

2016•2017
FACULTY OF MEDICINE AND LIFE SCIENCES
Master of Biomedical Sciences

Master's thesis

Heart rate responses during exercise in obese adolescents: relation with cardiovascular health

Supervisor :
Prof. Dr. Guy MASSA
Prof. dr. Dominique HANSEN

Marjolein Beyens

Thesis presented in fulfillment of the requirements for the degree of Master of Biomedical Sciences

Transnational University Limburg is a unique collaboration of two universities in two countries: the University of Hasselt and Maastricht University.



Universiteit Hasselt | Campus Hasselt | Martelarenlaan 42 | BE-3500 Hasselt
Universiteit Hasselt | Campus Diepenbeek | Agoralaan Gebouw D | BE-3590 Diepenbeek



2016•2017
FACULTY OF MEDICINE AND LIFE
SCIENCES
Master of Biomedical Sciences

Master's thesis

Heart rate responses during exercise in obese
adolescents: relation with cardiovascular health

Supervisor :
Prof. Dr. Guy MASSA
Prof. dr. Dominique HANSEN

Marjolein Beyens

*Thesis presented in fulfillment of the requirements for the degree of Master of
Biomedical Sciences*

Acknowledgements

Firstly, I would like to express my sincere gratitude to my supervisors, Prof. dr. Dominique Hansen and Prof. Dr. Guy Massa. They have allowed me to learn a tremendous amount throughout the eight months of training and provided me with extensive guidance in writing this thesis.

I am grateful to all of those with whom I have had the pleasure to work during this project: Cardiologist Dr. Torab Al Hatawe, for performing many an ultrasound exam, and analyzing them to boot; Dr. Renate Zeevaert, for helping us with the recruitment of many of our patients; and the whole Children's obesity team at Jessa Hospital, dieticians Trui Otte and Sigrid Vanhaesebrouck and psychologist Kelly Faust.

My sincere thanks also go to the nurse's team at the pediatric unit of Jessa hospital for their hard work and commitment to this project.

To the personnel at ReGo, a huge thank you for putting up with our questions and banter for this extended period.

I thank my fellow interns, Hajar Boujemaa and Wouter Franssen for the stimulating discussions and for all the fun we have had over the last months. Wouter especially, working with you has been a pleasure and a meaningful learning experience.

Finally, I would like to thank my parents for supporting me throughout writing this thesis and my whole school career. Finally, I wish to thank my partner Tim, who provides me with unending inspiration and support.

Table of contents

Acknowledgements	I
Table of contents	III
List of abbreviations	V
Abstract.....	VII
1. Introduction.....	1
1.1. Obesity: prevalence and health effects	1
1.2. Obesity and exercise.....	4
1.3. Hypothesis and objectives.....	5
2. Materials and methods	7
2.1. Subjects and study design.....	7
2.2. Anthropometry, assessment of physical activity and maturation stage.....	8
2.3. Fasting blood samples	8
2.4. Echocardiography and carotid intima thickness measurement	9
2.5. Maximal cardiopulmonary testing	10
2.6. Statistics.....	11
3. Results	13
3.1. Participant characteristics.....	13
3.2. Cardiopulmonary testing parameters	14
3.3. Blood parameters	17
3.4. Metabolic risk factor	19
3.5. Electrocardiogram parameters.....	19
3.6. Ultrasound parameters.....	20
3.7. Association between metabolic risk factor and heart rate responses.....	21
3.8. Multivariate models for heart rate responses.....	22
3.8.1. Metabolic parameter models for heart rate responses.....	22
3.8.2. Cardiovascular parameter models for heart rate responses.....	23
3.8.3. Electrophysiological parameter models for heart rate responses.....	23
4. Discussion	25
4.1. Markers for cardiovascular abnormalities	25

4.2.	Cardiopulmonary testing outcomes.....	26
5.	Conclusion	29
6.	Limitations.....	29
7.	Future perspectives.....	29
8.	References.....	31
9.	Appendices.....	37

List of abbreviations

A mitral	trans-mitral peak late diastolic velocity	PETO ₂	end-tidal O ₂ tension
AF	alkaline phosphatase	Psys	systolic blood pressure
ALT	alanine aminotransferase	QTc	corrected QT
AST	aspartate aminotransferase	RAS	renal renin-angiotensin system
BAT	brown adipose tissue	RER	respiratory exchange ratio
BF	breathing frequency	RIA	radioimmunoassay
BIA	bioelectrical impedance analysis	rpm	revolutions per minute
BMI	body Mass Index	SFC	skinfold calliper
BP	blood pressure	T2DM	type 2 diabetes mellitus
Ca ²⁺	calcium	T4	thyroxine
CIMT	carotid intima media thickness	TAG	triacylglycerol
CO	cardiac output	TNF- α	tumour necrosis factor- α
CPET	cardiopulmonary testing	TSH	thyroid stimulating hormone
CRI	chronotropic index	TTE	transthoracic echocardiography
CRP	c-reactive protein	VCO ₂	carbon dioxide output
DT	deceleration time	Vd/Vt	physiological dead-space/tidal volume ratio
E mitral	trans-mitral peak early diastolic velocity	VE	expired volume
E'	early diastolic mitral annular velocity	VE/VCO ₂	ventilatory equivalents for CO ₂
ECG	electrocardiogram	VE/VO ₂	ventilatory equivalents for O ₂
EF	ejection fraction	VO ₂	oxygen uptake
FFA	free fatty acids	VO ₂ /HR	oxygen pulse
GGT	gamma-glutamyl transpeptidase	Vt	tidal volume
Hb	haemoglobin	VT1	first ventilatory threshold
HbA1c	glycosylated haemoglobin	VT2	second ventilatory threshold
HC	hip circumference	WAT	white adipose tissue
HDL	high-density lipoprotein	WC	waist circumference
HOMA-IR	homeostatic model assessment for insulin resistance	WHO	World Health Organisation
HR	heart rate	WHR	waist-hip ratio
HRR	heart rate recovery	WIV-ISP	Belgian scientific institute of public health
HRreserve	heart rate reserve		
Ht	haematocrit		
IL-6	interleukin-6		
IOTF	International Obesity Task Force		
K ⁺	potassium		
LA	left atrium		
LDL	low-density lipoprotein		
LV	left ventricle		
MetS	metabolic syndrome		
MRF	metabolic risk factor		
NASH	non-alcoholic steatohepatitis		
NG	lean group		
OG	obese group		
OITF	International Obesity Task Force		
OUES	oxygen uptake efficiency slope		
PAQ-A	physical activity questionnaire for adolescents		
Pdia	diastolic blood pressure		
PETCO ₂	end-tidal CO ₂ tension		

Abstract

Introduction: Childhood obesity is a growing epidemic associated with important cardiovascular, gastrointestinal, respiratory and orthopaedic comorbidities. In adults with obesity, a changed peak heart rate (HR_{peak}) during exercise is independently related to an elevated risk for adverse cardiovascular events. However, the prognostic relevance of this parameter in adolescents with obesity remains to be explored.

Objectives: The goal of this project was to study the association between HR responses and (i) cardiac function, (ii) cardiovascular autonomic function, (iii) atherosclerosis and (iv) cardiac electrophysiology to assess the importance of HR responses during exercise as a prognostic tool for cardiovascular disease in obese adolescents.

Methods: Twenty-four obese (OG BMI: 32.1 ± 4.3 kg/m², age: 13.4 ± 1.1 years) and 24 lean (LG BMI: 19.6 ± 2.5 kg/m², age: 13.9 ± 1.5 years) adolescents performed a maximal cardiopulmonary exercise test (CPET), with assessment of cardiovascular and respiratory outcomes. Subject groups were age and gender matched, Tanner stage and physical activity levels were assessed. Transthoracic echocardiography (TTE) was performed, as well as measurement of carotid intima media thickness (CIMT). Fasting blood samples were collected to evaluate glycaemic control, liver function and cardiovascular risk factors. Multivariate regression models were used to explore relationships between CPET outcomes and secondary measurements.

Results: HR_{peak} was not different between two groups ($p=0.97$). However, heart rate reserve (HR reserve) ($p<0.01$) and heart rate recovery (HRR) at 30 ($p=0.03$), 60 ($p=0.02$) and 120 seconds ($p<0.01$) after exercise were decreased in the OG. Other CPET outcomes were not significantly altered except for maximal power output ($p=0.01$). HRreserve and HRR were associated with metabolic risk factors seen in obesity and factors indicative of cardiovascular abnormalities.

Conclusion: Significant alterations in HRreserve and HRR suggest cardiovascular autonomic function is distorted in obese adolescents. Although the data indicates that HR_{peak} is not changed in obese adolescents, HRreserve and HRR might be of prognostic value for cardiovascular disease risk in obese adolescents. Further research should be implemented to discover the associations between cardiac autonomic function and adverse cardiac events and the aetiology behind the development of distorted autonomic function in obese adolescents.

1. Introduction

1.1. Obesity: prevalence and health effects

The World Health Organisation (WHO) defines overweight and obesity as an abnormal or excessive fat accumulation that may have adverse health effects. Defining obesity is essential in observing prevalence rates as well as monitoring and intervening with the process of weight gain (1). Body mass index (BMI [kg/m^2]) is the ratio of weight to height that is commonly used to classify weight status. BMI is calculated by dividing one's weight in kilograms by the square of their height in meters.

On a population level, BMI provides a useful, rapid measure of overweight and obesity. However, when considering individuals, it should be used with caution as the same BMI between individuals might not correspond to the same degree of fatness or adiposity. In adults, overweight is defined as a BMI greater than or equal to $25 \text{ kg}/\text{m}^2$ and obesity as a BMI greater than or equal to $30 \text{ kg}/\text{m}^2$ by the WHO (2). When defining childhood overweight and obesity, age, as well as gender, needs to be considered additionally. The International Obesity Task Force (IOTF) cut-offs have been widely used to assess prevalence rates for childhood overweight and obesity and are defined by values of BMI at age of 18 years (3).

According to a report by the Belgian scientific institute of public health (WIV-ISP), the prevalence rates of obesity have risen to 7% in children from 2 to 17 years old in 2013 in Belgium. The prevalence of overweight children is even higher, one in five children are too heavy considering their age and height. Moreover, both these numbers have risen since 1997 (4). Worldwide, an estimated 170 million children (aged less than 18 years old) are now overweight (2012) (5). Although significant efforts are being made to prevent and reduce the prevalence of childhood obesity, obesity continues to become even more prevalent (5, 6)

Childhood obesity is caused by diverse genetic and non-genetic factors. The "thrifty gene" hypothesis partially explains the global obesity epidemic by natural selection favouring a thrifty energy metabolism in times of limited food supply and high energy expenditure (7). Current low-activity lifestyles and an overabundance of high-energy foods, both characteristics of western countries, combined with the genetic predisposition of the thrifty genes, could lead to obesity. Even disregarding this theory, low levels of physical activity and easier access to high energy foods, have been implicated as important factors in the development of paediatric obesity (8, 9).

The residual energy that is left over when caloric intake is higher than caloric expenditure leads to energy stored in the form of triacylglycerol (TAG) in adipose tissue (see Figure 1). The excessive storage that occurs in obesity eventually incites an excess release of free fatty acids (FFA), due to excessive sympathetic nervous system stimulation and resulting increase in lipolysis. Excess FFA in the bloodstream lead to lipotoxicity, as lipids create oxidative stress in cells (10). The fat cells or adipocytes that form the adipose tissue do not only store TAGs, but also form an important endocrine tissue, releasing a wide range of so-called adipokines. Adipokines range from proteo-hormones such as leptin and adiponectin, that help regulate fat mass and appetite (11, 12), to pro-inflammatory cytokines such as interleukin-6 (IL-6) and tumour necrosis factor- α (TNF- α) (13). Particularly white adipose tissue (WAT) in visceral fat depots, rather than brown adipose tissue (BAT), contribute to the creation of a low-grade inflammatory state by excretion of these cytokines (14). Adipocytes also activate fat-associated macrophages which in turn also excite inflammation (15).

The low-grade inflammation alongside the created lipotoxicity form an intricate network that makes obese patients at greater risk of developing cardiovascular diseases such as high blood pressure and atherosclerosis, endocrine disorders such as insulin resistance and type 2 diabetes mellitus (T2DM) and gastrointestinal problems e.g. gallstones and non-alcoholic fatty liver disease or non-alcoholic steatohepatitis (NASH) (7, 16, 17). Hypertension in obesity is mostly caused by adipokines that lead to excessive secretion of hormones of the renal renin-angiotensin system (RAS), which enhance endothelial vasomotor tone (18). Atherosclerosis is provoked by increased inflammation and further exacerbated by insulin resistance (19). The insulin resistance, in turn, is caused by inflammatory processes that decrease insulin sensitivity and the effect of lipotoxicity in the islets of the pancreas, which decrease secretion of insulin by the β -cells (20). T2DM emerges when insulin resistance worsens and when dyslipidaemia arises from the increased FFAs and TAGs in circulation (21). Gallstones become more prevalent in subjects with obesity due to enhanced lipolysis and in turn saturation of bile, promoting the formation of gallstones (22). NASH arises when inflammatory adipokines and lipotoxicity work together in the liver to cause the fatty and inflamed state (19).

