverse outcomes such as neural tube defects (Chi *et al.* 2014). However, the placental metabolome characteristic of prenatal exposure to ambient air pollution did not deserve any attention until now.

Metabolomic parameters linked to the effects of air pollution have been studied to a small extent in other human matrices such as umbilical cord blood plasma (e.g. oxylipins) (Martens *et al.* 2017) and lung lavage fluid (Surowiec *et al.* 2016). Since these results showed specific metabolic signatures associated with air pollution exposure, this should be an incentive to further investigate tissues such as the placenta to reveal early-life changes in metabolic pathways due to adverse exposures during pregnancy.

4.3. Triple relationship between exposure to ambient air pollution, placental biomarkers and disease development

As stated by Professor David Barker in the early 1990s, the occurrence of diseases later in life may already be initiated during fetal development as a result of detrimental *in utero* exposures and direct or indirect influences of placental involvement (Barker 1995). To fully comprehend the complexity of the fetal origin of disease, it is crucial to investigate the intricate triple relationship between exposure, molecular effect and clinical outcome (Table 6). In earlier research, morphological changes in placental tissue have been linked to chorangiosis, an adverse condition of the placenta itself, which is an indirect consequence for disease development and known to be associated with perinatal mortality and morbidity. Maternal exposure to urban ambient air pollution during the gestational period has been shown to lead to a significantly higher number of chorionic villi without a change in placental weight suggesting an increased possibility for developing chorangiosis (Akbulut *et al.* 2009). The associations between prenatal exposure to air pollution, the molecular changes in the placenta, and the consequences on developmental or disease characteristics later in life have not yet been studied extensively. Ghosh *et al.* (2013) investigated the effect of maternal gestational exposure to air pollution in relation to a specific placental genotype and the development of childhood bronchitis during the first two years of life.

614 **Table 6.** Studies describing the triple relationship between exposure to ambient air pollution during pregnancy, the associated placental -omics marker and the health outcome.

Author	Exposure measured in ambient air	Placental measurement	Disease / health condition
Hincal F. (1986)	Urban air pollution	Placental AHH activity	Low birth weight and shorter birth length
Sram <i>et al.</i> (2006)	PM _{2.5} , PM ₁₀	DNA adducts and <i>GSTM1</i> , <i>GSTP1</i> , <i>GSTT1</i> , <i>CYP1A1</i> *2A and <i>CYP1A1</i> *2C genotypes	Low birth weight and prematurity
Rossner <i>et al.</i> (2011)	PAHs, PM _{2.5}	SNP analysis for 95 genes and measurement of 8-oxodG adducts	Low birth weight and intrauterine growth restriction
Ghosh <i>et al.</i> (2013)	PAH, PM _{2.5}	Six SNPs (<i>GSTM1</i> , <i>GSTP1</i> , <i>GSTT1</i> , <i>CYP1A1 MspI</i> , <i>EPHX1</i> exon 3 and 4) and one <i>EPHX1</i> diplotype	Acute bronchitis in early childhood
Clemente <i>et al.</i> (2016)	NO ₂	mtDNA content	Low birth weight
Kingsley <i>et al.</i> (2016)	Residential proximity to a major road	DNA methylation (LINE and AluYb8 elements)	Low birth weight and small for gestational age

615

616 Abbreviations: 8-oxodG, (8-oxo-2'-deoxyguanosine); AHH, Aryl hydrocarbon hydroxylase; *CYP1A1*(*2A), Cytochrome (*CYP*) P450 1A1 (2A); *EPHX1*, Epoxide hydrolase 1; *GSTM1*, Glutathione S-
617 transferase M1; *GSTP1*, Glutathione S-transferase P1; *GSTT1*, Glutathione S-transferase T1; His, Histidine; *LEP*, Leptin; miR, MicroRNA; mtDNA, Mitochondrial DNA; *MspI*, Substitution of
618 isoleucine to valine in the 3' non-coding region of *CYP1A1*, NO₂, Nitrogen dioxide; PAH, Polycyclic aromatic hydrocarbon; PM_{2.5}, Particulate matter with a diameter smaller than 2.5 µm; PM₁₀,
619 Particulate matter with a diameter smaller than 10 µm; SNP, Single nucleotide polymorphism.

A significant relationship was identified between the development of childhood bronchitis and the presence of a low activity *EPHX1* polymorphism in the placenta with increased exposure to PAH and PM_{2.5}. These authors were thereby the first to identify a link between prenatal exposure to air pollution, a placental -omics marker and disease development.

