

Review

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Overview of the optimal perioperative intraperitoneal chemotherapy regimens used in current clinical practice

DOI 10.1515/pap-2017-0003

Received February 5, 2017; accepted March 20, 2017;
previously published online April 7, 2017

Abstract: Peritoneal surface malignancy (PSM) is a common manifestation of digestive and gynecologic malignancies alike. At present, patients with isolated PSM are treated with a combination therapy of cytoreductive surgery (CRS) and hyperthermic intraperitoneal chemotherapy (HIPEC). The combination of CRS and intraperitoneal (IP) chemotherapy should now be considered standard of care for PSM from appendiceal epithelial cancers, colorectal cancer and peritoneal mesothelioma. Although there is a near universal standardization regarding the CRS, we are still lacking a much-needed standardization among the various IP chemotherapy treatment modalities used today in clinical practice. Pharmacologic evidence should be generated to answer important questions raised by the myriad of variables associated with IP chemotherapy.

Keywords: bidirectional intraoperative chemotherapy (BIC), early postoperative intraperitoneal chemotherapy (EPIC), hyperthermic intraperitoneal chemotherapy (HIPEC), peritoneal surface malignancy

Introduction

Peritoneal surface malignancy (PSM) is a common manifestation of digestive and gynecologic malignancies alike. At present, patients with isolated PSM are treated with a combination therapy of cytoreductive surgery (CRS) and hyperthermic intraperitoneal chemotherapy (HIPEC) [1].

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CRS and HIPEC have evolved over three decades and have demonstrated encouraging clinical results in several phase II and III trials [2–10]. Eveno et al. provided a thorough overview of randomized controlled trials evaluating CRS and HIPEC vs. other strategies in prevention and therapy of PSM [11]. The combined treatment modality should now be considered standard of care for PSM from appendiceal epithelial cancers, colorectal cancer and peritoneal mesothelioma [12–14]. Promising results have also been published for HIPEC in ovarian cancer and gastric cancer [5, 9, 15]. Although there is now a clearly defined standardization of CRS, based on the work by Sugarbaker et al. [16, 17], no standardized intraperitoneal (IP) chemotherapy treatment modalities exist. Variables to be considered are normothermic vs. HIPEC, open vs. closed HIPEC technique, but also the use of HIPEC with or without early postoperative intraperitoneal chemotherapy (EPIC) or the sole administration of EPIC. This also implicates the important pharmacologic variables associated with the chemotherapy agents that are currently available for the administration of HIPEC [18] and EPIC [19]. There is a pressing need to generate pharmacologic data working toward standardization among the myriad of IP treatment protocols currently applied. The current IP chemotherapy regimens are non-standardized, poorly predictable and based largely on extrapolation of systemic chemotherapy data (Figure 1). In this manuscript, we review the current IP chemotherapy practice for colorectal, appendiceal, gastric, ovarian PSM and peritoneal mesothelioma.

Ideal IP chemotherapy regimens

The ideal IP chemotherapy regimen has to meet two important requirements: First, we have to construct the pharmacologic best way of delivery of our cancer chemotherapy agent from the moment of application until the penetration into the individual tumor cell. Pharmacology of IP chemotherapy can be artificially divided between pharmacokinetics and pharmacodynamics. Whereas pharmacokinetics

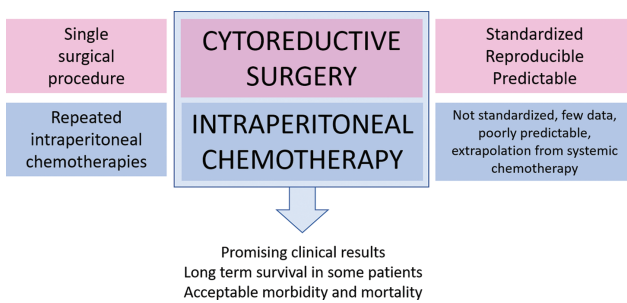


Figure 1: Treatment of peritoneal surface malignancy.

describes what the body does to the drug, pharmacodynamics looks at what the drug does to the body. Pharmacokinetics of IP chemotherapy studies the alterations between the moment of administration of the IP chemotherapy and the cancer chemotherapy drug showing up at the level of the tumor nodule. Important pharmacokinetic variables include drug dose, volume, duration, carrier solution, pressure and molecular weight. The basic way of depicting pharmacokinetic data is by a concentrations \times time graph. The area-under-the-curve (AUC) ratio IP/IV is important in that it quantifies the dose intensity expected in the treatment of PSM. Pharmacodynamics subsequently looks into the effect of that cancer chemotherapy drug on the tumor, considering tumor nodule size, density, vascularity, interstitial fluid pressure, binding and temperature. Pharmacodynamics data are depicted in a concentrations \times effect graph.

A second equally important requirement is the choice of the correct cancer chemotherapy drug with a cytotoxic activity against that specific cancer cell [19]. This emphasizes the increasing importance of chemosensitivity testing, toward a patients-tailored approach of selecting the ideal drug for IP and/or IV administration. At present several preclinical work has been conducted in this field using a wide variety of *in vitro*, *in vivo* and *ex vivo* assays using several patient-derived tumor cell lines in combination with several chemotherapy agents [20, 21]. However, important to note is that during the *in vitro* assays, the 3-D structure of the tumors and hence the important pharmacodynamics of the nodules are lost. Moreover, metabolization which is very important for the cytotoxic effect of several drugs is not taking in to account. *Ex vivo* assays using patient-derived xenografts and orthotopic animal models also present an impaired view of the clinical situation. For example, implantation of tumor cells subcutaneously, due to differences in microenvironment, will result in the formation of one tumor nodule which fails to progress and metastasize. Further research, and careful validation of such assays are needed, taking into account the