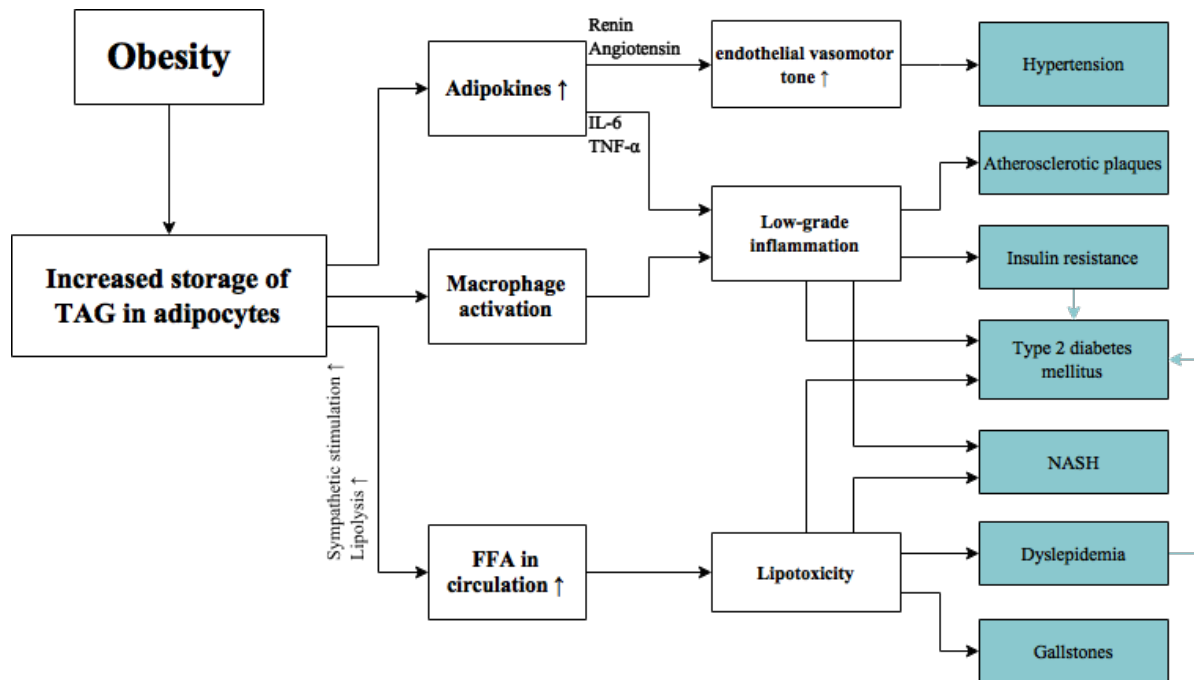


Figure 1: Pathology of obesity.

Abbreviations: TAG, triacylglyceride; FFA, free fatty acids; NASH, non-alcoholic steatohepatitis (10).

Before these conditions fully establish themselves, a cluster of risk factors identified as the metabolic syndrome (MetS) is seen in patients. The MetS risk factors include hypertension, hypertriglyceridemia, low HDL cholesterol and hyperinsulinemia, and are early, though reversible, stages of the comorbidities mentioned above (23).

Beyond these issues, obesity can cause problems of respiratory and orthopaedic origins. Comorbidities involving the respiratory system include obesity hypoventilation syndrome or Pickwick syndrome, resulting from the accumulation of adipose tissue, confining the upper respiratory tracts and adversely affecting breathing (16). The low-grade inflammatory state can exacerbate this condition by affecting bronchioles (24). The increased pressure on the joints due to weight can cause flatfoot and combined with inflammatory effects of adipokines can cause osteoarthritis. Obese adolescents can also experience psychosocial effects because of the social stigma associated with obesity, quality of life can, therefore, be reduced when compared to lean peers (16). Furthermore, not only does obesity contribute to illness in adolescence, it leads to increased morbidity and mortality in adulthood, even if the weight is normalised in adulthood (25). Lastly, obesity significantly increases the risk of certain cancers, such as breast cancer and kidney cancer (26).

Next to the disease and psychosocial burden on the patients, society carries a substantial medical and financial burden. Should the obese status persist through adulthood, costs are estimated to be much higher. A German study estimates a cost up to € 207 000 per obese adult (27). The total cost of overweight and obesity in European countries is 0.7% to 8% of the total healthcare expenditure, in which obesity is the main contributor (4).

1.2. Obesity and exercise

Other negative effects of excess body mass in adolescents are suboptimal responses to exercise. Often found is a reduced peak oxygen uptake per kg of lean tissue (28). Some studies reported a reduced peak cycling power output (29, 30) or a reduction in peak heart rate (HR_{peak})(29, 31) in obese adolescents compared to their lean counterparts. Other studies mention changes in heart rate reserve (HR_{reserve}) and heart rate recovery (HRR) after exercise (32, 33). In obese adults, HR_{peak} is independently related to an elevated glycosylated haemoglobin (HbA1c) concentration, which is used as a biomarker in diabetes diagnosis (34). Moreover, HR_{peak} is correlated with the risk of major adverse cardiovascular events and premature death and lower exercise tolerance (34, 35). Despite the established association between HR_{peak} and prognosis in obese adults, the validity of HR_{peak} as a prognostic marker in obese adolescents has not yet been explored. Assessing HR_{peak} using cardiopulmonary testing (CPET) would then provide a non-invasive, inexpensive and safe early predictor for cardiovascular incidents later in life (36). CPET allows the researcher to study the responses of both the cardiovascular and ventilatory systems to exercise stress. Gas exchange measurements of oxygen (O₂) and carbon dioxide (CO₂) are accompanied by an electrocardiogram (ECG), heart rate monitoring and blood pressure measurements. Gas exchange at the airways is interconnected with cardiac output and pulmonary blood flow as well as extraction of O₂ and feedback of CO₂ by the muscles, which is visualised in Figure 2. Because of the multifaceted outcomes CPET provides, a large range of dysfunctions can be discovered, even if they are only present when stress, i.e. exercise is put upon the body (37).

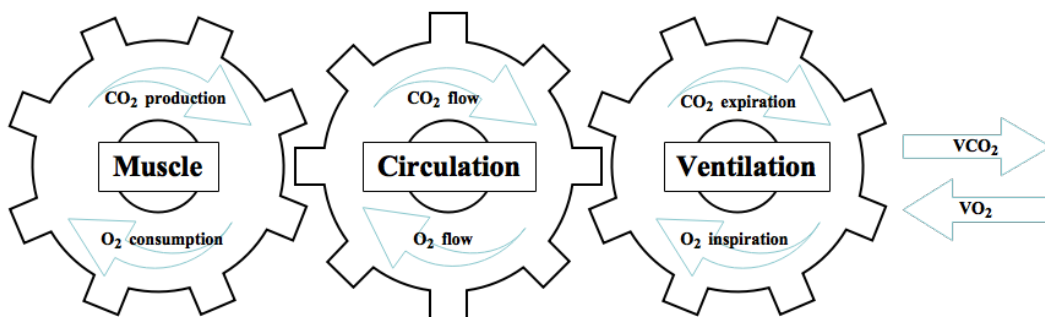


Figure 2: Coupling between muscle metabolism, blood circulation and ventilation, the foundation of cardiopulmonary testing(37).

In line with findings in obese adults, exercise testing and particularly HRpeak and heart rate responses may indicate several asymptomatic abnormalities in obese adolescents (38). For example, blood catecholamine (39) and potassium (40) concentrations during exercise are lowered in some obese adolescents compared to leans. Furthermore, obesity in adolescents can be associated with structural abnormalities of the heart or ventricular stiffness (41) and impaired blood pressure regulation and cardiovascular autonomic dysfunction (42). Some studies have also found atherosclerosis (43) in obese adolescents or cardiac electrophysiological abnormalities (44).

1.3. Hypothesis and objectives

The goal of this investigation was to examine whether HR responses during exercise testing may be an early indicator for cardiovascular abnormalities in obese adolescents. We hypothesised that HR responses during exercise testing are a sensitive and specific marker for cardiac dysfunction, cardiovascular autonomic abnormalities, atherosclerosis and cardiac electrophysiology abnormalities in obese adolescents.

The first aim of the proposed project was to compare CPET outcomes, HR responses in particular, between two groups, an obese group and a lean group. Secondly, the association between HR responses and (i) cardiac function, (ii) cardiovascular autonomic function, (iii) blood potassium concentrations at rest and at peak exercise, (iv) atherosclerosis, and (v) cardiac electrophysiology will be studied. In this regard, the diagnostic value of HR responses during maximal exercise testing will be clarified in obese adolescents, and the physiology behind the development of heart disease in obese adolescents can be explored.

2. Materials and methods

2.1. Subjects and study design

Forty-eight adolescents between the ages of 12 and 16 were recruited between November 2016 and May 2017 into the current cohort case-control clinical study. Subjects were split up into two groups of 24 subjects: the obese group (OG) and lean group (NG). The subjects were classified as obese or of normal weight based on BMI (kg/m^2) according to the age- and gender-specific IOTF centile curves. Overweight and obesity were classified using BMI cut-off points matching an adult BMI of over 25 and 30 kg/m^2 respectively. Normal weight was classified with BMI cut-off points consistent with adult BMI of between 18,5 and 25 kg/m^2 .

Obese participants were recruited from the childhood obesity clinic at Jessa Hospital vzw (Hasselt, Belgium) where they were referred to for overweight/obesity or indications of underlying pathologies, (for instance insulin resistance). Patients were invited to take part in the study if they were not using any medications with effects on exercise performance at the time and if they were free from clinically diagnosed T2DM or cardiovascular or respiratory diseases. The lean group was recruited in several manners: recruitment posters with a small description and contact information were circulated in different places, a classified advertisement was placed in a local newspaper and word-of-mouth also provided a few participants. Interested subjects were invited to participate in the study if they were not currently using any performance-altering medication and if they were free from chronic cardiovascular, respiratory or metabolic diseases.

Ethics approval was acquired from the medical ethical committees of both Jessa Hospital vzw and Hasselt University. Written informed consent was obtained from every participant and their parent or legal guardian after careful explanation of the study aims and methods. The study was conducted according to the principles outlined in the Helsinki Declaration as stated by the World Medical Association in 2013.

During one day hospitalisation, all participants were subjected to following exams in a chronological order.

1. Anthropometry, assessment of physical activity and maturation stage
2. Fasting blood sampling
3. Echocardiography and carotid intima thickness measurement
 - Standardized meal

4. Maximal cardiopulmonary exercise testing with blood lactate and potassium assessment at rest and peak exercise

2.2. Anthropometry, assessment of physical activity and maturation stage

Height was measured to the nearest 0.1 cm using a Harpenden wall-mounted stadiometer (De Grood metaaltechniek, Nijmegen, The Netherlands). Body weight was determined in fasting condition and in underwear to the nearest 0.1 kg on a digital scale (Seca, California, USA). BMI was then calculated from the ratio of weight (in kg) to squared height (m^2). BMI was defined using the age- and gender-specific IOTF centile curves (3). Waist and hip circumference (WC; HC) were determined using measurement tape. WC was measured across the umbilicus, HC was determined at the height of the iliac crest. Concurrently waist-hip ratio was calculated (WHR, waist circumference (cm)/ hip circumference (cm)). Fat percentage was assessed in two manners. Firstly, by using bioelectrical impedance analysis (BIA) with the Bodystat 1500 MDD (EuroMedix POC, Leuven, Belgium) (45) and secondly by means of a Harpenden skinfold calliper (SFC) (Baty International, West Sussex, UK). Skinfold thickness at the medial triceps area and subscapular area were then used to calculate body fat percentage according to a protocol by Slaughter et al., 1988 (46, 47). Blood pressure was evaluated using an automatic blood pressure monitor from Omron (Hoofddorp, The Netherlands). Physical maturity was assessed using the Tanner scale, which defines physical measurements of development based on external primary and secondary sex characteristics (48, 49). It was assessed in accordance with the observation by a paediatrician and the adolescents' own opinion using a graphic depiction. The level of physical activity was determined using the validated Dutch physical activity questionnaire for adolescents (PAQ-A) (50). The result is an activity score between 1 and 5, a score of 1 indicates low physical activity, whereas a score of 5 indicates high physical activity. The score is based on physical activity over the past week during school time, over lunch, right after school, in the evening and during the weekend.

2.3. Fasting blood samples

Next, fasting blood samples were collected for the analysis of: plasma glucose, insulin, iron, ferritin, thyroid-stimulating hormone (TSH), free T4, cortisol, aspartate aminotransferase (AST), gamma-glutamyl transpeptidase (GGT), alanine aminotransferase (ALT), alkaline phosphatase (ALP), uric acid, calcium (Ca^{2+}), potassium (K^+), total cholesterol, high-density lipoprotein (HDL) cholesterol, low-density lipoprotein (LDL) cholesterol, triglycerides, and c-reactive protein (CRP) concentrations (Cobas®8000 modular analyser (F. Hoffmann-La Roche Ltd., Basel, Switzerland)). Blood glycated haemoglobin concentration (HbA_{1c}) was measured using ion exchange chromatography (Menarini HA-8180 HbA1C auto-analyser, Menarini Diagnostics, Diegem, Belgium). Blood leptin concentration was

measured using radioimmunoassay (RIA) (LINCO Research Inc., Saint Louis, MI, USA). Blood haemoglobin, haematocrit, and white blood cells were automatically evaluated using Siemens Advia 2120 (Siemens Healthcare Diagnostics, Deerfield, IL, USA). Information on which collection tubes were used can be found in Appendix 1. From fasting glucose and insulin concentration, homeostatic model assessment for insulin resistance (HOMA-IR) was calculated by: $\text{fasting glucose (mg/dl)} * \text{fasting insulin concentration } (\mu\text{U/ml}) / 405$ (51). From blood parameter data, together with anthropometric data, a single-factor model, metabolic risk factor (MRF), underlying the MetS was created (52). MRF was calculated as the sum of standardized WC (cm), standardized insulin concentration (pmol/l), standardized triglyceride/HDL ratio and standardized mean arterial pressure (MAP), divided by four. Standard score or z-scores were calculated by subtracting the population mean from the value and dividing this number by the standard deviation (53).