Several studies have focused on placental -omics signatures and adverse birth outcomes such as reduced birth weight, intrauterine growth restriction, small for gestational age, and prematurity. In earlier studies, no significant associations were found between ambient air pollution exposure, birth weight and either the levels of DNA adducts in the placenta (Sram *et al.* 2006) or the activity of aryl hydrocarbon hydroxylase in placenta (AHH) (Hincal 1986), although a link was shown between air pollution exposure and AHH activity (see proteomics section). Three more recent studies found significant negative correlations between birth weight and the levels of NO₂ (Clemente *et al.* 2016), PM_{2.5} and PAHs (Rossner *et al.* 2011), and the residential proximity to a major road (Kingsley *et al.* 2016). However, Rossner *et al.* (2011) concluded that the levels of 8-oxo-deoxyguanosine (8-oxodG) DNA adducts, a marker of direct oxidative DNA damage, measured in placental tissue were only correlated with PM_{2.5} levels but not with birth weight. The same conclusion was reached for the association between residential distance to a major road and LINE element methylation: LINE methylation levels were only negatively associated with a distance to a major road, but no link with birth weight could be identified (Kingsley *et al.* 2016). The only mediation analysis to investigate the triple association between air pollution exposure, placental -omics and disease development was performed with data of the INMA birth cohort (Clemente *et al.* 2016). This analysis showed that 10% of the association between a 10 µg/m³ increase in NO₂ exposure during pregnancy and reduced birth weight could be mediated by a decrease in mitochondrial DNA levels. The results of these three studies could suggest that prenatal exposure to air pollution might exert its effects on birth outcomes by altering more subtle regulations such as those of the placental energy system and not by direct damage to the placental DNA. However, since a broad array of possible molecular factors could be responsible for the link between *in utero* air pollution exposure and an effect on disease development later in life,

integration of results on all -omics fields and the potential associations with prenatal exposure to ambient air pollution and childhood development should be prioritized in future research. This could eventually aid in the understanding of the complex etiology of adult diseases.

The strength of this review is that this is, to our knowledge, the first descriptive work to summarize and discuss the current knowledge on all placental -omics signatures that have been analyzed in association with prenatal air pollution exposure. This article has identified the current areas with the greatest gaps of knowledge which need to be addressed in future research and can therefore be a base to guide placental -omics research. A limitation in this review is the heterogeneity of the study designs of the 25 discussed articles. Differences were apparent in the approaches used to obtain the -omics data, the placental sampling protocols, and in both exposure assessment as well as the definition of the specific exposure windows. For these reasons a formal meta-analysis to combine the study results was not possible.

5. CONCLUSIONS AND FUTURE DIRECTIONS

Exposure to air pollution in daily life is unavoidable. A crucial time window of exposure in the course of human life is fetal development. The feto-placental unit is subjected to maternal conditions and exposures that can adversely affect -omics characteristics of the placenta. Eventually, these placental changes could potentially lead to alterations in metabolic capacities of the fetus and an increased risk of disease development later in life. This systematic review shows that the placenta is a suitable tissue to investigate the effects of prenatal exposure to ambient air pollution by examining -omics biomarkers. The placenta is a temporary organ that reflects various exposures throughout pregnancy. Important in this branch of research is to have a representation of the effects of these exposures on the whole placenta. Since not only inter-, but also intra-placental differences should always be taken into account, researchers should try to find a consensus on a unified, standardized method to work

with a pooled sample of each placenta. Especially in -omics research, placental sampling is crucial because of the fragility of DNA, protein and especially RNA structures. Therefore, standardization and communication about sampling methods is crucial in -omics research. In this way, results over different cohorts could be more easily compared and discussed.

At this point in time, this systematic review shows that some -omics fields are more represented than others in the research on the effects of prenatal exposure to air pollution on placental biomarkers. The most focus has been put on the presence of placental DNA adducts, although only a minority of studies found significant effects of air pollution exposure on these biomarkers. Therefore, more attention should be put on other, more promising -omics fields such as epigenetics, transcriptomics, and proteomics. At present, several placental -omics markers have been suggested that could provide a better insight on how the consequences of exposure to ambient air pollution are manifested during pregnancy. However, most studies only focus on specific components of molecular systems and pathways. Integrating a top-down approach is crucial for epigenomics, transcriptomics and proteomics for a full understanding of the array of molecular changes that result from detrimental environmental exposures. More studies containing large qualitative datasets should combine candidate -omics markers with the exploration of entire metabolic pathways in the full genome, transcriptome, epigenome, proteome or metabolome. Eventually, this should provide a complete molecular signature of key players describing the effects of prenatal environmental exposure on placental functioning and fetal (disease) development.

Two -omics fields, - metabolomics and exposomics -, could not be sufficiently covered in the context of this systematic review because of the current paucity of such studies. More attention should be put on these fields to further expand the knowledge on placental biomolecular signatures of prenatal air pollution exposure. In general, the effects of detrimental exposures on placental molecular changes and the subsequent effects on the programming of pathologies later in life are rather scarcely documented. Therefore, future research should focus more on integrative projects such as the

epigenome-wide association studies (EWAS) to identify key molecular regulators in the etiology of disease processes. Also, more longitudinal follow-up research is needed to identify and clarify the triple link between *in utero* exposure to ambient air pollution, changes in placental -omics categories, and disease initiation/progression later in life. Two projects that are already integrating several hazardous exposures such as ambient air pollution, the molecular signatures of these exposures in several tissues and the health effects on newborns, children and adults are the Human Early-Life Exposome (HELIX) project (Vrijheid *et al.* 2014) and the EXPOsOMICS project (Vineis *et al.* 2016). In conclusion, future integrative long-term research looks promising in elucidating the underlying placental mechanisms that potentially influence disease development later in life, as a consequence of gestational air pollution exposure.