heterogeneity of tumors and the important pharmacodynamic variables. In the present era of omics assays, gene expression profiling has gained increasing interest in clinical applications to predict oncologic outcomes. Levine et al. analyzed gene expression profiles of appendiceal and colorectal PSM samples from patients undergoing HIPEC after complete CRS. They reported distinct genomic signatures for colorectal PSM when compared to appendiceal PSM. Three distinct phenotypes, two consisting of predominantly appendiceal samples (low-risk appendiceal and high-risk appendiceal) and the third with predominantly primary colorectal samples (high-risk colorectal), were identified. Furthermore, overall survival (120 months) after optimal CRS and HIPEC was significantly different between the low-risk appendiceal and the high-risk colorectal group [22]. Fujishima et al. used immunohistochemistry to evaluate mucin (MUC) protein expression in tumor nodules of patients with peritoneal dissemination from colorectal cancer as the only synchronous distant metastasis, who had received HIPEC. They report that in patients positive for MUC2 expression the 3-year overall survival rate was 0.0%, whereas in patients negative for MUC2 expression, the 3-year overall survival rate was 61.1% [23]. This emphasizes the importance of omics assays to help define better candidates for certain therapies and possibly, in the near future, the choice of chemotherapeutic agents.

IP cancer chemotherapy regimens for colorectal or appendiceal PSM

Table 1 summarizes the most frequently used IP cancer chemotherapy regimens in colorectal and appendiceal PSM. The two dominant cancer drugs that form the backbone of these regimens are oxaliplatin and mitomycin C.

Oxaliplatin

Oxaliplatin (oxalato-1,2-diaminocyclohexane-platinum(II)) is a third-generation platinum complex with proven cytotoxicity in colon and appendiceal neoplasms [24]. In a dose escalation and pharmacokinetic study, Elias et al. demonstrated that 460 mg/m² of oxaliplatin in 2L/m² of chemotherapy solution over 30 min was well tolerated [25, 26]. The low AUC ratio is compensated by the rapid absorption of the drug into the tissue, being the reason for the short application time. Since there initiation into the IP chemotherapy regimens at the beginning of the 2000s, we see a trend toward lower oxaliplatin-dosed

Table 1: Hyperthermic intraperitoneal chemotherapy (HIPEC)- and bidirectional intraoperative chemotherapy (BIC)-regimens.**Oxaliplatin-based regimens****Elias high dose oxaliplatin regimen**

1. Add oxaliplatin to 2 L/m² 5 % dextrose solution
 2. Dose of oxaliplatin is 460 mg/m²
 3. 30-min HIPEC treatment
- Intravenous component
4. Add 5-fluorouracil 400 mg/m² and leucovorin 20 mg/m² to separate bags of 250 mL normal saline. Begin rapid intravenous infusion of both drugs 1 h before intraperitoneal chemotherapy

Glehen medium dose oxaliplatin regimen

1. Add oxaliplatin to 2 L/m² 5 % dextrose solution
 2. Dose of oxaliplatin is 360 mg/m²
 3. 30-min HIPEC treatment
- Intravenous component
4. Add 5-fluorouracil 400 mg/m² and leucovorin 20 mg/m² to separate bags of 250 mL normal saline. Begin rapid intravenous infusion of both drugs 1 h before intraperitoneal chemotherapy

Wake forest university oxaliplatin regimen

1. Add oxaliplatin to 3 L 5 % dextrose solution
2. Dose of oxaliplatin is 200 mg/m²
3. Two hour HIPEC treatment

Mitomycin C-based regimens**Sugarbaker regimen**

1. Add mitomycin C to 2 L 1.5 % dextrose peritoneal dialysis solution
2. Add doxorubicin to the same 2 L 1.5 % peritoneal dialysis solution
3. Dose of mitomycin C and doxorubicin is 15 mg/m² for each chemotherapy agent
4. Add 5-fluorouracil (400 mg/m²) and leucovorin (20 mg/m²) to separate bags of 250 mL normal saline. Begin rapid intravenous infusion of both drugs simultaneous with intraperitoneal chemotherapy

Dutch high dose mitomycin C regimen: “Triple Dosing Regimen”

1. Add mitomycin C to 3 L 1.5 % dextrose peritoneal dialysis solution
2. Add mitomycin C to the 1.5 % peritoneal dialysis solution at a dose of 17.5 mg/m² followed by 8.8 mg/m² at 30 min and 8.8 mg/m² at 60 min
3. Total dose of mitomycin C 35 mg/m² for 90-min HIPEC treatment

American society of peritoneal surface malignancy low dose mitomycin C regimen: “Concentration-Based Regimen”

1. Add mitomycin C to 3 L 1.5 % dextrose peritoneal dialysis solution
2. Add mitomycin C to the 1.5 % peritoneal dialysis solution at a dose of 30 mg/3L followed by 10 mg at 60 min
3. Dose of mitomycin C 40 mg/3L for 90-min HIPEC treatment

regimens. This is based on a growing concern for unacceptable bleeding complications with the initial 460 mg/m²-based regimens. In a phase I trial, Elias et al. himself

evaluated the pharmacokinetics of heated IP oxaliplatin administered in increasingly hypotonic solutions of 5 % dextrose [27]. They reported that oxaliplatin clearance from the IP cavity was similar regardless of the osmolarity, but that very hypotonic solutions induce high incidence of IP hemorrhage and thrombocytopenia. As a result of high incidence hemorrhagic complications in another prospective multicenter trial organized by Pomel et al., the dose of oxaliplatin was reduced to 350 mg/m². However, the incidence of the hemorrhagic complications (29 %) did not decrease and the trial was closed prematurely [28]. Chalret du Rieu et al. performed a population pharmacokinetics study, including 75 patients, treated with CRS and oxaliplatin-based HIPEC [29]. They report grade 3/4 thrombocytopenia in 14 % of treated patients. Moreover, they concluded that the higher the absorbed dose from the peritoneal cavity, highly dependent on the initial oxaliplatin concentration, the deeper the resultant thrombocytopenia. In an analysis of 701 patients treated with CRS and HIPEC with oxaliplatin or other chemotherapeutic agents, Charrier et al. reported that oxaliplatin-based HIPEC increased the risk of hemorrhagic complications compared to other drugs [30]. In contrast to cisplatin and mitomycin, oxaliplatin traditionally is considered not stable in chloride-containing solutions. This necessitates a dextrose-based carrier which may result in serious electrolyte disturbances and hyperglycemia during the intracavitary therapy [31]. Unknown to most this degradation of oxaliplatin in normal saline only accounts for less than 10 % of the total amount at 30 min, as when applied during HIPEC. Moreover, oxaliplatin degradation was associated with the formation of its active drug form [Pt(dach)Cl₂] [32, 33]. Different oxaliplatin-based HIPEC regimens are used in current clinical practice: “Elias High Dose Oxaliplatin Regimen” [25], “Glehen Medium Dose Oxaliplatin Regimen” and the “Wake Forest University Oxaliplatin Regimen” [24].