2.4. Echocardiography and carotid intima thickness measurement

Ultrasound procedures were performed using the Vivid 7 Ultrasound Machine by GE Healthcare (Milwaukee, USA). Transthoracic echocardiography (TTE) with tissue Doppler was executed by an experienced cardiologist with the patient in a reclined position, using the GE M4S Matrix Array sector transducer (GE Healthcare, Milwaukee, USA). Left-ventricular (LV) systolic and diastolic function was assessed using following parameters: ejection fraction (EF), LV septum width (mm), LV diameter (mm), left-atrial (LA) diameter (mm), trans-mitral peak early diastolic velocity (E mitral [cm/sec]), trans-mitral peak late diastolic velocity (A mitral [cm/sec]), mitral E/A ratio, deceleration time (DT [sec]), E/E'-ratio (E'=Early diastolic mitral annular velocity) and cardiac output (CO [l/min]) (54). Ejection fraction was estimated using Simpson's rule algorithm (55) on the apical two and four chamber views. LV septum width, LV diameter and LA diameter were assessed on the parasternal long axis (PLAX). E mitral, A mitral, DT and CO were evaluated in the apical four chamber view. An algorithm was used to evaluate left ventricular diastolic function by assessment of left atrial pressure (LAP), visualised in Appendix 2 (56).

Following echocardiography, carotid intima media thickness (CIMT) was measured by ultrasound of the lateral cervical area using the GE 9L-D linear transducer with the patient in supine position. CIMT is measured in end-diastolic position on the far wall of the left and right common carotid artery just inferior to the bifurcation (57). All ultrasound images were analysed EchoPAC Clinical Workstation Software (GE Healthcare, Milwaukee, USA).

2.5. Maximal cardiopulmonary testing

After consumption of a standardized meal (382 kcal, 9.75 g fat, 62.5 g carbohydrates, 10.6 g protein), a maximal CPET was executed on an electronically braked cycle and corresponding CardioSoft software from GE Healthcare (Milwaukee, USA). After one minute of sitting on the bike and one minute of unloaded cycling, the exercise test was initiated with pedalling at a starting load of 40 W, thereafter the resistance increased continuously with 20 W every minute. A cycling frequency of 60 to 70 revolutions per minute (rpm) had to be maintained. By continuous pulmonary gas exchange analysis, oxygen uptake (VO_2), carbon dioxide output (VCO_2) and other respiratory parameters (expired volume (VE), ventilatory equivalents for O_2 (VE/VO_2), ventilatory equivalents for CO_2 (VE/VCO_2), tidal volume (Vt), physiological dead-space-tidal volume ratio (Vd/Vt), breathing frequency (BF), respiratory exchange ratio ($\text{RER}(\text{VO}_2/\text{VCO}_2)$), end-tidal O_2 tension (PETO_2), end-tidal CO_2 tension (PETCO_2), oxygen pulse (VO_2/HR)) were collected breath-by-breath and averaged every 10 seconds. Heart rate (HR) was monitored and averaged every 10 seconds and more specific cardiac electrophysiological parameters were continuously monitored using a 12-lead ECG device (KISS™ multilead, GE Healthcare, Milwaukee, USA). The test was ended when the patient failed to maintain a pedal frequency of at least 60 rpm or indicated that they were exhausted. The subjects were verbally encouraged during exercise testing to achieve a maximal exercise test, based on a respiratory gas exchange ratio (RER) ≥ 1.05 and subjective opinion of an experienced tester, based on subjective features (fatigue, laboured and fast breathing, leg muscle pain). Perceived exertion of the respiratory system and muscles was evaluated using the Borg scale, to assess the intensity level the patient experienced. After stopping the test, 2 minutes of recovery was tracked while subjects continued to cycle unloaded at 50 rpm. Parameters were assessed at rest, defined by the lowest HR during the one minute of rest on the bike, and at peak exercise. Peak exercise was defined by the highest VO_2 value the subject could achieve before starting recovery. In recovery, heart rate recovery ($\text{HRR} (\Delta\text{HR}_{\text{peak}}-\text{HR})$) was assessed at 30, 60, 90 and 120 seconds (where data was available at this time point). Heart rate reserve ($\text{HR}_{\text{reserve}}$) was calculated by extracting HR at rest (HR_{rest}) from HR at peak (HR_{peak}). Chronotropic index (CRI) was calculated using the following formula: $(\text{HR at peak exercise} - \text{resting HR})/(\text{predicted HR}_{\text{peak}} - \text{resting HR})$ (58). Predicted HR_{peak} was estimated using the formula proposed by Tanaka et al. (2001) $(208 - 0.7 \times \text{age})$ (59, 60).

The oxygen uptake efficiency slope (OUES) was calculated using all exercise data (from start up to recovery) by a linear least square regression of VO_2 on the logarithmic of VE (61). The first ventilatory threshold (VT1) was estimated from the slope of HR against VO_2 , at the point that HR starts to increase disproportionately compared to VO_2 . The second ventilatory threshold (VT2) was determined on the VE/VCO_2 slope, at the point where VE starts to increase more than VCO_2 does.

Venous blood was drawn at two time points, at rest, just before the CPET, and at peak exercise. In these samples, potassium and lactate concentrations were determined. Venous blood was collected in lithium heparin tubes (Vacutainer®, BD, New Jersey, USA) and potassium concentration was established using Cobas ®8000 modular analyser (F. Hoffmann-La Roche Ltd., Basel, Switzerland). Lactate concentrations in venous blood were determined using Cobas Accutrend®Plus from F. Hoffmann-La Roche Ltd. (Basel, Switzerland).

The electrocardiogram was analysed using 50 second ECG-traces at rest and at peak exercise, and averaging the measurements. PR-interval, RR-interval, p-top width, QRS-complex width, Sokolow-index and QT-interval were measured using CardioSoft software from GE Healthcare (Milwaukee, USA) (see Appendix 3). Corrected QT-interval (QTc) was then calculated using Bazett's Formula ($QT \text{ Interval} / \sqrt{RR\text{-interval}}$).

2.6. Statistics

Continuous data was checked for normality within both groups. Afterwards, continuous parameters were compared between OG and LG using either a Mann-Whitney U test (in case of non-parametric data) or an independent samples t-test (normally distributed data). Chi-Square analysis was used to compare discrete data (Tanner stage, sex). P-values of ≤ 0.05 were considered significant (2-tailed). Correlations were made using Spearman's rank correlation coefficient for non-parametric data. Multivariate linear regression was used to determine the relationship between CPET parameters that were significantly different between groups and other significantly altered parameters, grouped as metabolic parameters, cardiovascular parameters and electrocardiography parameters. All statistical analyses were performed using IBM® SPSS® Statistics for Macintosh, Version 24.0 (IBM Corp, Armonk, NY).

3. Results

3.1. Participant characteristics

Characteristics of the participants are shown in Table 1. To summarize, the study population consisted of 48 adolescent males and females divided into the OG (n=24) and the LG (n=24). Groups were age matched (age OG: 13.4±1.1 years, age LG: 13.9±1.5 years, p=0.32) and male and female subjects were proportioned equally (p=0.77). Physical maturity levels were uniformly distributed among groups (p=0.52). Body length was not different between two groups (OG: 166.4±8.7 cm, LG: 167.3±9.1 cm, p=0.51). In line with the study design, mean weight and BMI were respectively 61% and 64% higher in the OG (weight OG: 89.3±16.3 kg, weight LG: 55.3±11.4 kg, p=<0.01; BMI OG: 32.1±4.3 kg/m², BMI LG: 19.6±2.5 kg/m², p=<0.01). WC and HC, as well as WHR were higher in the oG (WC OG: 104.4±13.4 cm, WC LG 67.8±6.5 cm, p=<0.0; HC OG 105.0±8.6 cm, HC LG 78.9±8.9 cm, p=<0.01; WHR OG: 0.99±0.07, WHR LG 0.87±0.11, p=<0.01). BIA showed fat percentage to be 161% higher in OG, SFC calculations indicates a 181% higher fat percentage in OG. (BIA Fat% OG: 36.7±6.9, BIA Fat% LG 14.0±7.9, p=<0.01; SFC Fat% OG: 48.9±11.1, SFC Fat% LG 17.4±5.9, p= <0.01). Physical activity level was higher in the LG (PAQ-A score OG: 2.1±0.6, PAQ-A score LG 2.6±0.5, p=0.02).

Table 1: Patient characteristics

	Obese subjects (n=24)		Lean subjects (n=24)		p-value
	Mean	± SD	Mean	± SD	
Age (Years)	13.4	± 1.1	13.9	± 1.5	0.32
Sex					0.77
Male (n)	13		14		
Female (n)	11		10		
Weight (kg)	89.3	± 16.3	55.3	± 11.4	<0.01
Length (cm)	166.4	± 8.7	167.3	± 9.1	0.51
BMI (kg/m ²)	32.1	± 4.3	19.6	± 2.5	<0.01
WC (cm)	104.4	± 13.4	67.8	± 6.5	<0.01
HC (cm)	105.0	± 8.6	78.9	± 8.9	<0.01
WHR	0.99	± 0.07	0.87	± 0.11	<0.01
% Fat mass					<0.01
BIA	36.7	± 6.9	14.0	± 7.9	<0.01
SFC	48.9	± 11.1	17.4	± 5.9	<0.01
Tanner stage					0.52
I (n)	2		2		
II (n)	0		1		
III (n)	6		4		
IV (n)	2		6		
V (n)	11		11		
PAQ-A score	2.1	± 0.6	2.6	± 0.5	0.02

Abbreviations: BMI, body mass index; WC, waist circumference; HC, hip circumference; WHR, waist hip ratio; BIA, bioelectrical impedance analysis; SFC, skinfold calliper

3.2. Cardiopulmonary testing parameters

Table 2 lists the parameters of the CPET from OG and LG, compared between groups. Blood lactate concentration at rest was different between the two groups (OG: 1.3 ± 0.7 , LG: 1.7 ± 0.7 , $p=0.02$). Systolic and diastolic blood pressure were higher in the obese patients (Psys OG: 124.9 ± 12.1 mmHg, Psys LG: 113.7 ± 10.3 mmHg, $p < 0.01$; Pdias OG: 77.2 ± 9.8 mmHg, Pdias LG: 69.1 ± 7.5 mmHg, $p < 0.01$).

At peak exercise only one parameter was significantly different between groups: cycling power output. With a mean of 158.8 W (SD: 36.2 W), peak cycling power output in the OG was 20% lower than in the LG (190.3 ± 44.5 , $p=0.01$).

Table 2: Cardiopulmonary testing parameters

	Obese subjects (n=24)		Lean subjects (n=24)		p-value
	Mean	± SD	Mean	± SD	
Rest					
VO ₂ (ml/min)	351.9	± 83.7	315.4	± 111.9	0.05
VCO ₂ (ml/min)	315.9	± 80.5	264.8	± 96.2	0.05
VE (l/min)	10.6	± 2.6	9.1	± 2.9	0.07
VE/VO ₂	0.031	± 0.007	0.030	± 0.006	0.53
VE/VCO ₂	0.034	± 0.004	0.035	± 0.004	0.36
Vt (l)	0.66	± 0.27	0.65	± 0.23	0.09
Vd/Vt	14.7	± 4.8	15.8	± 6.2	0.52
BF (breaths/min)	17.4	± 4.4	15.7	± 5.0	0.22
RER	0.90	± 0.12	0.84	± 0.11	0.09
PETO ₂ (mmHg)	112.0	± 6.1	111.1	± 6.8	0.65
PETCO ₂ (mmHg)	34.8	± 3.1	33.7	± 3.4	0.25
VO ₂ HR (ml/beat)	3.9	± 1.0	4.4	± 1.7	0.30
Psys (mmHg)	124.9	± 12.1	113.7	± 10.3	<0.01
Pdia (mmHg)	77.2	± 9.8	69.1	± 7.5	<0.01
Borg Legs	9.9	± 1.8	8.6	± 2.0	0.02
Borg Lungs	9.3	± 2.5	8.3	± 1.4	0.22
Lactate Rest (mmol/l)	1.3	± 0.7	1.7	± 0.7	0.02
K Rest (mmol/l)	3.8	± 0.2	4.0	± 0.3	0.05
Peak exercise					
VO ₂ (ml/min)	2059.1	± 454.7	2251.4	± 553.5	0.21
VCO ₂ (ml/min)	2525.0	± 595.0	2782.0	± 714.1	0.14
VE (l/min)	75.6	± 19.8	84.5	± 22.3	0.15
VE/VO ₂	0.037	± 0.005	0.038	± 0.006	0.59
VE/VCO ₂	0.030	± 0.003	0.031	± 0.004	0.41
Vt (l)	1.9	± 0.5	1.9	± 0.5	0.83
Vd/Vt	18.8	± 2.8	18.4	± 2.8	0.61
BF (breaths/min)	41.2	± 8.1	45.3	± 9.2	0.11
RER	1.23	± 0.07	1.23	± 0.07	0.69
PETO ₂ (mmHg)	114.1	± 18.4	119.3	± 5.1	0.17
PETCO ₂ (mmHg)	40.9	± 18.6	35.8	± 4.3	0.07
VO ₂ HR (ml/beat)	11.1	± 2.2	12.1	± 2.9	0.16
Borg Legs	15.8	± 3.3	16.5	± 2.0	0.64
Borg Lungs	16.3	± 2.2	15.6	± 1.9	0.55
Cycling power output (W)	158.8	± 36.2	190.3	± 44.5	0.01
Lactate Peak (mmol/l)	4.9	± 2.5	4.5	± 1.4	0.82
K Peak (mmol/l)	4.2	± 0.6	4.4	± 0.5	0.43

Abbreviations: VO₂, oxygen uptake; VCO₂, carbon dioxide output; VE, expired volume; VE/VO₂, ventilatory equivalents for O₂; VE/VCO₂, ventilatory equivalents for CO₂; Vt, tidal volume; Vd/Vt, Physiological dead-space-tidal volume ratio; BF, breathing frequency; RER, respiratory exchange ratio; PETO₂, end-tidal O₂ tension; PETCO₂, end-tidal CO₂ tension; VO₂/HR, oxygen pulse; Psys, systolic blood pressure; Pdia, diastolic blood pressure; K, potassium;

The VT1 was reached at a higher percentage of VO₂peak in obese subjects (OG: 52.9±6.1%, LG:47.7±6.8%, p=0.01). VT1 however was not different between groups (OG:1072.5±185.2 ml/min; LG: 1072.5±296. p=1.00). The VT2 and the percentage of the VO₂peak at which it was attained were not different between groups (Table 3).