6. DECLARATIONS

6.1. COMPETING FINANCIAL INTERESTS DECLARATION

All authors declare they do not have any competing financial interests.

6.2. FUNDING

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6.4. AUTHORS' CONTRIBUTIONS

Selection criteria for the in- or exclusion of articles were determined by LJL, NDS, and TSN. The literature search was performed by LJL and NDS, and TSN was consulted if any discrepancies remained between these two researchers. LJL wrote the first draft of the systematic review, with contributions from NDS (suggestions on section "4.1. Placental tissue in epidemiological research: advantages and disadvantages"), BGJ (section "4.2.2. Epigenetics" on (mt)DNA methylation) and KV (section "4.2.2. Epigenetics" on miRNA). HAR put special attention to the construction and finalization of the manuscript. All authors read and discussed the final version of the manuscript and approved it before submission.

7. REFERENCES

- Akbulut M, Sorkun HC, Bir F, Eralp A, Duzcan E. 2009. Chorangiosis: The potential role of smoking and air pollution. *Pathol. Res. Pract.* 205:75–81; doi:10.1016/j.prp.2008.05.004.
- Bahr BL, Price MD, Merrill D, Mejia C, Call L, Bearss D, et al. 2014. Different expression of placental pyruvate kinase in normal, preeclamptic and intrauterine growth restriction pregnancies. *Placenta* 35:883–890; doi:10.1016/j.placenta.2014.09.005.
- Ballester F, Estarlich M, Iñiguez C, Llop S, Ramón R, Esplugues A, et al. 2010. Air pollution exposure during pregnancy and reduced birth size: a prospective birth cohort study in Valencia, Spain. *Environ. Health* 9:6; doi:10.1186/1476-069X-9-6.
- Barker DJ. 1995. Fetal origins of coronary heart disease. *BMJ* 311:171–4; doi:10.1136/bmj.311.6998.171.
- Bartsch H, Nair U, Risch A, Rojas M, Wikman H, Alexandrov K. 2000. Genetic polymorphism of CYP genes, alone or in combination, as a risk modifier of tobacco-related cancers. *Cancer Epidemiol. Biomarkers Prev.* 9: 3–28.
- Bates S, Clapton J, Coren E. 2007. Systematic maps to support the evidence base in social care. 3: 539–551.
- Bijnens E, Zeegers MP, Gielen M, Kicinski M, Hageman GJ, Pachen D, et al. 2015. Lower placental telomere length may be attributed to maternal residential traffic exposure; a twin study. *Environ. Int.* 79:1–7; doi:10.1016/j.envint.2015.02.008.
- Bloxam D. 1985. Human placental energy metabolism: its relevance to in vitro perfusion. *Contrib Gynecol Obs.* 13: 59–69.
- Bonnin A, Goeden N, Chen K, Wilson ML, King J, Shih C, et al. 2011. A transient placental source of serotonin for the fetal forebrain. *Nature* 472:347–350; doi:10.1038/nature09972.A.
- Bouchard L, Thibault S, Guay S-P, Santure M, Monpetit A, St-Pierre J, et al. 2010. Leptin gene

754 epigenetic adaptation to impaired glucose metabolism during pregnancy. *Diabetes Care* 33:
755 2436–2441.

756 Burton GJ, Fowden AL, Thornburg KL. 2016. Placental origins of chronic disease. *Physiol. Rev.*
757 96:1509–1565; doi:10.1152/physrev.00029.2015.

758 Burton GJ, Sebire NJ, Myatt L, Tannetta D, Wang Y, Sadovsky Y, et al. 2014. Optimising sample
759 collection for placental research. *Placenta* 35:9–22; doi:10.1016/j.placenta.2013.11.005.

760 Buss C, Entringer S, Wadhwa PD. 2013. Fetal programming of brain development: intrauterine stress
761 and susceptibility to psychopathology. *Sci Signal* 5; doi:10.1126/scisignal.2003406.Fetal.

762 Castegna A, Iacobazzi V, Infantino V. 2015. The mitochondrial side of epigenetics. *Physiol. Genomics*
763 47:299–307; doi:10.1152/physiolgenomics.00096.2014.

764 Cha D-H, Kim GJ. 2010. Dynamic role of trophoblast in human placental development. In *Human*
765 *Placenta: Structure and Development, Circulation and Functions*, pp. 49–77.

766 Chi Y, Pei L, Chen G, Song X, Zhao A, Chen T, et al. 2014. Metabonomic profiling of human placentas
767 reveals different metabolic patterns among subtypes of neural tube defects. *J. Proteome Res.*
768 13:934–945; doi:10.1021/pr4009805.