Mitomycin C

Mitomycin C is an alkylating tumor antibiotic extracted from *Streptomyces* species which most important mechanism of action is through DNA cross-linking. Jacquet et al. reported a clear pharmacokinetic advantage after IP administration with an AUC IP/IV ratio of 23.5 [34]. It is used for PSM from colorectal cancer, appendiceal cancer, ovarian cancer, gastric cancer and, for diffuse malignant peritoneal mesothelioma both as HIPEC and as EPIC [2, 3, 34–38]. Barlogie et al. suggested *in vitro* thermal enhancement of mitomycin C [39, 40]. Our pharmacokinetic data in 145

HIPEC patients suggest that the largest proportion (62%) of the total drug administered remained in the body at 90 min [36]. Controversies still exist regarding the proper dosimetry of the chemotherapy solution. Triple dosing regimen may result in more stable peritoneal levels of the drug throughout the time of IP chemotherapy. Current applied HIPEC dosing regimens are the “Sugarbaker Regimen” [36], The “Dutch High Dose Mitomycin C Regimen: Triple Dosing Regimen” [41] and the “American Society of Peritoneal Surface Malignancy Low Dose Mitomycin C Regimen: Concentration-based Regimen” [42].

Body surface area-based or concentration-based IP chemotherapy

The current dosing regimens of IP chemotherapy can be divided into body surface area (BSA)-based and concentration-based. Most groups use a drug dose based on calculated BSA (mg/m^2) in analogy to systemic chemotherapy regimens. These regimens take BSA as a measure for the effective peritoneal contact area, the peritoneal surface area in the Dedrick formula [43]. The Dedrick formula on itself is an application of Fick’s law of diffusion. Rubin et al. [44] however, demonstrated there is an imperfect correlation between actual peritoneal surface area and calculated BSA. BSA-based IP chemotherapy will result in a fixed dose (BSA-based) diluted in varying volumes of perfusate, i.e.; different concentrations depending on substantial differences in the body composition of patients and differences in the HIPEC technique (open vs. closed abdomen). From the Dedrick formula we know that peritoneal concentration and not peritoneal dose is the driving diffusion force [43]. The importance of this has been discussed by Elias et al. [45] in a clinical investigation where 2-, 4-, and 6-liters of chemotherapy solution was administered with a constant dose of chemotherapy solution. A more dilute IP chemotherapy concentration retarded the clearance of chemotherapy and resulted in less systemic toxicity [46]. Therefore, it can be assumed that by the diffusion model, less concentrated chemotherapy would penetrate to a lesser extent into the cancer nodules and normal tissues. On the other hand,

concentration-based chemotherapy offers a more predictable exposure of the tumor nodules to the IP chemotherapy and thus efficacy [47]. Unfortunately, the prize to be paid for a better prediction of the efficacy of the IP chemotherapy is a high unpredictability of the plasmatic cancer chemotherapy levels and thus toxicity. Currently, there is an ongoing randomized trial at our hospital evaluating the pharmacology and morbidity of both dosing methods, entitled “concentration-based vs. body surface area-based perioperative intraperitoneal chemotherapy after optimal cytoreductive surgery in colorectal peritoneal carcinomatosis treatment: randomized non-blinded phase III clinical trial (COBOXtrial)” (<https://clinicaltrials.gov/ct2/show/NCT03028155>).

Clinical results

Table 2 summarizes the clinical trials comparing oxaliplatin-based and mitomycin c-based HIPEC [45–47]. Except for the study by Leung et al. all these studies show similar clinical outcome which seems to suggest non-inferiority of either drug [47]. All of these reports, however, have serious methodological issues (selection bias, historical bias, ...). A randomized control trial is in order to solve this issue. A multi-center, open-label, randomized phase II trial has been conducted to evaluate hematologic toxicities after HIPEC with oxaliplatin and mitomycin C in patients with appendiceal tumors (<https://clinicaltrials.gov/ct2/show/NCT01580410>). Time to progression after oxaliplatin- or mitomycin C-based HIPEC has also been evaluated. The accrual is completed and we are now awaiting the results.

IP cancer chemotherapy for PSM of gastric cancer, ovarian cancer, mesothelioma and sarcoma

The predominant IP regimens in this setting are cisplatin-based.

Table 2: Clinical studies of oxaliplatin-based vs. mitomycin-C-based HIPEC.

Year	Author	n	Type	Uni/multi	Result
2008	Elias	523	Retrospective	Multi [23]	MMC = Oxali
2014	Hompes	95	Retrospective	Bicentric	MMC = Oxali
2014	Prata-Villaverde	539	Retrospective	Multi (>15)	MMC = Oxali except PSDSS I/II (MMC 54.3 vs. 28.2 months)
2016	Leung	201	Retrospective	Unicentric	Oxali (OS 56 vs. 29 months)

Cisplatin

Cisplatin (cis-diamminedichloroplatinum-III, CDDP) is an alkylating agent that causes apoptotic cell death by formation of DNA adducts [48]. Both normothermic and hyperthermic IP applications have been explored in the treatment of ovarian cancer, gastric cancer and peritoneal mesothelioma [4, 18, 49–54]. It is eliminated by renal excretion and consequently the main concern with its use is renal toxicity [55]. Urano et al. showed an excellent *in vitro* and *in vivo* thermal augmentation of cisplatin [56]. Current applied cisplatin-based HIPEC regimens are the “Sugarbaker Regimen” [57] and the “National Cancer Institute Milan Regimen” [58] (Table 3).