Table 3: Ventilatory efficiency parameters during CPET

	Obese subjects (n=24)		Lean subjects (n=24)		p-value
	Mean	± SD	Mean	± SD	
OUES	2195.8	± 501.2	2229.4	± 501.5	0.82
VT1(ml/min)	1072.5	± 185.2	1072.5	± 296.2	1.00
VT2(ml/min)	1565.4	± 379.4	1802.9	± 502.0	0.12
VT1 (% of VO ₂ peak)	52.9	± 6.1	47.7	± 6.8	0.01
VT2 (% of VO ₂ peak)	76.4	± 9.9	80.0	± 9.1	0.19

Abbreviations: OUES, oxygen uptake efficiency slope (VE/VCO₂ slope); VT1, first ventilatory threshold; VT2, second ventilatory threshold

Resting HR was elevated in OG with 79.9 bpm compared to 64.7 bpm in LG (SD OG: 13.3, SD LG: 10.7, p=<0.01). Peak HR was not lowered in the OG (OG: 185.7±11.5bpm; LG: 185.8±9.6 bpm, p-value 0.97). Chronotropic responses between groups were not different, as indicated by CRI which was similar in both groups (OG: 0.86±0.09; LG: 0.90±0.07; p=0.74). HRreserve however was higher in lean subjects compared to their obese counterparts (OG: 106.4±17.8 bpm, LG:126.5±25.6, p=<0.01). During recovery from peak exercise, heart rate decline in the OG was delayed compared to the LG, as demonstrated by HRR at 30, 60 and 120 sec which are all lowered in obese subjects (HRR30s OG: 9.8±7.2, HRR30s LG: 16.5±10.1, p=0.01; HRR60s OG: 22.5±10.8, HRR60s LG: 31.4±14, p=0.02; HRR120s OG: 35.4±11.8, HRR120s LG: 46.1±11.5, p=0.01)(Table 4). This is visualised in Figure 3.

Table 4: Heart rate responses

	Obese subjects (n=24)		Lean subjects (n=24)		p-value
	Mean	± SD	Mean	± SD	
HRrest (bpm)	79.9	± 13.3	64.7	± 10.7	<0.01
HRpeak (bpm)	185.7	± 11.5	185.8	± 9.6	0.97
CRI	0.86	± 0.09	0.90	± 0.07	0.74
HRreserve	106.4	± 17.8	126.5	± 25.6	<0.01
HRR					
30 s	9.8	± 7.2	16.5	± 10.1	0.01
60 s	22.5	± 10.8	31.4	± 14	0.02
120 s	35.4	± 11.8	46.1	± 11.5	0.01

Abbreviations: HR, heart rate; HRreserve, heart rate reserve (Δ HRpeak-HRrest); HRR, heart rate recovery (Δ HRpeak - HR at 30, 60, 120s)

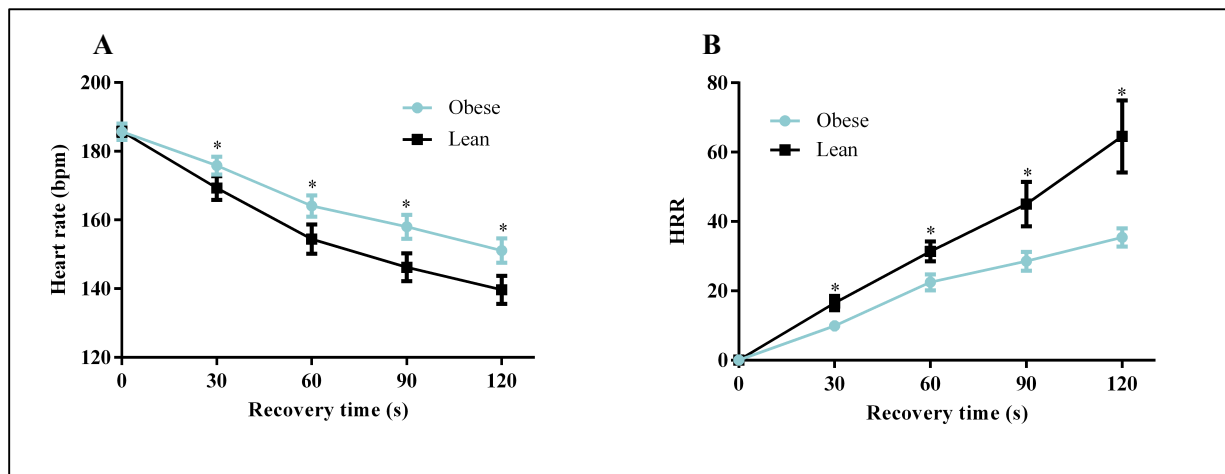


Figure 3: Heart rate changes during recovery from peak exercise in obese and lean subjects. (A) The decline in heart rate over time, starting at HRpeak. (B) Course of HRR. * p-value<0.05 between groups

Abbreviations: HRR, heart rate recovery

3.3. Blood parameters

Markers of glycaemic control are listed in Table 5. Fasting blood glucose was not altered between groups (OG: 88.9 ± 6.1 mg/dl, LG: 86.1 ± 6.6 , $p=0.14$), blood insulin concentration was higher in the OG (OG: 176.7 ± 117.3 pmol/l, LG: 73.1 ± 38.4 pmol/l, $p < 0.01$). HOMA-IR, also had significant different values between groups (OG: 5.6 ± 3.9 , LG: 2.2 ± 1.3 , $p < 0.01$). Mean blood leptin concentration in the obese subjects was higher than in lean subjects (OG: 25.4 ± 16.9 ng/ml, LG 10.5 ± 5.5 ng/ml, $p < 0.01$).

Liver function was disturbed in obese subjects, as indicated by elevated blood ALT and GGT concentrations (ALT OG: 29.8 ± 16.8 IU/L, ALT LG: 16.6 ± 7.3 IU/L, $p < 0.01$; GGT OG: 19.4 ± 8.1 IU/L, GGT LG: 13.2 ± 4.2 IU/L, $p < 0.01$).

Blood HDL concentrations were reduced and blood LDL and triglyceride concentrations were increased (HDL OG: 46.2 ± 11.4 mg/dl, LG: 62.3 ± 11.7 mg/dl, $p < 0.01$; LDL OG: 93.5 ± 26.5 mg/dl, LG 76.0 ± 22.2 mg/dl, $p=0.02$; triglycerides OG: 105.0 ± 63.7 mg/dl, LG 68.5 ± 32.1 mg/dl, $p=0.03$). In obese subjects blood CRP concentration was $4.8 (\pm 7.5)$ mg/dl and in lean subjects $0.6 (\pm 2.0)$ mg/dl and thus lower ($p < 0.01$) (Table 5).

The remaining biochemical and haematological blood values tested did not show many dissimilarities among the two groups. Blood iron concentrations in obese subjects were reduced in the OG (OG: 81.8 ± 34.8 μ g/dl, LG: 108.8 ± 39.4 μ g/dl, $p=0.02$). Haemoglobin and haematocrit concentrations were

both lowered in obese subjects (Hb OG: 13.4±0.9 g/dl, LG: 14.1±1.1 g/dl, p=0.02; Ht OG: 38.7±2.3%, 39.4±7.9%, p=0.01) (Table 5).

Table 5: Blood parameters

	Obese subjects (n=24)		Lean subjects (n=24)		p-value
	Mean	± SD	Mean	± SD	
Glycemic control					
Glucose (mg/dl)	88.9	± 6.1	86.1	± 6.6	0.14
Insulin (pmol/l)	176.7	± 117.3	73.1	± 38.4	<0.01
HbA1C (mg/dl)	5.3	± 0.3	5.2	± 0.2	0.13
HOMA-IR	5.6	± 3.9	2.2	± 1.3	<0.01
Leptin (ng/ml)	25.4	± 16.9	10.5	± 5.5	<0.01
Liver function markers					
AST (IU/L)	23.7	± 7.3	24.4	± 8.7	0.90
ALT (IU/L)	29.8	± 16.8	16.6	± 7.3	<0.01
GGT (IU/L)	19.4	± 8.1	13.2	± 4.2	<0.01
ALP (IU/L)	188.0	± 91.6	200.1	± 120.3	0.91
Uric acid (µmol/L)	7.4	± 8.9	5.1	± 1.1	0.45
Atherosclerosis risk factors					
Total cholesterol (mg/dl)	158.4	± 33.5	152.0	± 22.2	0.74
HDL (mg/dl)	46.2	± 11.4	62.3	± 11.7	<0.01
LDL (mg/dl)	93.5	± 26.5	76.0	± 22.2	0.02
Triglycerides (mg/dl)	105.0	± 63.7	68.5	± 32.1	0.03
CRP (mg/dl)	4.8	± 7.5	0.6	± 2.0	<0.01
Other bloodparameters					
Iron (µg/dl)	81.8	± 34.8	108.8	± 39.4	0.02
Calcium (mmol/l)	2.3	± 0.3	2.4	± 0.1	0.20
ferritin (ng/ml)	58.5	± 31.8	48.6	± 27.4	0.26
TSH (IU/ml)	2.7	± 1.0	2.2	± 0.8	0.07
free T4 (pmol/L)	14.1	± 3.1	14.2	± 1.8	0.56
cortisol (ng/ml)	9.8	± 6.2	7.8	± 3.7	0.34
Hb (g/dl)	13.4	± 0.9	14.1	± 1.1	0.02
Ht (%)	38.7	± 2.3	39.4	± 7.9	0.01
Leukocytes (cells/µl)	7.0	± 1.4	8.7	± 8.4	0.15

Abbreviations: HbA1C, glycosylated haemoglobin; AST, aspartate aminotransferase; GGT, gamma-glutamyl transpeptidase; ALT, alanine aminotransferase; ALP, alkaline phosphatase; HDL, high-density lipoprotein; LDL, low-density lipoprotein; CRP, c-reactive protein; TSH, thyroid stimulating hormone; T4, thyroxine; Hb, Haemoglobin; Ht, Haematocrit

3.4. Metabolic risk factor

Obese subjects had a significantly higher MRF, with a mean of 0.63(\pm 0.63), compared to a mean MRF in the lean subjects of -0,47 (\pm 0.28) (p < 0.01).

3.5. Electrocardiogram parameters

At rest, RR-intervals of the obese subjects were smaller than those of the lean subjects (OG: 641.7 \pm 319.0, LG: 802.8 \pm 172.8, p < 0.01) (Table 6). Both at rest and at peak exercise the PR-interval was longer in the LG, both times by around 10% (Rest PR-interval OG: 135.9 \pm 398.8 ms, LG: 149.3 \pm 16.3 ms, p = 0.03, Peak PR-interval OG: 94.5 \pm 18.3, LG: 105.0 \pm 14.4, p = 0.03). Sokolow-index was also significantly different at both time points. At rest, it was 3.1(\pm 1.7) mV in the OG and 4.0 (\pm 1.2) mV in the LG (p < 0.01). At peak exercise the same relationship is seen, again Sokolow-index in the OG was smaller than in de LG (OG: 2.6 \pm 1.0mV, LG: 3.7 \pm 1.1, p < 0.01).

Table 6: ECG-parameters

	Obese subjects (n=24)		Lean subjects (n=24)		p-value
	Mean	\pm SD	Mean	\pm SD	
Rest					
RR-interval (ms)	641.7	\pm 319.0	802.8	\pm 172.8	<0.01
PR-interval (ms)	135.9	\pm 398.8	149.3	\pm 16.3	0.03
P-top-width (ms)	86.0	\pm 13.7	88.8	\pm 13.1	0.47
QRS-width (ms)	87.4	\pm 10.5	82.5	\pm 10.7	0.12
Sokolow index (mV)	3.1	\pm 1.7	4.0	\pm 1.2	<0.01
QT-interval (ms)	319.0	\pm 53.0	343.6	\pm 50.3	0.05
QTc (ms)	398.8	\pm 60.3	389.7	\pm 67.1	0.15
Peak Exercise					
RR-interval (ms)	321.3	\pm 20.8	324.4	\pm 16.6	0.57
PR-interval (ms)	94.5	\pm 18.3	105.0	\pm 14.4	0.03
P-top-width (ms)	71.0	\pm 13.6	70.7	\pm 13.3	0.95
QRS-width (ms)	79.0	\pm 9.6	75.7	\pm 10.0	0.24
Sokolow index (mV)	2.6	\pm 1.0	3.7	\pm 1.1	<0.01
QT-interval (ms)	222.2	\pm 35.5	222.8	\pm 13.2	0.37
QTc (ms)	392.2	\pm 61.1	391.0	\pm 21.2	0.14

QTc : QT interval corrected for heart rate

3.6. Ultrasound parameters

CIMT in lean subjects was on average 0.04 mm thicker than CIMT in obese subjects (OG: 0.50±0.05 mm, LG: 0.54±0.06 mm, p=0.02).