769 Clemente DBP, Casas M, Vilahur N, Begiristain H, Bustamante M, Carsin A, et al. 2016. Prenatal
770 ambient air pollution, placental mitochondrial DNA content, and birth weight in the INMA
771 (Spain) and ENVIRONAGE (Belgium) birth cohorts. *Environ. Health Perspect.* 124:659–65;
772 doi:10.1289/ehp.1408981.

773 Cohen Hubal EA, Moya J, Selevan SG. 2008. A lifestage approach to assessing children’s exposure.
774 *Birth Defects Res. B. Dev. Reprod. Toxicol.* 83:522–529; doi:10.1002/bdrb.20173.

775 Cox B, Leavey K, Nosi U, Wong F, Kingdom J. 2015. Placental transcriptome in development and
776 pathology: Expression, function, and methods of analysis. *Am. J. Obstet. Gynecol.* 213:S138–
777 S151; doi:10.1016/j.ajog.2015.07.046.

778 Deng Q, Lu C, Jiang W, Zhao J, Deng L, Xiang Y. 2017. Association of outdoor air pollution and indoor
779 renovation with early childhood ear infection in China. *Chemosphere* 169:288–296;
780 doi:10.1016/j.chemosphere.2016.11.079.

781 Deng Q, Lu C, Li Y, Sundell J, Norbäck D. 2016. Exposure to outdoor air pollution during trimesters of
782 pregnancy and childhood asthma , allergic rhinitis , and eczema. *Environ. Res.* 150:119–127;
783 doi:10.1016/j.envres.2016.05.050.

784 Dodd-Butera T, Quintana PJE, Ramirez-Zetina M, Batista-Castro AC, Sierra MM, Shaputnic C, et al.
785 2016. Placental biomarkers of PAH exposure and glutathione-S-transferase biotransformation
786 enzymes in an obstetric population from Tijuana, Baja California, Mexico. *Environ. Res.*
787 152:360–368; doi:10.1016/j.envres.2016.04.019.

788 Dötsch J, Trollmann R, Tzschoppe A, Struwe E, Schild R. 2010. Predicting development and disease in
789 infancy and childhood from placental function. In *Human Placenta: Structure and Development,*
790 *Circulation and Functions*, pp. 173–180.

791 Eidem H, Ackerman W, McGary K, Abbot P, Rokas A. 2015. Gestational tissue transcriptomics in term
792 and preterm human pregnancies: a systematic review and meta-analysis. *BMC Med. Genomics*
793 8:27; doi:10.1186/s12920-015-0099-8.

794 Ellis JA, Kemp AS, Ponsonby AL. 2014. Gene-environment interaction in autoimmune disease. *Expert*
795 *Rev. Mol. Med.* 16; doi:10.1017/erm.2014.5.

796 Fowler BA. 2012. Biomarkers in toxicology and risk assessment. *Mol. Clin. Environ. Toxicol.* 101: 459–
797 470.

798 Gallego Romero I, Pai A a, Tung J, Gilad Y. 2014. RNA-seq: impact of RNA degradation on transcript
799 quantification. *BMC Biol.* 12:42; doi:10.1186/1741-7007-12-42.

800 Ghosh R, Topinka J, Joad JP, Dostal M, Sram RJ, Hertz-Picciotto I. 2013. Air pollutants, genes and early
801 childhood acute bronchitis. *Mutat. Res. - Fundam. Mol. Mech. Mutagen.* 749:80–86;
802 doi:10.1016/j.mrfmmm.2013.04.001.

803 Gu Y, Sun J, Groome LJ, Wang Y. 2013. Differential miRNA expression profiles between the first and
804 third trimester human placentas. *Am J Physiol Endocrinol Metab.* 304: E836–E843.

805 Haque AK, Mancuso MG, Williams MG, Dodson RF. 1992. Asbestos in organs and placenta of five
806 stillborn infants suggests transplacental transfer. *Environ. Res.* 58:163–175; doi:10.1016/S0013-
807 9351(05)80212-9.

808 Heidinger BJ, Blount JD, Boner W, Griffiths K, Metcalfe NB, Monaghan P. 2012. Telomere length in
809 early life predicts lifespan. *Proc. Natl. Acad. Sci. U. S. A.* 109:1743–8;
810 doi:10.1073/pnas.1113306109.

811 Hincal F. 1986. Effects of exposure to air pollution and smoking on the placental aryl hydrocarbon
812 hydroxylase (AHH) activity. *Arch. Environ. Health* 41:377–383;
813 doi:10.1080/00039896.1986.9935782.

814 Hogg K, Blair JD, von Dadelszen P, Robinson WP. 2013. Hypomethylation of the LEP gene in placenta
815 and elevated maternal leptin concentration in early onset pre-eclampsia. *Mol. Cell. Endocrinol.*
816 367:64–73; doi:10.1016/j.mce.2012.12.018.