Table 3: Cisplatin-based HIPEC regimens.

Cisplatin-based regimens	
Sugarbaker regimen	
1.	Add cisplatin to 2 L 1.5 % dextrose peritoneal dialysis solution
2.	Add doxorubicin to the same 2 L 1.5 % peritoneal dialysis solution
3.	Dose of cisplatin is 50 mg/m ² and doxorubicin is 15 mg/m ² for 90-min HIPEC treatment
Intravenous chemotherapy	
4.	Add ifosfamide 1,300 mg/m ² to 1 L 0.9 % sodium chloride. Begin continuous IV infusion over 90 min simultaneous with intraperitoneal chemotherapy
5.	Add mesna disulfide 260 mg/m ² in 100 mL 0.9 % sodium chloride to be given IV as a bolus 15 min prior to ifosfamide infusion
6.	Add mesna disulfide 260 mg/m ² in 100 mL 0.9 % sodium chloride to be given IV as a bolus 4 h after ifosfamide infusion
7.	Add mesna disulfide 260 mg/m ² in 100 mL 0.9 % sodium chloride to be given IV as a bolus 8 h after ifosfamide infusion
National Cancer Institute Milan regimen	
1.	15.25 mg/L of doxorubicin and 43 mg/L of cisplatin for 90-min HIPEC treatment
2.	Chemotherapy solution 4–6 L based on capacity of the peritoneal space

Doxorubicin

Cisplatin-based regimens are often augmented with IP doxorubicin. Doxorubicin or hydroxydaunorubicin (adriamycin) is an anthracycline antibiotic. Initial research categorized it as a DNA-intercalating drug. It was later demonstrated that the actual mechanism of action is a temperature-dependent interaction of doxorubicin with the cell surface membrane [59–61]. Doxorubicin was considered a candidate for IP application based on its wide *in vitro* and *in vivo* activity against a broad range of malignancies, its

slow clearance from the peritoneal compartment due to the high molecular weight of the hydrochloride salt, its favorable AUC ratio of IP to IV concentration times of 230 [18, 62–66]. More recently PEGylated liposomal doxorubicin has generated interest for HIPEC application due to its favorable pharmacokinetics [67, 68].

Bidirectional intraoperative chemotherapy (BIC)

Most current protocols advocate bidirectional intraoperative chemotherapy (BIC). By combining intraoperative IV and intraoperative IP cancer chemotherapy, a bidirectional diffusion gradient is created through the intermediate tissue layer containing the cancer nodules. In 2002, Elias et al. first reported the clinical use of intraoperative IV 5-fluorouracil and leucovorin in conjunction with oxaliplatin-based HIPEC, to potentiate the effect of oxaliplatin [69]. We also reported a clear pharmacokinetic advantage for the intraoperative IV administration of 5-fluorouracil [69]. A similar pharmacokinetic advantage and heat targeting of intraoperative IV ifosfamide was demonstrated [57]. Ifosfamide is a prodrug that needs the cytochrome P450 system of liver or red blood cells to be activated to its active metabolite 4-hydroxyifosfamide. Consequently, it requires IV administration rather than IP instillation for its cytotoxic activity. The drug also shows true heat synergy. It may be an ideal systemic drug to increase the cytotoxicity of HIPEC. The bidirectional approach offers the possibility of optimizing cancer chemotherapy delivery to the target peritoneal tumor nodules. Further pharmacologic studies are needed to clarify the most efficient method of administration (continuous, bolus or, repeated bolus), doses and, choice of cancer chemotherapy drugs for this bidirectional approach.

Early postoperative intraperitoneal chemotherapy

EPIC has some conceptual advantages. It is administered shortly after CRS at the time of minimal residual tumor burden. Moreover, IP treatments initiated before wound healing occurs can minimize non-uniform drug distribution and eliminate residual cancer cell entrapment in postoperative fibrin deposits. Disadvantages associated with EPIC are the increased risks of infection and postoperative complications [70–73]. EPIC does not involve hyperthermia and is administered postoperatively (typically day 1 to

day 4/5) through both an inflow catheter and outflow drains inserted at the time of CRS and can be applied with or without HIPEC. Proper selection of chemotherapy agents based on pharmacologic principles suggests the use of cell-cycle specific drugs such as 5-fluorouracil and the taxanes (Table 4). This implies administering multiple cycles, each with a dwell time of around 23 h before renewal. This ensures that all the residual tumor cells are susceptible for the cell cycle specific drug.

5-Fluorouracil

The fluorinated pyrimidines have been successfully used for a wide variety of tumors and are still an essential component of all successful gastrointestinal cancer chemotherapy regimens [74, 75]. This thymidylate synthase inhibitor binds covalently with the enzyme and prevents the formation of thymidine monophosphate, the DNA nucleoside precursor. Also, 5-FU by its metabolites 5-fluoro-uridine diphosphate and 5-fluoro-uridine triphosphate gets incorporated in RNA, resulting in a second cytotoxic pathway. The action of 5-fluorouracil is therefore cell cycle specific. These characteristics limit the use of IP 5-fluorouracil to EPIC [19, 76–79]. 5-fluorouracil is not chemically compatible with other drugs in a mixed solution for infusion or instillation.

Taxanes

Paclitaxel and docetaxel, with their high molecular weight these molecules, have a remarkable high AUC ratio of respectively 853 and 861 [80]. The taxanes stabilize the microtubule against depolymerization, thereby disrupting normal microtubule dynamics [81]. There is evidence supporting additional mechanisms of action [82]. They exert cytotoxic activity against a broad range of tumors. This translates itself into a clear pharmacokinetic advantage for IP administration [83]. The data regarding possible thermal augmentation of taxanes are conflicting [82]. Taxanes have been used in a neoadjuvant intraperitoneal and systemic chemotherapy (NIPS) setting as well as intraoperatively and postoperatively. Their cell cycle specific mechanism of action makes them a better candidate for repetitive application such as in EPIC, NIPS or normothermic adjuvant postoperative IP chemotherapy.