LV septum was widened in obese subjects with an average of 0.9 mm compared to lean subjects (OG: 8.0±0.8 mm, LG: 7.1±1.0 mm, p=<0.01) (Table 7). The diameter of the LA was also broader in the OG (OG: 34.1±4.5 mm, LG: 29.0±3.9 mm, p=<0.01). Although no significant differences were seen in other parameter, trends are seen in velocity of the mitral A wave and the E/E'-ratio. Both are higher in the obese subjects. An algorithm was used to the rise in LAP, however in both groups no subjects classified as LAP↑.

Table 7: Ultrasound parameters

	Obese subjects (n=24)		Lean subjects (n=24)		p-value
	Mean	± SD	Mean	± SD	
CIMT (mm)	0.50	± 0.05	0.54	± 0.06	0.02
TTE parameters					
EF (%)	60.9	± 6.0	60.7	± 6.1	0.89
LV septum width (mm)	8.0	± 0.8	7.1	± 1.0	<0.01
LV diameter (mm)	45.4	± 4.0	46.2	± 3.5	0.50
LA diameter (mm)	34.1	± 4.5	29.0	± 3.9	<0.01
E mitral (cm/sec)	83.6	± 16.5	82.3	± 13.9	0.60
A mitral (cm/sec)	52.5	± 13.1	45.9	± 8.0	0.06
mitral E/A ratio	1.7	± 0.4	1.8	± 0.5	0.16
DT (ms)	154.8	± 22.3	145.8	± 26.2	0.22
E/E'	9.2	± 1.9	8.2	± 1.6	0.06
CO (l/min)	5.2	± 0.8	4.9	± 1.1	0.26
LAP ↑ (n)	0		0		1

CIMT, carotid intima media thickness; EF, Ejection fraction; LV, left ventricle; LA, left atrium; E mitral, transmitral peak early diastolic velocity; A mitral, transmitral peak late diastolic velocity; DT, deceleration time; E', Early diastolic mitral annular velocity; CO, Cardiac output; LAP, left atrial pressure

3.7. Association between metabolic risk factor and heart rate responses

Since metabolic risk factor and HRreserve, HRR30s, HRR60s and HRR120s proved changed in obese the OG, the association between these factors was further investigated. The associations were first evaluated using linear regression and concomitant correlation (Figure 4). All three HRR parameters were significantly correlated with metabolic risk (MRF). HRreserve was negatively correlated with a correlation coefficient (r) of -0.297 ($p=0.05$), but the correlation was not significant. The correlations between HRR and MRF differ between time points, although all relationships are significantly negative. At 30 seconds, HRR is correlated with an r of -0.393 ($p=0.006$). At 60 seconds the relation becomes even stronger, and the correlation coefficient is -0.414 ($p=0.004$). After another minute of recovery, the relationship with MRF returns to the same value as at 30 seconds ($r: -0.392$, $p=0.014$).

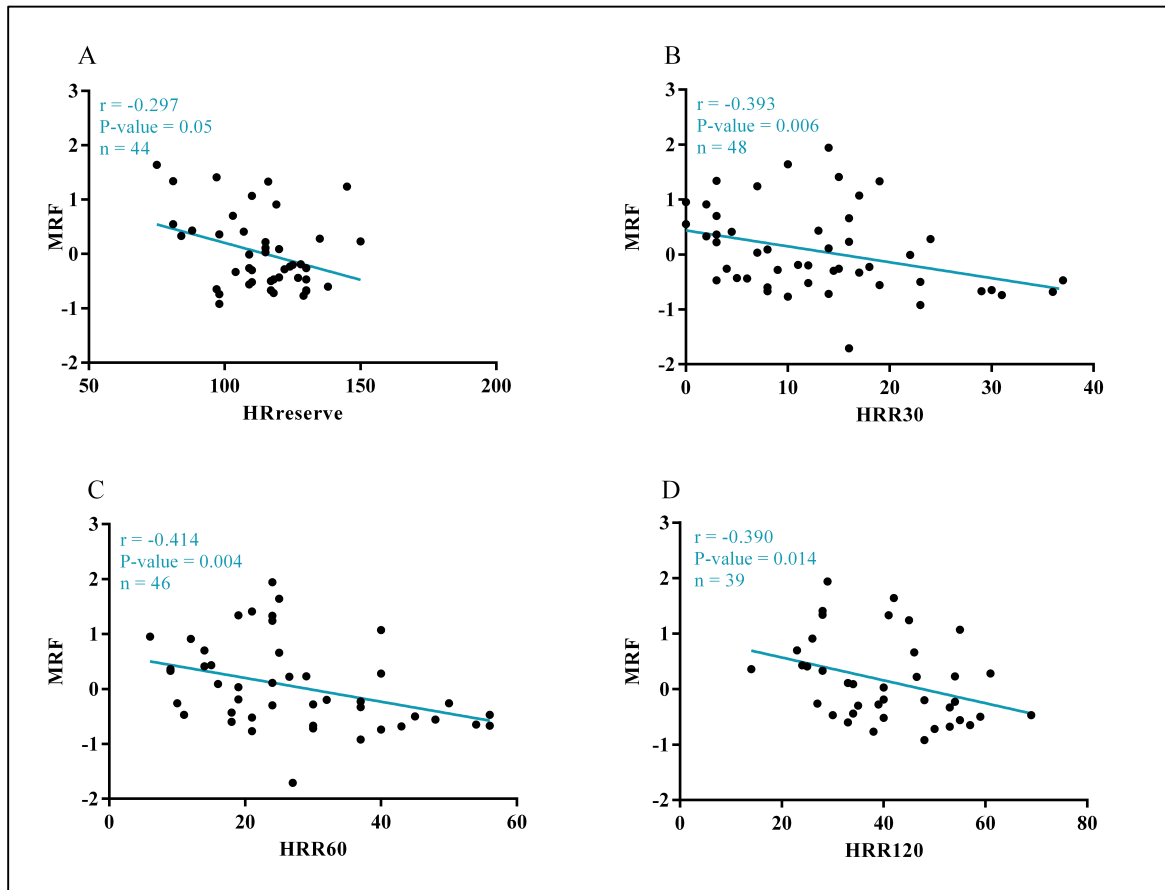


Figure 4: Correlations between MRF and (A) HRreserve, (B), HRR30s, (C) HRR60s, (D) HRR120s.

Abbreviations: MRF, metabolic risk factor; HRreserve, heart rate reserve (Δ HR_{peak}-HR_{rest}); HRR, heart rate recovery (Δ HR_{peak} - HR at 30, 60, 120s)

Next, an adjusted linear model was used to associate MRF with each of the four autonomic heart function parameters. For each parameter, the best fitted model was chosen (according to adjusted R^2), initially adjusting the models for age, length, tanner stage and sex (Table 8). Even after adjusting, MRF stays a significant negative predictor for HRreserve, HRR30sec, HRR60sec, and HRR120sec. The model for MRF and HRreserve had a coefficient of determination (adjusted R^2) of 0.253 and the standardized coefficient β (SC β) for MRF in this model was -0.408 ($p=0.009$). MRF had an SC β of -0.33($p=0.017$) in the model for HRR30 with an adjusted R^2 of 0.295. The corrected model between HRR60 gives MRF an SC β of -0.381($p=0.006$), the goodness of fit in this model is represented by adjusted R^2 of 0.301. The last model, for MRF and HRR120, has an adjusted R^2 of 0.139 and gives MRF an SC β of -0.466.

3.8. Multivariate models for heart rate responses

Multivariate models were created for autonomic heart function parameters HRreserve, HRR30s, HRR60s and HRR120s and (i) metabolic parameters, (ii) cardiovascular parameters and (iii) ECG parameters (Table 8). For all predictors in the models, correlations were evaluated, results can be found in Appendix 4.

3.8.1. Metabolic parameter models for heart rate responses

HRreserve was negatively associated with WHR (adjusted R^2 : 0.243, SC β : -0.298, $p<0.01$). HRR30sec was significantly associated with blood leptin and ALP concentrations (adjusted R^2 : 0.482). The relationships were different in direction, as blood leptin had a negative association with an SC β of -0.454($p<0.01$) and blood ALP was positively associated (SC β : 0.406, $p<0.01$). The model compiled for HRR60s contained blood ALP, triglycerides, iron and Hb concentration (adjusted R^2 : 0.540). Triglyceride concentration had a negative relationship (SC β : -0.394, $p<0.01$), the rest of the predictors were positively related to HRR60s (SC β ALP: 0.405, $p<0.01$; SC β iron: 0.229, $p=0.05$; SC β Hb: 0.297, $p=0.01$). HRR at 120s is predicted with an adjusted R^2 of 0.515 by blood CRP, triglyceride and ferritin concentrations. CRP had an SC β of -0.303 ($p=0.013$), triglyceride concentration had an SC β of -0.275 ($p=0.01$), and lastly SC β for ferritin is 0.5472 ($p<0.01$).

3.8.2. Cardiovascular parameter models for heart rate responses

HRreserve was associated with $\dot{V}CO_2$ at peak exercise (SC β : 0.404, $p < 0.01$), adjusted R^2 for this model is 0.142. The model assembled for HRR30s contained only mitral A-wave velocity as a predictor and had an adjusted R^2 of 0.295 (SC β A mitral -0.558, $p = 0.01$). HRR at 60sec was positively correlated with peak $\dot{V}O_2/HR$ (SC β : 0.369, $p = 0.02$) and negatively associated with mitral A-wave velocity (SC β : -0.421, $p = 0.01$). This model had a goodness of fit indicated with an adjusted R^2 of 0.314. The model for HRR120s predictive with an adjusted R^2 of 0.076 compiling only Mitral E/A ratio (SC β : 0.314, $p = 0.04$).

3.8.3. Electrophysiological parameter models for heart rate responses

Again, no significant predictors could be found for HRreserve and consequently a model was not generated. HRR at 30sec was correlated with peak PR-interval (adjusted R^2 : 0.246) (SC β : 0.337, $p = 0.02$). HRR60s was significantly associated with PR-interval both at rest and at peak exercise (adjusted R^2 : 0.390). SC β for resting PR-interval is 0.278 ($p = 0.04$) and for peak PR-interval it is 0.369 ($p = 0.01$). The model composed for HRR120s contained only resting PR-interval (SC β : 0.490, $p < 0.01$) and the model was predictive with an adjusted R^2 of 0.221.

Table 8: Multivariate adjusted models for heart rate responses

	HRreserve			HRR 30			HRR60			HRR120		
	Adj. R ²	SC β	p-value	Adj. R ²	SC β	p-value	Adj. R ²	SC β	p-value	Adj. R ²	SC β	p-value
<u>Metabolic Risk model</u>	0.253 ^a			0.299 ^a			0.301 ^a			0.139 ^b		
Metabolic Risk		-0.408	0.01		-0.330	0.02		-0.358	0.01		-0.466	<0.01
<u>Metabolic parameters model</u>	0.243 ^c			0.482			0.540			0.515		
WHR		-0.298	0.037									
Leptin (ng/ml)					-0.454	<0.01						
ALP (IU/L)					0.406	<0.01		0.405	<0.01			
Triglycerides (mg/dl)								-0.394	<0.01		-0.334	<0.01
CRP (mg/dl)											-0.303	0.013
Iron (μg/dl)								0.229	0.05			
ferritin (ng/ml)											0.472	<0.01
Hb (g/dl)								0.297	0.01			
<u>Cardiovascular parameters model</u>	0.142			0.295			0.314			0.076		
Peak VCO ₂ (ml/min)		0.404	0.008									
Peak VO ₂ /HR								0.369	0.02			
A mitral (cm/sec)					-0.558	0.01		-0.421	0.01			
Mitral E/A ratio											0.314	0.04
<u>ECG parameters model</u>				0.246 ^a			0.390 ^a			0.221		
Rest PR-interval (ms)								0.278	0.04		0.490	<0.01
Peak PR-interval (ms)					0.337	0.02		0.369	<0.01			

Abbreviations: Adj., adjusted; HRreserve, heart rate reserve (Δ HR_{peak}-HR_{rest}); HRR, heart rate recovery (Δ HR_{peak} - HR); SCβ, standardized coefficient β; AST, aspartate aminotransferase; GGT, gamma-glutamyl transpeptidase; ALP, alkaline phosphatase; CRP, c-reactive protein; Hb, Haemoglobin; VCO₂, carbon dioxide output; VO₂/HR, oxygen pulse; E mitral, transmitral peak early diastolic velocity; A mitral, transmitral peak late diastolic velocity

^a, adjusted for sex

^b, adjusted for age, length, tanner stage and sex

^c, adjusted for body length

4. Discussion

The aim of this clinical study was to examine whether HR responses during cardiopulmonary exercise testing may be an early indicator for cardiovascular anomalies in obese adolescents. The main findings of our study are that heart rate reserve as well as heart rate recovery are significantly altered in obese subjects. Moreover, significant associations are found between these parameters and parameters indicative of cardiovascular function.