817 Hogg K, Price EM, Hanna CW, Robinson WP. 2012. Prenatal and perinatal environmental influences
818 on the human fetal and placental epigenome. *Clin. Pharmacol. Ther.* 92:716–26;
819 doi:10.1038/clpt.2012.141.

820 Janssen BG, Byun H-M, Gyselaers W, Lefebvre W, Baccarelli AA, Nawrot TS. 2015. Placental
821 mitochondrial methylation and exposure to airborne particulate matter in the early life
822 environment: An ENVIR *ON* AGE birth cohort study. *Epigenetics* 10:536–544;
823 doi:10.1080/15592294.2015.1048412.

824 Janssen BG, Godderis L, Pieters N, Poels K, Kici Ski M, Cuypers A, et al. 2013. Placental DNA
825 hypomethylation in association with particulate air pollution in early life. *Part. Fibre Toxicol.*
826 10:22; doi:10.1186/1743-8977-10-22.

827 Janssen BG, Munters E, Pieters N, Smeets K, Cox B, Cuypers A, et al. 2012. Placental mitochondrial

828 DNA content and particulate air pollution during in utero life. *Environ. Health Perspect.*
829 120:1346–52; doi:10.1289/ehp.1104458.

830 Jedrychowski WA, Perera FP, Camann D, Spengler J, Butscher M, Mroz E, et al. 2015. Prenatal
831 exposure to polycyclic aromatic hydrocarbons and cognitive dysfunction in children. *Environ.*
832 *Sci. Pollut. Res.* 22:3631–3639; doi:10.1007/s11356-014-3627-8.

833 Jirtle RL, Skinner MK. 2007. Environmental epigenomics and disease susceptibility. *Nat. Rev. Genet.*
834 8:253–262; doi:10.1038/embor.2011.125.

835 Kedryna T, Gumińska M, Lucyna Z. 2004. Pyruvate kinase activity in the placentas of women living in
836 polluted and unpolluted environments. *Med. Sci. Monit.* 10: 672–8.

837 Kelsey G, Michels KB. 2012. Epigenome changes during development. In *Epigenetic epidemiology*, pp.
838 77–103.

839 Kim J, Zhao K, Jiang P, Lu Z, Wang J, Murray JC, et al. 2012. Transcriptome landscape of the human
840 placenta. *BMC Genomics* 13:115; doi:10.1186/1471-2164-13-115.

841 Kingsley SL, Eliot MN, Whitsel EA, Huang Y, Karl T, Marsit CJ, et al. 2016. Maternal residential
842 proximity to major roadways, birth weight, and placental DNA methylation Samantha. 92–
843 93:43–49; doi:10.1016/j.envint.2016.03.020.Maternal.

844 Lambertini L, Chen J, Nomura Y. 2015. Mitochondrial gene expression profiles are associated with
845 maternal psychosocial stress in pregnancy and infant temperament. *PLoS One* 10:1–20;
846 doi:10.1371/journal.pone.0138929.

847 Lamichhane DK, Leem J, Lee J, Kim H. 2015. A meta-analysis of exposure to particulate matter and
848 adverse birth outcomes.

849 Levkovitz R, Zaretsky U, Gordon Z, Jaffa AJ, Elad D. 2013. In vitro simulation of placental transport:
850 Part I. Biological model of the placental barrier. *Placenta* 34:699–707;
851 doi:10.1016/j.placenta.2013.03.014.

852 Lewis RM, Demmelmair H, Gaillard R, Godfrey KM, Hauguel-De Mouzon S, Huppertz B, et al. 2013.
 853 The placental exposome: Placental determinants of fetal adiposity and postnatal body
 854 composition. *Ann. Nutr. Metab.* 63:208–215; doi:10.1159/000355222.

855 Linnane A, Ozawa T, Marzuki S, Tanaka M. 1989. Mitochondrial Dna Mutations As an Important
 856 Contributor To Ageing and Degenerative Diseases. *Lancet* 333:642–645; doi:10.1016/S0140-
 857 6736(89)92145-4.

858 Lu C, Deng L, Ou C, Yuan H, Chen X, Deng Q. 2017. Preconceptional and perinatal exposure to traffic-
 859 related air pollution and eczema in preschool children. *J. Dermatol. Sci.* 85: 85–95.

860 Machaalani R, Ghazavi E, Hinton T, Waters KA, Hennessy A. 2014. Cigarette smoking during
 861 pregnancy regulates the expression of specific nicotinic acetylcholine receptor (nAChR) subunits
 862 in the human placenta. *Toxicol. Appl. Pharmacol.* 276:204–212; doi:10.1016/j.taap.2014.02.015.

863 Magda Price E, Cotton AM, Peñaherrera MS, McFadden DE, Kobor MS, Robinson WP. 2012. Different
 864 measures of “genome-wide” DNA methylation exhibit unique properties in placental and
 865 somatic tissues. *Epigenetics* 7:652–663; doi:10.4161/epi.20221.