Neoadjuvant intraperitoneal and systemic chemotherapy

Neoadjuvant bidirectional chemotherapy uses both the IP and IV routes of chemotherapy administration prior to the CRS. It has been suggested as an option for reducing dissemination to the extra-abdominal space, testing the

Table 4: Early postoperative intraperitoneal chemotherapy (EPIC)-regimens.

Early postoperative intraperitoneal chemotherapy with 5-fluorouracil on postoperative days 1 through 4 for adenocarcinoma from appendiceal, colonic and gastric cancer

1. 5-Fluorouracil _____ mg (400 mg/m² for females and 600 mg/m² for males, maximum dose = 1,400 mg) and 50 meq sodium bicarbonate in _____ mL 1.5% dextrose peritoneal dialysis solution via the Tenckhoff catheter daily for 4 days: start date _____, stop date _____.
2. The intraperitoneal fluid volume is 1L for patients ≤2.0 m² and 1.5 L for those >2.0 m².
3. Drain all fluid from the abdominal cavity prior to instillation, then clamp abdominal drains.
4. Run the chemotherapy solution into the abdominal cavity through the Tenckhoff catheter as rapidly as possible. Dwell for 23 h and drain for 1 h prior to next instillation.
5. Use gravity to maximize intraperitoneal distribution of the 5-fluorouracil. Instill the chemotherapy with the patient in a full right lateral position. After 30 min, direct the patient to turn to the full left lateral position. Change position right to left every 30 min. Continue turning for the first 6 h after instillation of chemotherapy solution.
6. Monitor with pulse oximeter during the first 6 h of intraperitoneal chemotherapy.
7. Continue to drain abdominal cavity after final dwell until Tenckhoff catheter is removed.

Early postoperative intraperitoneal chemotherapy with paclitaxel on postoperative days 1–5 for peritoneal mesothelioma and ovarian cancer

1. Paclitaxel _____ mg (20 to 40 mg/m² × _____ m²) (maximum dose = 80 mg) in 1,000 mL 6% Hespan® (B. Braun, Irvine, CA) via Tenckhoff catheter daily: start date _____, stop date _____.
 2. Instill as rapidly as possible via Tenckhoff catheter. Dwell for 23 h. Drain from Jackson-Pratt drains for 1 h prior to next instillation.
 3. During the initial 6 h after chemotherapy infusion, the patient's bed should be kept flat. The patient should be on the right side during instillation. Turn at 30 min post instillation onto the left side and continue to change sides at 30-min intervals for 6 h.
 4. Monitor with pulse oximeter during the first 6 h of intraperitoneal chemotherapy.
 5. Continue to drain abdominal cavity by Jackson-Pratt drains after the last dose of intraperitoneal chemotherapy.
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tumor biology and for reducing the extent of small PC nodules. Theoretically this approach, called NIPS, may facilitate definitive CRS after initial exploratory laparoscopy or laparotomy [84]. Radiological and clinical responses with NIPS have been reported by several groups [84–87]. However, although NIPS may reduce the tumor load to be addressed by CRS, it has several disadvantages. Adhesions from prior surgical interventions may interfere with adequate IP drug distribution and, as complete responses are unusual, further cytoreduction-chemotherapy is necessary if the approach is to be curative. NIPS is reported to add to morbidity and mortality of further surgical treatment [88]. Furthermore, extensive fibrosis, as a response to chemotherapy, may occur and render judgments concerning the extent of PC difficult or impossible. Table 5 lists the most commonly used NIPS regimens [83–88].

Table 5: Neoadjuvant intraperitoneal and systemic chemotherapy (NIPS) – regimens.

Yonemura regimen (2016)

1. Oral S-1 is administered for 14 days at a dose of 60 mg/m²/day, followed by 7 days rest prior to intraperitoneal chemotherapy administration
2. On day 1 docetaxel (30 mg/m²) and cisplatin (30 mg/m²) are administered by IP infusion
3. On day 8 (30 mg/m²) and cisplatin (30 mg/m²) are administered IV

Ishigami regimen

1. Oral S-140 mg/m² twice daily for 14 consecutive days followed by 7 days rest—I
2. Paclitaxel 50 mg/m² IV simultaneously with 20 mg/m² IP in 1L normal saline over 1 h on day 1 and day 8
3. Regimen repeated every 3 weeks

Fujiwara regimen

1. Oral S-140 mg/m² twice daily for 14 consecutive days
2. Docetaxel 40–60 mg/m² IP in 1L normal saline
3. Regimen repeated every 3 weeks

Sugarbaker regimen

1. Paclitaxel 20 mg/m² in 1L 6% Hespan (B. Braun, Irvine, CA, USA) via IP port or Tenckhoff catheter
 2. Instill by gravity flow as rapidly as possible 5 days in a row
 3. On day 3 of 5 day cycle, instill oxaliplatin 100–150 mg/m² IV in 250 mL D5W over 120 min. Start 30 min after IP paclitaxel
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Conclusions

The combination of CRS and IP chemotherapy should now be considered standard of care for PSM from appendiceal epithelial cancers, colorectal cancer and peritoneal mesothelioma. Although there is a near universal

standardization regarding the CRS, there is still a much-needed standardization among the various IP chemotherapy treatment modalities used today in clinical practice. This manuscript reviewed the most commonly used IP regimens for HIPEC, EPIC and NIPS. Although today, trends in the IP protocols, such as the reduced dosing of oxaliplatin and the triple dosing regimen of mitomycin C are observed; more pharmacologic and clinical evidence should be generated to answer important questions raised by the myriad of variables associated with IP chemotherapy.

Author contributions: All the authors have accepted responsibility for the entire content of this submitted manuscript and approved submission.