4.1. Markers for cardiovascular abnormalities

Comparing the OG and LG, many important different changes are visible, which are associated with cardiovascular outcomes. First, evidence of the MetS is seen in the obese population. This cluster of four risk factors, leading to increased risk of cardiovascular disease, can be observed the OG (62). Psys and Pdia are elevated (hypertension), triglyceride concentration in the blood is augmented (hypertriglyceridemia), the concentration of blood HDL cholesterol is lower and blood insulin concentration is higher (hyperinsulinemia). MRF confirms this finding: obese subjects have an overall much higher score. This factor is based on triglyceride/HDL ratio, insulin concentration, MAP (a measure blood pressure) and additionally, WC (52). WC had been indicated as a marker of cardiovascular health, and WHR, another parameter that was substantially heightened in obese subjects, is often used in cardiometabolic risk assessment in children (63, 64).

In the results from the ECG analyses, differences in resting RR-interval are seen, however, RR-interval in this case is only a proxy for HR, it represents the time between two heart depolarisations and consequent heart beats (65). HR_{rest} is significantly higher in the obese groups and thus follows that the RR-interval is shorter. PR-interval again is shortened in the OG, though, this time both at rest and at peak exercise. At rest, the shorter PR-interval could be associated with the faster HR, but since the HR_{peak} is the same in both groups, the PR-interval difference at peak exercise cannot be explained in this way. What is more, other studies have found opposite results, that PR-interval is longer in obese adolescents (66) and that in adults it prolonged PR interval has been associated with adverse cardiovascular outcomes (67).

Sokolow-index was significantly higher both at rest and at peak exercise in the LG. A high Sokolow-index can indicate left ventricular hypertrophy (68). Sokolow-index, however, is based upon the signal transduction of the electrical activity of the heart. In obese individuals, the signal must travel through a

larger amount of tissue, and thus some of the signal is lost (69). In this way, the higher Sokolow-index in the LG can be rationalised and it does not imply any effects on cardiovascular health.

CIMT in lean subjects is significantly higher in lean subjects, this is contradictory to other findings (70, 71). A possible clarification for this finding is the relationship between androgen concentrations and CIMT, in which higher androgen concentrations are associated with thicker carotid intima media (71, 72). However, androgens were not tested for in our study population, so this remains speculation. Nonetheless, lower concentrations of testosterone have been reported in obese adolescents (73).

In echocardiography, a larger left ventricular septum width is seen in obese subjects. LV septum width can be a predictor for left ventricular hypertrophy, and this has been shown to be present in obese adolescent, possibly due to the effects of high blood pressure (74). Left atrial diameter, an important risk factor for atrial fibrillation in adults (75), is also enlarged in the OG. No other parameters were significantly different between the two groups, but a trend is seen in E/e' ratio, which is higher in obese subjects and is a measure of left ventricular diastolic function. A higher ratio, in this case, can predict a worse outcome, it indicates reduced passive filling capacity of the left ventricle during diastole (76).

4.2. Cardiopulmonary testing outcomes

Our data suggests that peak cycling power output is significantly lowered in obese subjects. Nadeau (2009) and Mendelson (2012) confirm these findings (29, 30). HR_{peak}, however, was not different between groups in our population, which is contradictory to some other findings (29, 31), but again, other studies find the same (30). Peak cycling power output is dependent on the interconnection between intrinsic characteristics of the muscle tissue (amount of tissue, efficiency of energy metabolism,...), the cardiovascular function (efficient distribution of oxygen) and ability of the respiratory system to extract oxygen from the air and elimination of carbon dioxide from the bloodstream (37). Since VO₂ peak is the same in both groups, metabolic demand must be greater in the OG, for the same workload. A few mechanisms can explain this phenomenon. Obese subjects experience a higher demand on muscles due to the increased weight (77) and ventilatory work requires extra metabolic power to compensate for the pressure that is put on the chest by the increase in weight (78). Moreover, obese adults show an increased proportion of type IIb- muscle fibers. This type of fibers, which are more glycolytic, is less efficient in metabolism and is less resistant to fatigue (79).

During recovery from exercise, HRR is significantly slowed in obese subjects. Also, HR_{reserve} is significantly lowered in the OG. This difference in HR reserve, which is defined as the peak HR minus the resting HR, is the effect of the higher HR_{rest} in the obese patients since the HR_{peak} is not different

between groups. Other studies have also seen this occurrence of a lower resting HR in obese adolescents (80, 81). A possible clarification for this higher resting HR is dysregulation of the autonomic nervous system (ANS). The sympathetic nervous system (SNS) has been shown to be over-active, while parasympathetic nervous system (PNS) activity is reduced in obese adolescents (33, 82). However, other studies have found an opposite effect and found an under-activation of the SNS as well as the PNS (83, 84). Our findings confirm the first outcomes since a higher resting HR is the symptom of over-activation of the SNS and possible reduced PNS activity. The finding that HRR recovery is slowed in the obese subjects confirms this and was established by other studies that find the same (32, 85). HRR was calculated as the difference between HR_{peak} and the HR 30, 60 and 120 seconds after the cessation of the maximal exercise test. HRR can be used as a measure of ANS dysfunction. The short term HRR, at 30 and 60 seconds at could be considered the consequence of cardiac parasympathetic activity, while HRR at 120 seconds is related to withdrawal of sympathetic activity and to the clearance of adrenal influences (86). Furthermore, slowed HRR is associated with higher mortality risk (87, 88).

The correlations between MRF and these autonomic heart function parameters are often seen in literature. Many studies speak of dysfunctional autonomic function in the presence of metabolic syndrome in obese adolescents, of which MRF is a predictor (89, 90).

In the models with other metabolic parameters for HRR, an association is seen between blood leptin concentration and HRR₃₀. Evidence suggests leptin stimulates sympathetic activity (91), and as leptin resistance and an increase in blood leptin are often seen in obese patients (92), it could be one of the initiating factors of dysfunction of the ANS.

An association with both HRR₃₀ and HRR₆₀ is seen with blood ALP concentration. The higher the concentration, the better the outcome in the form of higher HRR. ALP is mostly formed in the liver and bone tissue and higher concentrations of serum ALP have been associated with MetS (93). However, multiple forms of ALP are found in the body, evidence suggests that intestinal alkaline phosphatase deficiency is linked to development of MetS and T2DM, and that higher concentrations are protective (94). This could explain the link with HRR, although further evaluation is needed.

Triglyceride concentration in the blood is negatively associated with HRR₆₀ and HRR₁₂₀, this can be explained through the mechanisms of MetS, since hypertriglyceridemia is a risk factor included in the MetS (23). Blood CRP, a marker for inflammation, is negatively correlated with HRR₁₂₀, evidence suggest an association between inflammatory pathways and the ANS (95). Lastly in the metabolic parameters models, positive associations were found between blood iron and Hb concentrations and

HRR60 and ferritin and HRR120. Although no evidence exist linking these mechanisms, blood iron, Hb and ferritin are all involved in the transport of oxygen in the blood, and deficiencies can implicate anemia (96). Moreover, anemia has been associated with ASN dysfunction (97). Linking these two mechanisms can explain the associations between HRR and blood iron, Hb and ferritin concentrations.

The current study suggests HRreserve is associated positively with peak $\dot{V}CO_2$. A high peak $\dot{V}CO_2$ suggest a better overall exercise performance (98), possibly this explains the association with better HRreserve outcomes. Peak $\dot{V}O_2/HR$ is also positively associated with HRR 60. A higher oxygen pulse, again, is a sign of improved exercise performance since more oxygen per heart beat is pumped, possibly due to higher stroke volumes (99). Stroke volume during exercise is regulated by the SNS (100), it is thus likely that normal function of the SNS is related to better HRR outcomes.

HRR30 and HRR60 are negatively associated with mitral A-wave velocity. Decreased velocity of the A-wave can point to diastolic dysfunction. Although SNS over-activation has been linked to systolic dysfunction(101), its role in diastolic dysfunction remains to be investigated(102). Mitral E/A ratio is positively associated with HRR120. In normal diastolic function, E-wave velocity is higher than the A-wave velocity, and a higher ratio is thus better. However, a much higher E/A ratio can indicate diastolic dysfunction (103). Conversely, because all our patients were below this boundary of E/A function, this association is positive.

The lasts models include ECG-parameters. They indicated PR-interval at rest and at peak exercise are associated with HRR. It is likely that PR-interval is related more to HR than any abnormalities in electrophysiological function, and that the association is more related to HR. Resting HR confirms this theory, a longer PR-interval would indicate a lower HR and that is a positive predictor for HRR. However, because HRpeak is not different between groups, the correlation with peak PR-interval is not clear. Evidence suggests that, normally, peak PR-intervals decrease with exercise due to withdrawal of parasympathetic tone(104). Since HRR dysregulation implies that PNS activity is reduced, this cannot elucidate the association.

5. Conclusion

Significant alterations in HRreserve and HRR suggest cardiovascular autonomic function is distorted in obese adolescents. HRreserve and HRR are associated with metabolic risk factors seen in obesity and factors indicative of cardiovascular abnormalities. Although the data indicates that HR at peak exercise is not changed in obese adolescents, HRreserve and HRR might be of prognostic value for cardiovascular disease risk in obese adolescents. Further research should be implemented to discover the associations between cardiac autonomic function and adverse cardiac events and the aetiology behind the development of distorted autonomic function in obese adolescents.

6. Limitations

Our study has a few limitations. Although the target sample size was achieved in this study and sufficient power was generated, a larger sample size could provide more sturdy associations between outcomes. Fat percentage was assessed using BIA and SCF, but more valid methods of assessing body composition are available, for example dual-energy X-ray absorptiometry (DXA). An important implication for performance in the CPET could be amount of physical activity. The PAQ-A that was used in this study provides an idea about physical activity, but questionnaires are always subject to bias from the participant. Systematic data-collection using accelerometers could provide more objective and correct information on the physical activity levels.

7. Future perspectives

Heart rate recovery and heart rate reserve in adolescents deserve more attention. Possible longer recovery time with cardiopulmonary evaluation could provide more information still about the heart rate recovery. Echocardiography during exercise testing would also provide more insight into cardiac function during exercise. Heart rate variability (HRV) is also often used to evaluate cardiac autonomic function, using spectrum analysis on the RR-intervals of the ECG could provide an insight into the HRV during exercise.

8. References

1. Obesity and overweight: Fact sheet 2016 [Available from: <http://www.who.int/mediacentre/factsheets/fs311/en/>].
2. WHO. Physical status: the use and interpretation of anthropometry: report of a WHO Expert Committee. Geneva: World Health Organization; 1995.
3. Cole TJ, Lobstein T. Extended international (IOTF) body mass index cut-offs for thinness, overweight and obesity. *Pediatric obesity*. 2012;7(4):284-94.
4. Gisle L, Demarest S. Gezondheidsenquête 2013: Rapport 2: gezondheidsgedrag en leefstijl In: Volksgezondheid WI, editor. 2014.
5. WHO. Population-based approaches to childhood obesity prevention. 2012.
6. Lobstein T, Jackson-Leach R, Moodie ML, Hall KD, Gortmaker SL, Swinburn BA, et al. Child and adolescent obesity: part of a bigger picture. *Lancet (London, England)*. 2015;385(9986):2510-20.
7. Han JC, Lawlor DA, Kimm SYS. Childhood Obesity – 2010: Progress and Challenges. *Lancet (London, England)*. 2010;375(9727):1737-48.
8. Jimenez-Pavon D, Kelly J, Reilly JJ. Associations between objectively measured habitual physical activity and adiposity in children and adolescents: Systematic review. *International journal of pediatric obesity : IJPO : an official journal of the International Association for the Study of Obesity*. 2010;5(1):3-18.
9. Wang Y, Lobstein T. Worldwide trends in childhood overweight and obesity. *International journal of pediatric obesity : IJPO : an official journal of the International Association for the Study of Obesity*. 2006;1(1):11-25.
10. Redinger RN. The Pathophysiology of Obesity and Its Clinical Manifestations. *Gastroenterology & Hepatology*. 2007;3(11):856-63.
11. Niswender KD, Baskin DG, Schwartz MW. Insulin and its evolving partnership with leptin in the hypothalamic control of energy homeostasis. *Trends in endocrinology and metabolism: TEM*. 2004;15(8):362-9.
12. Matsuzawa Y, Funahashi T, Kihara S, Shimomura I. Adiponectin and metabolic syndrome. *Arterioscler Thromb Vasc Biol*. 2004;24(1):29-33.
13. Lafontan M. Fat cells: afferent and efferent messages define new approaches to treat obesity. *Annual review of pharmacology and toxicology*. 2005;45:119-46.
14. Gonzalez N, Moreno-Villegas Z, Gonzalez-Bris A, Egido J, Lorenzo O. Regulation of visceral and epicardial adipose tissue for preventing cardiovascular injuries associated to obesity and diabetes. *Cardiovascular diabetology*. 2017;16(1):44.
15. Kawanami D, Maemura K, Takeda N, Harada T, Nojiri T, Imai Y, et al. Direct reciprocal effects of resistin and adiponectin on vascular endothelial cells: a new insight into adipocytokine-endothelial cell interactions. *Biochemical and biophysical research communications*. 2004;314(2):415-9.
16. Neef M, Weise S, Adler M, Sergeev E, Dittrich K, Korner A, et al. Health impact in children and adolescents. *Best Pract Res Clin Endocrinol Metab*. 2013;27(2):229-38.
17. Ebbeling CB, Pawlak DB, Ludwig DS. Childhood obesity: public-health crisis, common sense cure. *The Lancet*. 2002;360(9331):473-82.
18. Chinetti G, Fruchart JC, Staels B. Peroxisome proliferator-activated receptors and inflammation: from basic science to clinical applications. *International journal of obesity and related metabolic disorders : journal of the International Association for the Study of Obesity*. 2003;27 Suppl 3:S41-5.
19. Wellen KE, Hotamisligil GS. Inflammation, stress, and diabetes. *The Journal of clinical investigation*. 2005;115(5):1111-9.
20. Boden G, Chen X, Ruiz J, White JV, Rossetti L. Mechanisms of fatty acid-induced inhibition of glucose uptake. *The Journal of clinical investigation*. 1994;93(6):2438-46.
21. McKeigue PM, Shah B, Marmot MG. Relation of central obesity and insulin resistance with high diabetes prevalence and cardiovascular risk in South Asians. *Lancet (London, England)*. 1991;337(8738):382-6.