866 Marafie EM, Marafie I, Emery SJ, Waters R, Jones NJ. 2000. Biomonitoring the human population
 867 exposed to pollution from the oil fires in Kuwait: analysis of placental tissue using (32)P-
 868 postlabeling. *Environ. Mol. Mutagen.* 36:274–82; doi:10.1002/1098-2280(2000)36:4<274::AID-
 869 EM3>3.0.CO;2-D [pii].

870 Martens DS, Gouveia S, Madhloum N, Janssen BG, Plusquin M, Vanpoucke C, et al. 2017. Neonatal
 871 cord blood oxylipins and exposure to particulate matter in the early-life environment: An
 872 ENVIRONAGE birth cohort study. *Environ. Health Perspect.* 125:691–698; doi:10.1289/EHP291.

873 Martens DS, Nawrot TS. 2016. Air pollution stress and the aging phenotype: The telomere
 874 connection. *Curr. Environ. Heal. reports* 3:258–69; doi:10.1007/s40572-016-0098-8.

875 Mayeur S, Lancel S, Theys N, Lukaszewski MA, Duban-Deweert S, Bastide B, et al. 2014. Maternal
 876 calorie restriction modulates placental mitochondrial biogenesis and bioenergetic efficiency:

877 Putative involvement in fetoplacental growth defects in rats. *World Rev. Nutr. Diet.* 109:106–
878 107; doi:10.1159/000356110.

879 Mayeur S, Silhol M, Moitrot E, Barbaux S, Breton C, Gabory A, et al. 2010. Placental BDNF/TrkB
880 signaling system is modulated by fetal growth disturbances in rat and human. *Placenta* 31:785–
881 791; doi:10.1016/j.placenta.2010.06.008.

882 Meyer JN, Leung MCK, Rooney JP, Sendoel A, Hengartner MO, Kisby GE, et al. 2013. Mitochondria as
883 a target of environmental toxicants. *Toxicol. Sci.* 134:1–17; doi:10.1093/toxsci/kft102.

884 Miura K, Higashijima A, Murakami Y, Fuchi N, Tsukamoto O, Abe S, et al. 2016. Circulating levels of
885 pregnancy-associated, placenta-specific microRNAs in pregnant women with placental
886 abruption. *Reprod. Sci.* 1–7; doi:10.1177/1933719116653837.

887 Mumford JL, Lee X, Lewtas J, Young TL, Santella RM. 1993. DNA adducts as biomarkers for assessing
888 exposure to polycyclic aromatic hydrocarbons in tissues from Xuan Wei women with high
889 exposure to coal combustion emissions and high lung cancer mortality. *Environ. Health*
890 *Perspect.* 99: 83–87.

891 Mund M, Louwen F, Klingelhoefer D, Gerber A. 2013. Smoking and pregnancy - A review on the first
892 major environmental risk factor of the unborn. *Int. J. Environ. Res. Public Health* 10:6485–6499;
893 doi:10.3390/ijerph10126485.

894 Nomura Y, Lambertini L, Rialdi A, Lee M, Mystal EY, Grabie M, et al. 2014. Global methylation in the
895 placenta and umbilical cord blood from pregnancies with maternal gestational diabetes,
896 preeclampsia, and obesity. *Reprod. Sci.* 21:131–137; doi:10.1177/1933719113492206.

897 Nugent BM, Bale TL. 2015. The omniscient placenta: Metabolic and epigenetic regulation of fetal
898 programming. 39:28–37; doi:10.1161/CIRCRESAHA.116.303790.The.

899 Obolenskaya MY, Teplyuk NM, Divi RL, Poirier MC, Filimonova NB, Zadrozna M, et al. 2010. Human
900 placental glutathione S-transferase activity and polycyclic aromatic hydrocarbon DNA adducts
901 as biomarkers for environmental oxidative stress in placentas from pregnant women living in

902 radioactivity- and chemically-polluted regions. *Toxicol. Lett.* 196:80–86;
 903 doi:10.1016/j.toxlet.2010.03.1115.

904 Pandey a, Mann M. 2000. Proteomics to study genes and genomes. *Nature* 405:837–846;
 905 doi:10.1038/35015709.

906 Payne BAI, Wilson IJ, Yu-Wai-Man P, Coxhead J, Deehan D, Horvath R, et al. 2013. Universal
 907 heteroplasmy of human mitochondrial DNA. *Hum. Mol. Genet.* 22:384–390;
 908 doi:10.1093/hmg/ddt435.

909 Pedersen M, Wichmann J, Autrup H, Dang DA, Decordier I, Hvidberg M, et al. 2009. Increased
 910 micronuclei and bulky DNA adducts in cord blood after maternal exposures to traffic-related air
 911 pollution. *Environ. Res.* 109:1012–1020; doi:10.1016/j.envres.2009.08.011.

912 Rappazzo KM, Daniels JL, Messer LC, Poole C, Lobdell DT. 2014. Exposure to fine particulate matter
 913 during pregnancy and risk of preterm birth among women in New Jersey, Ohio, and
 914 Pennsylvania, 2000–2005. *Environ. Health Perspect.* 122:992–997; doi:10.1289/ehp.1307456.