Research funding: Lemoine L is supported by the Agency for Innovation by Science and Technology (IWT) in Brussels, Belgium. Lemoine L is a researcher for the Limburg Clinical Research Program (LCRP) UHasselt-ZOL-Jessa, supported by the foundation Limburg Sterk Merk (LSM), Hasselt University, Ziekenhuis Oost-Limburg and Jessa Hospital, Belgium.

Employment or leadership: None declared.

Honorarium: None declared.

Competing interests: The funding organization(s) played no role in the study design; in the collection, analysis, and interpretation of data; in the writing of the report; or in the decision to submit the report for publication.

References

1. Mirnezami R, Mehta AM, Chandrakumaran K, Cecil T, Moran BJ, Carr N, et al. [Cytoreductive surgery in combination with hyperthermic intraperitoneal chemotherapy improves survival in patients with colorectal peritoneal metastases compared with systemic chemotherapy alone](#). *Br J Cancer* 2014;111:1500–8.
2. Verwaal VJ, Bruin S, Boot H, Van Slooten G, Van Tinteren H. 8-year follow-up of randomized trial: Cytoreduction and hyperthermic intraperitoneal chemotherapy versus systemic chemotherapy in patients with peritoneal carcinomatosis of colorectal cancer. *Ann Surg Oncol* 2008;15:2426–32.
3. Verwaal VJ, Van Ruth S, De Bree E, Van Sloothen GW, Van Tinteren H, Boot H, et al. [Randomized trial of cytoreduction and hyperthermic intraperitoneal chemotherapy versus systemic chemotherapy and palliative surgery in patients with peritoneal carcinomatosis of colorectal cancer](#). *J Clin Oncol* 2003;21:3737–43.
4. Baratti D, Kusamura S, Cabras AD, Bertulli R, Hutanu I, Deraco M. Diffuse malignant peritoneal mesothelioma: Long-term survival with complete cytoreductive surgery followed by hyperthermic intraperitoneal chemotherapy (HIPEC). *Eur J Cancer* 2013;49:3140–8.