22. Redinger RN, Small DM. Bile composition, bile salt metabolism and gallstones. *Archives of internal medicine*. 1972;130(4):618-30.
23. Shirai K. Obesity as the core of the metabolic syndrome and the management of coronary heart disease. *Current medical research and opinion*. 2004;20(3):295-304.
24. Bergeron C, Boulet LP, Hamid Q. Obesity, allergy and immunology. *The Journal of allergy and clinical immunology*. 2005;115(5):1102-4.
25. Inge TH, King WC, Jenkins TM, Courcoulas AP, Mitsnefes M, Flum DR, et al. The effect of obesity in adolescence on adult health status. *Pediatrics*. 2013;132(6):1098-104.
26. Carroll KK. Obesity as a risk factor for certain types of cancer. *Lipids*. 1998;33(11):1055-9.
27. Effertz T, Engel S, Verheyen F, Linder R. The costs and consequences of obesity in Germany: a new approach from a prevalence and life-cycle perspective. *Eur J Health Econ*. 2015.
28. Hansen D, Marinus N, Remans M, Courtois I, Cools F, Calsius J, et al. Exercise tolerance in obese vs. lean adolescents: a systematic review and meta-analysis. *Obes Rev*. 2014;15(11):894-904.
29. Mendelson M, Michallet AS, Esteve F, Perrin C, Levy P, Wuyam B, et al. Ventilatory responses to exercise training in obese adolescents. *Respir Physiol Neurobiol*. 2012;184(1):73-9.
30. Nadeau KJ, Zeitler PS, Bauer TA, Brown MS, Dorosz JL, Draznin B, et al. Insulin resistance in adolescents with type 2 diabetes is associated with impaired exercise capacity. *J Clin Endocrinol Metab*. 2009;94(10):3687-95.
31. Drinkard B, Roberts MD, Ranzenhofer LM, Han JC, Yanoff LB, Merke DP, et al. Oxygen-uptake efficiency slope as a determinant of fitness in overweight adolescents. *Medicine and science in sports and exercise*. 2007;39(10):1811-6.
32. Laguna M, Aznar S, Lara MT, Lucia A, Ruiz JR. Heart rate recovery is associated with obesity traits and related cardiometabolic risk factors in children and adolescents. *Nutrition, metabolism, and cardiovascular diseases : NMCD*. 2013;23(10):995-1001.
33. Guizar JM, Ahuatzin R, Amador N, Sanchez G, Romer G. Heart autonomic function in overweight adolescents. *Indian Pediatr*. 2005;42(5):464-9.
34. Hansen D, Dendale P. Modifiable predictors of chronotropic incompetence in male patients with type 2 diabetes. *J Cardiopulm Rehabil Prev*. 2014;34(3):202-7.
35. Felsher J, Meissner MD, Hakki AH, Heo J, Kane-Marsch S, Iskandrian AS. Exercise thallium imaging in patients with diabetes mellitus. Prognostic implications. *Archives of internal medicine*. 1987;147(2):313-7.
36. Mahler DA, Franco MJ. Clinical applications of cardiopulmonary exercise testing. *Journal of cardiopulmonary rehabilitation*. 1996;16(6):357-65.
37. Wasserman K. ea. *Principles of Exercise Testing and Interpretation: Including Pathophysiology and Clinical Applications*. 5th edition ed: LIPPINCOTT WILLIAMS & WILKINS; 2012.
38. Keytsman C, Dendale P, Hansen D. Chronotropic Incompetence During Exercise in Type 2 Diabetes: Aetiology, Assessment Methodology, Prognostic Impact and Therapy. *Sports Med*. 2015;45(7):985-95.
39. Rubin DA, Clark SJ, Ng J, Castner DM, Haqq AM, Judelson DA. Hormonal and metabolic responses to endurance exercise in children with Prader-Willi syndrome and non-syndromic obesity. *Metabolism*. 2015;64(3):391-5.
40. Salvadori A, Fanari P, Tovaglieri I, Giacomotti E, Nibbio F, Belardi F, et al. Ventilation and its control during incremental exercise in obesity. *Respiration; international review of thoracic diseases*. 2008;75(1):26-33.
41. Tadic M, Cuspidi C. Childhood obesity and cardiac remodeling: from cardiac structure to myocardial mechanics *Journal of Cardiovascular Medicine*. 2015;16(8):538-46.
42. Latchman PL, Mathur M, Bartels MN, Axtell RS, De Meersman RE. Impaired autonomic function in normotensive obese children. *Clin Auton Res*. 2011;21(5):319-23.
43. McCrindle BW. Cardiovascular consequences of childhood obesity. *Can J Cardiol*. 2015;31(2):124-30.
44. Üner A, Doğan M, Epcacan Z, Epeçan S. The effect of childhood obesity on cardiac functions. *Journal of Pediatric Endocrinology and Metabolism*. 2014;27(3-4):261.

45. Houtkooper LB, Going SB, Lohman TG, Roche AF, Van Loan M. Bioelectrical impedance estimation of fat-free body mass in children and youth: a cross-validation study. *Journal of applied physiology* (Bethesda, Md : 1985). 1992;72(1):366-73.
46. Slaughter MH, Lohman TG, Boileau RA, Horswill CA, Stillman RJ, Van Loan MD, et al. Skinfold Equations for Estimation of Body Fatness in Children and Youth. *Human Biology*. 1988;60(5):709-23.
47. Rodriguez G, Moreno LA, Blay MG, Blay VA, Fleta J, Sarria A, et al. Body fat measurement in adolescents: comparison of skinfold thickness equations with dual-energy X-ray absorptiometry. *European journal of clinical nutrition*. 2005;59(10):1158-66.
48. Marshall WA, Tanner JM. Variations in pattern of pubertal changes in girls. *Archives of disease in childhood*. 1969;44(235):291-303.
49. Marshall WA, Tanner JM. Variations in the pattern of pubertal changes in boys. *Archives of disease in childhood*. 1970;45(239):13-23.
50. Bervoets L, Van Noten C, Van Roosbroeck S, Hansen D, Van Hoorenbeeck K, Verheyen E, et al. Reliability and Validity of the Dutch Physical Activity Questionnaires for Children (PAQ-C) and Adolescents (PAQ-A). *Archives of public health = Archives belges de sante publique*. 2014;72(1):47.
51. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*. 1985;28(7):412-9.
52. Martínez-Vizcaino V, Martínez MS, Aguilar FS, Martínez SS, Gutiérrez RF, López MS, et al. Validity of a Single-Factor Model Underlying the Metabolic Syndrome in Children. A confirmatory factor analysis. 2010;33(6):1370-2.
53. Kreyszig E. *Advanced Engineering Mathematics* Fourth ed: Wiley; 1979.
54. Leeson P ea. *Echocardiography*, second edition: Oxford Specialist Handbooks in Cardiology; 2012.
55. Söderqvist E, Cain P, Lind B, Winter R, Nowak J, Brodin L-Å. Feasibility of creating estimates of left ventricular flow-volume dynamics using echocardiography. *Cardiovascular Ultrasound*. 2006;4:40-.
56. Andersen OS, Smiseth OA, Dokainish H, Abudiab MM, Schutt RC, Kumar A, et al. Estimating Left Ventricular Filling Pressure by Echocardiography. *Journal of the American College of Cardiology*. 2017;69(15):1937-48.
57. Dalla Pozza R, Ehringer-Schetitska D, Fritsch P, Jokinen E, Petropoulos A, Oberhoffer R, et al. Intima media thickness measurement in children: A statement from the Association for European Paediatric Cardiology (AEPC) Working Group on Cardiovascular Prevention endorsed by the Association for European Paediatric Cardiology. *Atherosclerosis*. 2015;238(2):380-7.
58. Baba R, Iwagaki S, Tauchi N, Tsurusawa M. Is the chronotropic index applicable to children and adolescents? *Circulation journal : official journal of the Japanese Circulation Society*. 2005;69(4):471-4.
59. Tanaka H, Monahan KD, Seals DR. Age-predicted maximal heart rate revisited. *Journal of the American College of Cardiology*. 2001;37(1):153-6.
60. Machado FA, Denadai BS. Validity of maximum heart rate prediction equations for children and adolescents. *Arquivos brasileiros de cardiologia*. 2011;97(2):136-40.
61. Bongers BC, Hulzebos EH, Helbing WA, Ten Harkel A, van Brussel M, Takken T. Response profiles of oxygen uptake efficiency during exercise in healthy children. *Eur J Prev Cardiol*. 2016;23(8):865-73.
62. Higgins V, Adeli K. Pediatric Metabolic Syndrome: Pathophysiology and Laboratory Assessment. *EJIFCC*. 2017;28(1):25-42.
63. Khoury M, Manlhiot C, McCrindle BW. Role of the Waist/Height Ratio in the Cardiometabolic Risk Assessment of Children Classified by Body Mass Index. *Journal of the American College of Cardiology*. 2013;62(8):742-51.
64. Neovius M, Linne Y, Rossner S. BMI, waist-circumference and waist-hip-ratio as diagnostic tests for fatness in adolescents. *International journal of obesity and related metabolic disorders : journal of the International Association for the Study of Obesity*. 2004;29(2):163-9.

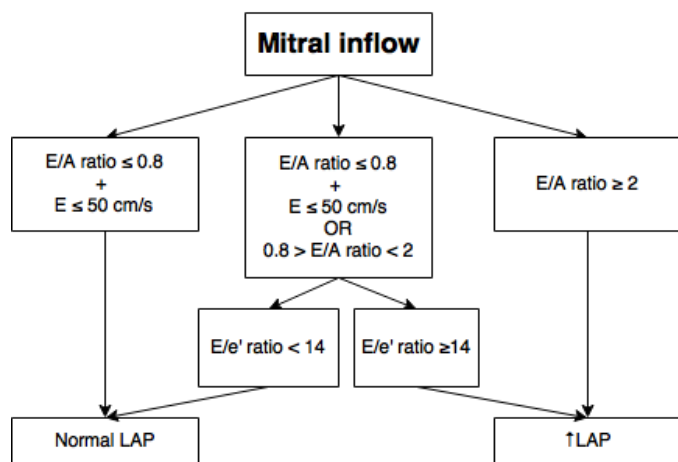
65. Thaler MS. *The Only EKG Book You'll Ever Need*: Lippincott Williams & Wilkins; 2007.
66. Sun G-Z, Li Y, Zhou X-H, Guo X-F, Zhang X-G, Zheng L-Q, et al. Association between obesity and ECG variables in children and adolescents: A cross-sectional study. *Experimental and Therapeutic Medicine*. 2013;6(6):1455-62.
67. Magnani JW, Wang N, Nelson KP, Connelly S, Deo R, Rodondi N, et al. The Electrocardiographic PR Interval and Adverse Outcomes in Older Adults: the Health, Aging and Body Composition Study. *Circulation Arrhythmia and electrophysiology*. 2013;6(1):84-90.
68. Peguero JG, Lo Presti S, Perez J, Issa O, Brenes JC, Tolentino A. Electrocardiographic Criteria for the Diagnosis of Left Ventricular Hypertrophy. *Journal of the American College of Cardiology*. 2017;69(13):1694-703.
69. Rider OJ, Ntusi N, Bull SC, Nethononda R, Ferreira V, Holloway CJ, et al. Improvements in ECG accuracy for diagnosis of left ventricular hypertrophy in obesity. *Heart (British Cardiac Society)*. 2016;102(19):1566-72.
70. Gao Z, Khoury PR, McCoy CE, Shah AS, Kimball TR, Dolan LM, et al. Adiposity Has No Direct Effect on Carotid Intima-Media Thickness in Adolescents and Young Adults: Use of Structural Equation Modeling to Elucidate Indirect & Direct Pathways. *Atherosclerosis*. 2016;246:29-35.
71. Yan Y, Hou D, Liu J, Zhao X, Cheng H, Yang P, et al. [Effect of childhood adiposity on long-term risks of carotid atherosclerosis and arterial stiffness in adulthood]. *Zhonghua yu fang yi xue za zhi [Chinese journal of preventive medicine]*. 2016;50(1):28-33.
72. Kim MS, Dao-Tran A, Davidowitz E, Tseng T, Gilsanz V, Ryabets-Lienhard A, et al. Carotid Intima-Media Thickness Is Associated with Increased Androgens in Adolescents and Young Adults with Classical Congenital Adrenal Hyperplasia. *Horm Res Paediatr*. 2016;85(4):242-9.
73. Kulshreshtha B, Arpita A, Rajesh PT, Sameek B, Dutta D, Neera S, et al. Adolescent gynecomastia is associated with a high incidence of obesity, dysglycemia, and family background of diabetes mellitus. *Indian Journal of Endocrinology and Metabolism*. 2017;21(1):160-4.
74. Brady TM. The Role of Obesity in the Development of Left Ventricular Hypertrophy Among Children and Adolescents. *Current hypertension reports*. 2016;18(1):3-.
75. Tsang TS, Barnes ME, Bailey KR, Leibson CL, Montgomery SC, Takemoto Y, et al. Left atrial volume: important risk marker of incident atrial fibrillation in 1655 older men and women. *Mayo Clin Proc*. 2001;76(5):467-75.
76. Park J-H, Marwick TH. Use and Limitations of E/e' to Assess Left Ventricular Filling Pressure by Echocardiography. *Journal of Cardiovascular Ultrasound*. 2011;19(4):169-73.
77. Huang L, Chen P, Zhuang J, Walt S. Metabolic cost, mechanical work, and efficiency during normal walking in obese and normal-weight children. *Research quarterly for exercise and sport*. 2013;84 Suppl 2:S72-9.
78. Faria AG, Ribeiro MA, Marson FA, Schivinski CI, Severino SD, Ribeiro JD, et al. Effect of exercise test on pulmonary function of obese adolescents. *Jornal de pediatria*. 2014;90(3):242-9.
79. Tanner CJ, Barakat HA, Dohm GL, Pories WJ, MacDonald KG, Cunningham PRG, et al. Muscle fiber type is associated with obesity and weight loss. *American Journal of Physiology - Endocrinology And Metabolism*. 2002;282(6):E1191.
80. Norman A-C, Drinkard B, McDuffie JR, Ghorbani S, Yanoff LB, Yanovski JA. Influence of Excess Adiposity on Exercise Fitness and Performance in Overweight Children and Adolescents. *Pediatrics*. 2005;115(6):e690-e6.
81. Park AE, Huynh P, Schell AM, Baker LA. Relationship between obesity, negative affect and basal heart rate in predicting heart rate reactivity to psychological stress among adolescents. *Int J Psychophysiol*. 2015;97(2):139-44.
82. Rabbia F, Silke B, Conterno A, Grosso T, De Vito B, Rabbone I, et al. Assessment of cardiac autonomic modulation during adolescent obesity. *Obesity research*. 2003;11(4):541-8.
83. Nagai N, Matsumoto T, Kita H, Moritani T. Autonomic nervous system activity and the state and development of obesity in Japanese school children. *Obesity research*. 2003;11(1):25-32.
84. Vanderlei LC, Pastre CM, Freitas Junior IF, Godoy MF. Analysis of cardiac autonomic modulation in obese and eutrophic children. *Clinics (Sao Paulo, Brazil)*. 2010;65(8):789-92.