915 Reddy M V, Kenny PC, Randerath K. 1990. 32P-assay of DNA adducts in white blood cells and
 916 placentas of pregnant women: lack of residential wood combustion-related adducts but
 917 presence of tissue-specific endogenous adducts. *Teratog. Carcinog. Mutagen.* 10: 373–384.

918 Rossner P, Tabashidze N, Dostal M, Novakova Z, Chvatalova I, Spatova M, et al. 2011. Genetic,
 919 biochemical, and environmental factors associated with pregnancy outcomes in newborns from
 920 the Czech Republic. *Environ. Health Perspect.* 119:265–71; doi:10.1289/ehp.1002470.

921 Rumbajan JM, Yamaguchi Y, Nakabayashi K, Higashimoto K, Yatsuki H, Nishioka K, et al. 2016. The
 922 HUS1B promoter is hypomethylated in the placentas of low-birth-weight infants. *Gene*
 923 583:141–146; doi:10.1016/j.gene.2016.02.025.

924 Ryan JG, Davis RK, Bloch JR. 2012. The placenta as a research biospecimen. *JOGNN - J. Obstet.*
 925 *Gynecol. Neonatal Nurs.* 41:834–845; doi:10.1111/j.1552-6909.2012.01420.x.

926 Saenen ND, Plusquin M, Bijmens E, Janssen BG, Gyselaers W, Cox B, et al. 2015. In utero fine particle
 927 air pollution and placental expression of genes in the brain-derived neurotrophic factor
 928 signaling pathway: An ENVIRONAGE birth cohort study. *Environ. Health Perspect.* 123:834–840;
 929 doi:10.1289/ehp.1408549.

930 Saenen ND, Vrijens K, Janssen BG, Madhloum N, Peusens M, Gyselaers W, et al. 2016. Placental
 931 nitrosative stress and exposure to ambient air pollution during gestation: A population study.
 932 *Am. J. Epidemiol.* 184:442–449; doi:10.1093/aje/kww007.

933 Saenen ND, Vrijens K, Janssen BG, Roels HA, Neven KY, Vanden Berghe W, et al. 2017. Lower
 934 placental leptin promoter methylation in association with fine particulate matter air pollution
 935 during pregnancy and placental nitrosative stress at birth in the ENVIRONAGE cohort. *Environ.*
 936 *Health Perspect.* 125:262–268; doi:10.1289/EHP38.

937 Shah PS, Balkhair T. 2011. Air pollution and birth outcomes: A systematic review. *Environ. Int.*
 938 37:498–516; doi:10.1016/j.envint.2010.10.009.

939 Sorkun HC, Bir F, Akbulut M, Divrikli U, Erken G, Demirhan H, et al. 2007. The effects of air pollution
 940 and smoking on placental cadmium, zinc concentration and metallothionein expression.
 941 *Toxicology* 238:15–22; doi:10.1016/j.tox.2007.05.020.

942 Sram RJ, Binkova B, Dejmek J, Chvatalova I, Solansky I, Topinka J. 2006. Association of DNA adducts
 943 and genotypes with birth weight. *Mutat. Res. - Genet. Toxicol. Environ. Mutagen.* 608:121–128;
 944 doi:10.1016/j.mrgentox.2006.04.022.

945 Sram RJ, Binkova B, Rossner P, Rubes J, Topinka J, Dejmek J. 1999. Adverse reproductive outcomes
 946 from exposure to environmental mutagens. *Mutat. Res. - Fundam. Mol. Mech. Mutagen.*
 947 428:203–215; doi:10.1016/S1383-5742(99)00048-4.

948 Stejskalova L, Pavek P. 2011. The function of cytochrome P450 1A1 enzyme (CYP1A1) and aryl
 949 hydrocarbon receptor (AhR) in the placenta. *Curr. Pharm. Biotechnol.* 12:16;
 950 doi:10.2174/138920111795470994.

951 Sun C, Velazquez MA, Fleming TP. 2016. DOHaD and the periconceptional period, a critical window in
 952 time. In *The Epigenome and Developmental Origins of Health and Disease*, pp. 33–47.

953 Surowiec I, Karimpour M, Gouveia-Figueira S, Wu J, Unosson J, Bosson JA, et al. 2016. Multi-platform
 954 metabolomics assays for human lung lavage fluids in an air pollution exposure study. *Anal.*
 955 *Bioanal. Chem.* 408:4751–4764; doi:10.1007/s00216-016-9566-0.

956 Topinka J, Binková B, Mracková G, Stávková Z, Benes I, Dejmek J, et al. 1997a. DNA adducts in human
 957 placenta as related to air pollution and to GSTM1 genotype. *Mutat. Res.* 390: 59–68.

958 Topinka J, Binkova B, Mrackova G, Stavkova Z, Peterka V, Benes I, et al. 1997b. Influence of GSTM1
 959 and NAT2 genotypes on placental DNA adducts in an environmentally exposed population.
 960 *Environ. Mol. Mutagen.* 30: 184–195.