5. Spiliotis J, Halkia E, Lianos E, Kalantzi N, Grivas A, Efstathiou E, et al. Cytoreductive surgery and HIPEC in recurrent epithelial ovarian cancer: A prospective randomized phase III study. *Ann Surg Oncol* 2015;22:1570–5.
6. Marcotte E, Dube P, Drolet P, Mitchell A, Frenette S, Leblanc G, et al. [Hyperthermic intraperitoneal chemotherapy with oxaliplatin as treatment for peritoneal carcinomatosis arising from the appendix and pseudomyxoma peritonei: A survival analysis.](#) *World J Surg Oncol* 2014;12:332.
7. Sideris L, Mitchell A, Drolet P, Leblanc G, Leclerc YE, Dube P. Surgical cytoreduction and intraperitoneal chemotherapy for peritoneal carcinomatosis arising from the appendix. *Can J Surg* 2009;52:135–41.
8. Yan TD, Deraco M, Baratti D, Kusamura S, Elias D, Glehen O, et al. [Cytoreductive surgery and hyperthermic intraperitoneal chemotherapy for malignant peritoneal mesothelioma: Multi-institutional experience.](#) *J Clin Oncol* 2009;27:6237–42.
9. Glehen O, Gilly FN, Arvieux C, Cotte E, Boutitie F, Mansvelt B, et al. Peritoneal carcinomatosis from gastric cancer: A multi-institutional study of 159 patients treated by cytoreductive surgery combined with perioperative intraperitoneal chemotherapy. *Ann Surg Oncol* 2010;17:2370–7.
10. Gonzalez Bayon L, Steiner MA, Vasquez Jimenez W, Asencio JM, Alvarez De Sierra P, Atahualpa Arenas F, et al. Cytoreductive surgery and hyperthermic intraperitoneal chemotherapy for the treatment of advanced epithelial ovarian carcinoma: Upfront therapy, at first recurrence, or later? *Eur J Surg Oncol* 2013;39:1109–15.
11. Clarisse Eveno MP. Randomized controlled trials evaluating cytoreductive surgery (CRS) and hyperthermic intraperitoneal chemotherapy (HIPEC) in prevention and therapy of peritoneal metastasis: A systemic review. *Pleura and Peritoneum* 2016;1:169–82.
12. Glehen O, Gilly FN, Boutitie F, Bereder JM, Quenet F, Sideris L, et al. Toward curative treatment of peritoneal carcinomatosis from nonovarian origin by cytoreductive surgery combined with perioperative intraperitoneal chemotherapy: A multi-institutional study of 1,290 patients. *Cancer* 2010;116:5608–18.
13. Sugarbaker PH. New standard of care for appendiceal epithelial neoplasms and pseudomyxoma peritonei syndrome? *Lancet Oncol* 2006;7:69–76.
14. Esquivel J, Piso P, Verwaal V, Bachleitner-Hofmann T, Glehen O, Gonzalez-Moreno S, et al. [American Society of Peritoneal Surface Malignancies opinion statement on defining expectations from cytoreductive surgery and hyperthermic intraperitoneal chemotherapy in patients with colorectal cancer.](#) *J Surg Oncol* 2014;110:777–8.
15. Bakrin N, Bereder JM, Decullier E, Classe JM, Msika S, Lorimier G, et al. Peritoneal carcinomatosis treated with cytoreductive surgery and Hyperthermic Intraperitoneal Chemotherapy (HIPEC) for advanced ovarian carcinoma: A French multicentre retrospective cohort study of 566 patients. *Eur J Surg Oncol* 2013;39:1435–43.
16. Sugarbaker PH. Parietal peritonectomy. *Ann Surg Oncol* 2012;19:1250.
17. Sugarbaker P. Cytoreductive surgery & perioperative chemotherapy for peritoneal surface malignancy: Textbook and video atlas. Woodbury, Connecticut, United States: Ciné-Med Publishing, 2012:214.
18. Van Der Speeten K, Stuart OA, Sugarbaker PH. Pharmacology of perioperative intraperitoneal and intravenous chemotherapy in patients with peritoneal surface malignancy. *Surg Oncol Clin N Am* 2012;21:577–97.
19. Sugarbaker PH, Graves T, DeBruijn EA, Cunliffe WJ, Mullins RE, Hull WE, et al. Early postoperative intraperitoneal chemotherapy as an adjuvant therapy to surgery for peritoneal carcinomatosis from gastrointestinal cancer: Pharmacological studies. *Cancer Res* 1990;50:5790–4.
20. Mahteme H, Von Heideman A, Grundmark B, Tholander B, Pahlman L, Glimelius B, et al. [Heterogeneous activity of cytotoxic drugs in patient samples of peritoneal carcinomatosis.](#) *Eur J Surg Oncol* 2008;34:547–52.
21. Cashin PH, Mahteme H, Graf W, Karlsson H, Larsson R, Nygren P. [Activity ex vivo of cytotoxic drugs in patient samples of peritoneal carcinomatosis with special focus on colorectal cancer.](#) *BMC Cancer* 2013;13:435.
22. Levine EA, Blazer DG, 3rd, Kim MK, Shen P, Stewart J, Guy C, et al. Gene expression profiling of peritoneal metastases from appendiceal and colon cancer demonstrates unique biologic signatures and predicts patient outcomes. *J Am Coll Surg* 2012;214:599–606; discussion -7.
23. Fujishima Y, Goi T, Kimura Y, Hirono Y, Katayama K, Yamaguchi A. MUC2 protein expression status is useful in assessing the effects of hyperthermic intraperitoneal chemotherapy for peritoneal dissemination of colon cancer. *Int J Oncol* 2012;40:960–4.
24. Stewart J, Shen P, Russell G, Fenstermaker J, McWilliams L, Coldrun FM, et al. [A phase I trial of oxaliplatin for intraperitoneal hyperthermic chemoperfusion for the treatment of peritoneal surface dissemination from colorectal and appendiceal cancers.](#) *Ann Surg Oncol* 2008;15:2137–45.
25. Elias D, Bonnay M, Puizillou JM, Antoun S, Demirdjian S, El OA, et al. [Heated intra-operative intraperitoneal oxaliplatin after complete resection of peritoneal carcinomatosis: Pharmacokinetics and tissue distribution.](#) *Ann Oncol* 2002;13:267–72.
26. Elias D, Raynard B, Bonnay M, Pocard M. [Heated intra-operative intraperitoneal oxaliplatin alone and in combination with intraperitoneal irinotecan: Pharmacologic studies.](#) *Eur J Surg Oncol* 2006;32:607–13.
27. Elias D, El Otmany A, Bonnay M, Paci A, Ducreux M, Antoun S, et al. [Human pharmacokinetic study of heated intraperitoneal oxaliplatin in increasingly hypotonic solutions after complete resection of peritoneal carcinomatosis.](#) *Oncology* 2002;63:346–52.
28. Pomel C, Ferron G, Lorimier G, Rey A, Lhomme C, Classe JM, et al. Hyperthermic intra-peritoneal chemotherapy using oxaliplatin as consolidation therapy for advanced epithelial ovarian carcinoma. Results of a phase II prospective multicentre trial. CHIPOVAC study. *Eur J Surg Oncol* 2010;36:589–93.
29. Chalret Du Rieu Q, White-Koning M, Picaud L, Lochon I, Marsili S, Gladieff L, et al. Population pharmacokinetics of peritoneal, plasma ultrafiltrated and protein-bound oxaliplatin concentrations in patients with disseminated peritoneal cancer after intraperitoneal hyperthermic chemoperfusion of oxaliplatin following cytoreductive surgery: Correlation between oxaliplatin exposure and thrombocytopenia. *Cancer Chemother Pharmacol* 2014;74:571–82.
30. Charrier T, Passot G, Peron J, Maurice C, Gocevska S, Quenet F, et al. [Cytoreductive surgery combined with hyperthermic intraperitoneal chemotherapy with oxaliplatin increases the risk of postoperative hemorrhagic complications: Analysis of predictive factors.](#) *Ann Surg Oncol* 2016;23:2315–22.