85. Singh TP, Rhodes J, Gauvreau K. Determinants of heart rate recovery following exercise in children. *Medicine and science in sports and exercise*. 2008;40(4):601-5.
86. Buchheit M, Papelier Y, Laursen PB, Ahmaidi S. Noninvasive assessment of cardiac parasympathetic function: postexercise heart rate recovery or heart rate variability? *American Journal of Physiology - Heart and Circulatory Physiology*. 2007;293(1):H8-H10.
87. Jouven X, Empana JP, Schwartz PJ, Desnos M, Courbon D, Ducimetiere P. Heart-rate profile during exercise as a predictor of sudden death. *The New England journal of medicine*. 2005;352(19):1951-8.
88. Cole CR, Blackstone EH, Pashkow FJ, Snader CE, Lauer MS. Heart-rate recovery immediately after exercise as a predictor of mortality. *The New England journal of medicine*. 1999;341(18):1351-7.
89. Taşçılar ME, Yokuşoğlu M, Boyraz M, Baysan O, Köz C, Dündaröz R. Cardiac Autonomic Functions in Obese Children - Original Article. *Journal of Clinical Research in Pediatric Endocrinology*. 2011;3(2):60-4.
90. Cho KI, Jo EA, Cho SH, Kim BH. The Influence of Epicardial Fat and Nonalcoholic Fatty Liver Disease on Heart Rate Recovery in Metabolic Syndrome. *Metabolic syndrome and related disorders*. 2017;15(5):226-32.
91. Paolisso G, Manzella D, Montano N, Gambardella A, Varricchio M. Plasma leptin concentrations and cardiac autonomic nervous system in healthy subjects with different body weights. *J Clin Endocrinol Metab*. 2000;85(5):1810-4.
92. Bravo PE, Morse S, Borne DM, Aguilar EA, Reisin E. Leptin and Hypertension in Obesity. *Vascular Health and Risk Management*. 2006;2(2):163-9.
93. Krishnamurthy VR, Baird BC, Wei G, Greene T, Raphael K, Beddhu S. Associations of Serum Alkaline Phosphatase with Metabolic Syndrome and Mortality. *The American journal of medicine*. 2011;124(6):566.e1-.e7.
94. Malo MS. A High Level of Intestinal Alkaline Phosphatase Is Protective Against Type 2 Diabetes Mellitus Irrespective of Obesity(). *EBioMedicine*. 2015;2(12):2016-23.
95. Aeschbacher S, Schoen T, Dorig L, Kreuzmann R, Neuhauser C, Schmidt-Trucksass A, et al. Heart rate, heart rate variability and inflammatory biomarkers among young and healthy adults. *Annals of medicine*. 2017;49(1):32-41.
96. Franchini M, Salvagno GL, Montagnana M, Lippi G. Serum ferritin levels correlate with haemoglobin concentration: a report on 589 outpatients from a single centre. *Blood Transfusion*. 2007;5(4):244-5.
97. Yokusoglu M, Nevruz O, Baysan O, Uzun M, Demirkol S, Avcu F, et al. The altered autonomic nervous system activity in iron deficiency anemia. *The Tohoku journal of experimental medicine*. 2007;212(4):397-402.
98. Datta D, Normandin E, ZuWallack R. Cardiopulmonary exercise testing in the assessment of exertional dyspnea. *Annals of Thoracic Medicine*. 2015;10(2):77-86.
99. Bhambhani Y, Norris S, Bell G. Prediction of stroke volume from oxygen pulse measurements in untrained and trained men. *Canadian journal of applied physiology = Revue canadienne de physiologie appliquee*. 1994;19(1):49-59.
100. Charkoudian N, Rabbitts JA. Sympathetic Neural Mechanisms in Human Cardiovascular Health and Disease. *Mayo Clinic Proceedings*. 2009;84(9):822-30.
101. Kishi T. Heart failure as an autonomic nervous system dysfunction. *Journal of Cardiology*. 2012;59(2):117-22.
102. Hogg K, McMurray J. Neurohumoral pathways in heart failure with preserved systolic function. *Progress in cardiovascular diseases*. 2005;47(6):357-66.
103. Kim H-L, Zo J-H, Seo J-B, Chung W-Y, Kim Y-J, Kim S-H, et al. Additional value of lateral tissue Doppler imaging in the assessment of diastolic dysfunction among subjects with pseudonormal pattern of mitral inflow. *Cardiovascular Ultrasound*. 2013;11:31-.
104. Atterhog JH, Loogna E. P-R interval in relation to heart rate during exercise and the influence of posture and autonomic tone. *Journal of electrocardiology*. 1977;10(4):331-6.

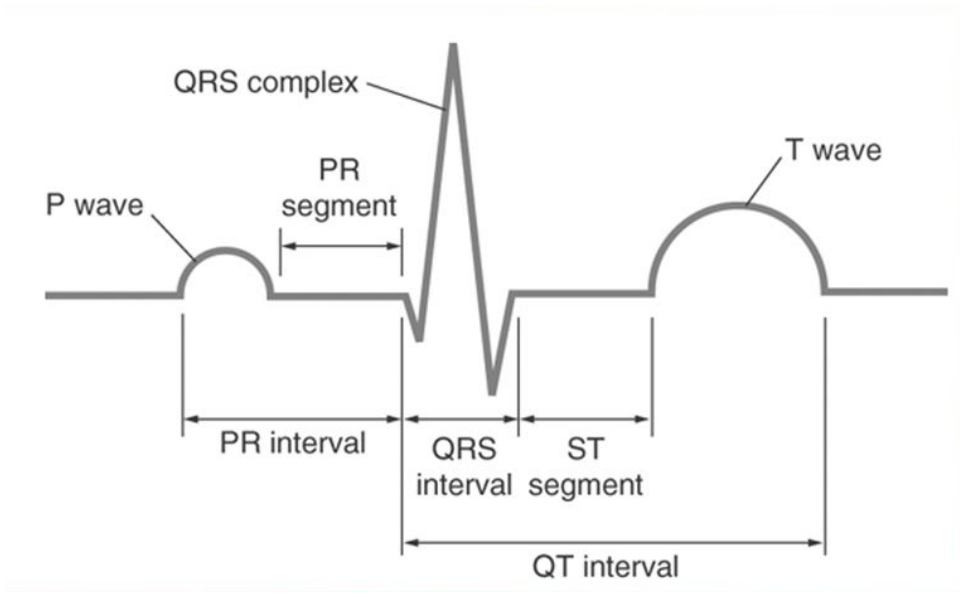
9. Appendices

Testing variable	Additive	Product	Brand
Glucose	Fluoride- Oxalaat	Vacutainer ® Blood collection tube	BD (New Jersey, USA)
CRP			
Iron	Lithium heparine		
AST			
GGT			
ALT			
Uric Acid			
K			
Ca			
Cholesterol			
HDL			
LDL			
Triglycerides	Clot activator		
ALP			
Ferritin			
TSH			
Free T4			
Cortisol			
Insulin			
Hb1ac			
Leptin		K2EDTA	
Hemoglobine			
Hematocrit			
White blood cells			

Appendix 1: Blood collection tubes used in fasting blood sampling



Appendix 2: Algorithm to evaluate left atrial pressure (LAP) (56)



Appendix 3: Intervals of the ECG (65)

	HRreserve		HRR 30		HRR60		HRR120	
	r	p-value	r	p-value	r	p-value	r	p-value
<u>Metabolic Risk model</u>								
Metabolic Risk	-0.297	0.05	-0.393	0.0006	-0.414	0.004	-0.39	0.014
<u>Metabolic parameters model</u>								
WHR	-0.369	0.014						
Leptin (ng/ml)			-0.462	0.001				
ALP (IU/L)			0.578	<0.001	0.508	<0.001		
Triglycerides (mg/dl)					-0.48	0.001	-0.411	0.009
CRP (mg/dl)							-0.468	0.003
Iron (µg/dl)					0.373	0.012		
ferritin (ng/ml)							0.033	0.844
Hb (g/dl)					0.222	0.138		
<u>Cardiovascular parameters model</u>								
Peak VCO ₂ (ml/min)	0.142	0.008						
Peak VO ₂ /HR					0.255	0.087		
A mitral (cm/sec)			-0.398	0.007	-0.36	0.018		
Mitral E/A ratio							0.617	<0.001
<u>ECG parameters model</u>								
Rest PR-interval (ms)					0.489	0.001	0.489	0.002
Peak PR-interval (ms)			0.387	0.007	0.562	0		

Abbreviations: r, Spearman's rho correlation coefficient; HRreserve, heart rate reserve (Δ HR_{peak}-HR_{rest}); HRR, heart rate recovery (Δ HR_{peak} - HR); AST, aspartate aminotransferase; GGT, gamma-glutamyl transpeptidase; ALP, alkaline phosphatase; CRP, c-reactive protein; Hb, Haemoglobin; VCO₂, carbon dioxide output; VO₂/HR, oxygen pulse; E mitral, transmitral peak early diastolic velocity; A mitral, transmitral peak late diastolic velocity

Appendix 4: Correlations between predictors of the heart rate response models

Auteursrechtelijke overeenkomst

Ik/wij verlenen het wereldwijde auteursrecht voor de ingediende eindverhandeling:
Heart rate responses during exercise in obese adolescents: relation with cardiovascular health

Richting: **Master of Biomedical Sciences-Environmental Health Sciences**

Jaar: **2017**

in alle mogelijke mediaformaten, - bestaande en in de toekomst te ontwikkelen - , aan de Universiteit Hasselt.

Niet tegenstaand deze toekenning van het auteursrecht aan de Universiteit Hasselt behoud ik als auteur het recht om de eindverhandeling, - in zijn geheel of gedeeltelijk -, vrij te reproduceren, (her)publiceren of distribueren zonder de toelating te moeten verkrijgen van de Universiteit Hasselt.

Ik bevestig dat de eindverhandeling mijn origineel werk is, en dat ik het recht heb om de rechten te verlenen die in deze overeenkomst worden beschreven. Ik verklaar tevens dat de eindverhandeling, naar mijn weten, het auteursrecht van anderen niet overtreedt.

Ik verklaar tevens dat ik voor het materiaal in de eindverhandeling dat beschermd wordt door het auteursrecht, de nodige toelatingen heb verkregen zodat ik deze ook aan de Universiteit Hasselt kan overdragen en dat dit duidelijk in de tekst en inhoud van de eindverhandeling werd genotificeerd.

Universiteit Hasselt zal mij als auteur(s) van de eindverhandeling identificeren en zal geen wijzigingen aanbrengen aan de eindverhandeling, uitgezonderd deze toegelaten door deze overeenkomst.

Voor akkoord,

Beyens, Marjolein

Datum: **8/06/2017**