961 Topinka J, Milcova A, Libalova H, Novakova Z, Rossner P, Balascak I, et al. 2009. Biomarkers of
 962 exposure to tobacco smoke and environmental pollutants in mothers and their transplacental
 963 transfer to the foetus. Part I: Bulky DNA adducts. *Mutat. Res. - Fundam. Mol. Mech. Mutagen.*
 964 669:13–19; doi:10.1016/j.mrfmmm.2009.04.011.

965 Tsamou M, Vrijens K, Madhloum N, Lefebvre W, Vanpoucke C, Nawrot TS. 2016. Air pollution-
 966 induced placental epigenetic alterations in early life: a candidate miRNA approach. *Epigenetics*
 967 2294; doi:10.1080/15592294.2016.1155012.

968 Tzoulaki I, Ebbels TMD, Valdes A, Elliott P, Ioannidis JPA. 2014. Design and analysis of metabolomics
 969 studies in epidemiologic research: A primer on-omic technologies. *Am. J. Epidemiol.* 180:129–
 970 139; doi:10.1093/aje/kwu143.

971 Vaiserman A. 2015. Epidemiologic evidence for association between adverse environmental
 972 exposures in early life and epigenetic variation: a potential link to disease susceptibility? *Clin.*
 973 *Epigenetics* 7:96; doi:10.1186/s13148-015-0130-0.

974 Vineis P, Chadeau-Hyam M, Gmuender H, Gulliver J, Herceg Z, Kleinjans J, et al. 2016. The exposome
 975 in practice: Design of the EXPOsOMICS project. *Int. J. Hyg. Environ. Health* 1–10;

doi:10.1016/j.ijheh.2016.08.001.

Vrijens K, Bollati V, Nawrot TS. 2015. MicroRNAs as potential signatures of environmental exposure or effect: A systematic review. *Environ. Health Perspect.* 123:399–411; doi:10.1289/ehp.1408459.

Vrijheid M, Slama R, Robinson O, Chatzi L, Coen M, van den Hazel P, et al. 2014. The human early-life exposome (HELIX): Project rationale and design. *Environ. Health Perspect.* 122:535–544; doi:10.1289/ehp.1307204.

Walsh JM, Byrne J, Mahony RM, Foley ME, McAuliffe FM. 2014. Leptin, fetal growth and insulin resistance in non-diabetic pregnancies. *Early Hum. Dev.* 90:271–274; doi:10.1016/j.earlhumdev.2014.03.007.

Weldy CS, Liu Y, Liggitt HD, Chin MT. 2014. In Utero exposure to diesel exhaust air pollution promotes adverse intrauterine conditions, resulting in weight gain, altered blood pressure, and increased susceptibility to heart failure in adult mice. *PLoS One* 9; doi:10.1371/journal.pone.0088582.

Whyatt RM, Bell D a, Jedrychowski W, Santella RM, Garte SJ, Cosma G, et al. 1998. Polycyclic aromatic hydrocarbon-DNA adducts in human placenta and modulation by CYP1A1 induction and genotype. *Carcinogenesis* 19:1389–92; doi:10.1093/carcin/19.8.1389.

Whyatt RM, Garte SJ, Cosma G, Bell DA, Jedrychowski W, Wahrendorf J, et al. 1995. CYP1A1 messenger RNA levels in placental tissue as a biomarker of environmental exposure. *Cancer Epidemiol. Biomarkers Prev.* 4: 147–153.

Wick P, Malek A, Manser P, Meili D, Maeder-Althaus X, Diener L, et al. 2010. Barrier capacity of human placenta for nanosized materials. *Environ. Health Perspect.* 118:432–436; doi:10.1289/ehp.0901200.

Wild CP. 2012. The exposome: From concept to utility. *Int. J. Epidemiol.* 41:24–32; doi:10.1093/ije/dyr236.

1000 Wilhelm-Benartzi CS, Houseman EA, Maccani MA, Poage GM, Koestler DC, Langevin SM, et al. 2012.
 1001 In utero exposures, infant growth, and DNA methylation of repetitive elements and
 1002 developmentally related genes in human placenta. *Environ. Health Perspect.* 120:296–302;
 1003 doi:10.1289/ehp.1103927.
 1004 World Health Organization. 2010. Biomarkers and human biomonitoring. WHO Train. Packag. Heal.
 1005 Sect.
 1006 Zhang X, Lin S, Funk WE, Hou L. 2013. Environmental and occupational exposure to chemicals and
 1007 telomere length in human studies. *Occup. Environ. Med.* 70:743–9; doi:10.1136/oemed-2012-
 1008 101350.
 1009 Zhang X, Pei L, Li R, Zhang W, Yang H, Li Y, et al. 2015. Spina bifida in fetus is associated with an
 1010 altered pattern of DNA methylation in placenta. *J. Hum. Genet.* 60:1–7;
 1011 doi:10.1038/jhg.2015.80.
 1012