31. De Somer F, Ceelen W, Delanghe J, De Smet D, Vanackere M, Pattyn P, et al. Severe hyponatremia, hyperglycemia, and hyperlactatemia are associated with intraoperative hyperthermic intraperitoneal chemoperfusion with oxaliplatin. *Perit Dial Int* 2008;28:61–6.
32. Jerremalm E, Hedeland M, Wallin I, Bondesson U, Ehrsson H. [Oxaliplatin degradation in the presence of chloride: Identification and cytotoxicity of the monochloro monooxalato complex.](#) *Pharm Res* 2004;21:891–4.
33. Mehta AM, Van Den Hoven JM, Rosing H, Hillebrand MJ, Nuijen B, Huitema AD, et al. Stability of oxaliplatin in chloride-containing carrier solutions used in hyperthermic intraperitoneal chemotherapy. *Int J Pharm* 2015;479:23–7.
34. Jacquet P, Averbach A, Stephens AD, Stuart OA, Chang D, Sugarbaker PH. Heated intraoperative intraperitoneal mitomycin C and early postoperative intraperitoneal 5-fluorouracil: Pharmacokinetic studies. *Oncology* 1998;55:130–8.
35. Fujita T, Tamura T, Yamada H, Yamamoto A, Muranishi S. Pharmacokinetics of mitomycin C (MMC) after intraperitoneal administration of MMC-gelatin gel and its anti-tumor effects against sarcoma-180 bearing mice. *J Drug Target* 1997;4:289–96.
36. Van Der Speeten K, Stuart OA, Chang D, Mahteme H, Sugarbaker PH. Changes induced by surgical and clinical factors in the pharmacology of intraperitoneal mitomycin C in 145 patients with peritoneal carcinomatosis. *Cancer Chemother Pharmacol* 2011;68:147–56.
37. Mohamed F, Cecil T, Moran B, Sugarbaker P. A new standard of care for the management of peritoneal surface malignancy. *Curr Oncol* 2011;18:e84–96.
38. Levine EA, Stewart J, Shen P, Russell GB, Loggie BL, Votanopoulos KI. Intraperitoneal chemotherapy for peritoneal surface malignancy: Experience with 1,000 patients. *J Am Coll Surg* 2014;218:573–85.
39. Barlogie B, Corry PM, Drewinko B. In vitro thermochemotherapy of human colon cancer cells with cis-dichlorodiammineplatinum (II) and mitomycin C. *Cancer Res* 1980;40:1165–8.
40. De Bree E, Tsiftsis DD. Experimental and pharmacokinetic studies in intraperitoneal chemotherapy: From laboratory bench to bedside. *Recent Results Cancer Res* 2007;169:53–73.
41. Witkamp A. Dose finding study of hyperthermic intraperitoneal chemotherapy with mitomycin C in patients with carcinosis of colorectal origin. *Eur J Surg Oncol* 1998;136:699–708.
42. Turaga K, Levine E, Barone R, Sticca R, Petrelli N, Lambert L, et al. Consensus guidelines from The American Society of Peritoneal Surface Malignancies on standardizing the delivery of hyperthermic intraperitoneal chemotherapy (HIPEC) in colorectal cancer patients in the United States. *Ann Surg Oncol* 2014;21:1501–5.
43. Dedrick RL, Myers CE, Bungay PM, DeVita VT, Jr.. Pharmacokinetic rationale for peritoneal drug administration in the treatment of ovarian cancer. *Cancer Treat Rep* 1978;62:1–11.
44. Rubin J, Clawson M, Planch A, Jones Q. [Measurements of peritoneal surface area in man and rat.](#) *Am J Med Sci* 1988;295:453–8.
45. Elias DM, Sideris L. Pharmacokinetics of heated intraoperative intraperitoneal oxaliplatin after complete resection of peritoneal carcinomatosis. *Surg Oncol Clin N Am* 2003;12:755–69, xiv.
46. Sugarbaker PH, Stuart OA, Carmignani CP. [Pharmacokinetic changes induced by the volume of chemotherapy solution in patients treated with hyperthermic intraperitoneal mitomycin C.](#) *Cancer Chemother Pharmacol* 2006;57:703–8.
47. Mas-Fuster MI, Ramon-Lopez A, Nalda-Molina R. Importance of standardizing the dose in hyperthermic intraperitoneal chemotherapy (HIPEC): A pharmacodynamic point of view. *Cancer Chemother Pharmacol* 2013;72:273–4.
48. Los G, Mutsaers PH, Van Der Vijgh WJ, Baldew GS, De Graaf PW, McVie JG. Direct diffusion of cis-diamminedichloroplatinum(II) in intraperitoneal rat tumors after intraperitoneal chemotherapy: A comparison with systemic chemotherapy. *Cancer Res* 1989;49:3380–4.
49. Conti M, De Giorgi U, Tazzari V, Bezzi F, Baccini C. [Clinical pharmacology of intraperitoneal cisplatin-based chemotherapy.](#) *J Chemother* 2004;16:23–5.
50. Gladieff L, Chatelut E, Dalenc F, Ferron G. Pharmacological bases of intraperitoneal chemotherapy. *Bull Cancer* 2009;96:1235–42.
51. Coccolini F, Cotte E, Glehen O, Lotti M, Poiasina E, Catena F, et al. Intraperitoneal chemotherapy in advanced gastric cancer. Meta-analysis of randomized trials. *Eur J Surg Oncol* 2014;40:12–26.
52. Alberts DS, Liu PY, Hannigan EV, O'Toole R, Williams SD, Young JA, et al. [Intraperitoneal cisplatin plus intravenous cyclophosphamide versus intravenous cisplatin plus intravenous cyclophosphamide for stage III ovarian cancer.](#) *N Engl J Med* 1996;335:1950–5.
53. Armstrong DK, Bundy B, Wenzel L, Huang HQ, Baergen R, Lele S, et al. [Intraperitoneal cisplatin and paclitaxel in ovarian cancer.](#) *N Engl J Med* 2006;354:34–43.
54. Zivanovic O, Abramian A, Kullmann M, Fuhrmann C, Coch C, Hoeller T, et al. HIPEC ROC I: A phase I study of cisplatin administered as hyperthermic intraoperative intraperitoneal chemoperfusion followed by postoperative intravenous platinum-based chemotherapy in patients with platinum-sensitive recurrent epithelial ovarian cancer. *Int J Cancer* 2014;136(3): 699–708.
55. Hakeam HA, Breakiet M, Azzam A, Nadeem A, Amin T. The incidence of cisplatin nephrotoxicity post hyperthermic intraperitoneal chemotherapy (HIPEC) and cytoreductive surgery. *Ren Fail* 2014;36:1486–91.
56. Urano M, Kuroda M, Nishimura Y. [For the clinical application of thermochemotherapy given at mild temperatures.](#) *Int J Hyperthermia* 1999;15:79–107.
57. Van Der Speeten K, Stuart OA, Mahteme H, Sugarbaker PH. Pharmacokinetic study of perioperative intravenous Ifosfamide. *Int J Surg Oncol* 2011;2011:185092.
58. Deraco M, Baratti D, Cabras AD, Zaffaroni N, Perrone F, Villa R, et al. [Experience with peritoneal mesothelioma at the Milan National Cancer Institute.](#) *World J Gastrointest Oncol* 2010;2:76–84.
59. Tritton TR. [Cell surface actions of adriamycin.](#) *Pharmacol Ther* 1991;49:293–309.
60. Tritton TR, Yee G. The anticancer agent adriamycin can be actively cytotoxic without entering cells. *Science* 1982;217:248–50.
61. Lane P, Vichi P, Bain DL, Tritton TR. Temperature dependence studies of adriamycin uptake and cytotoxicity. *Cancer Res* 1987;47:4038–42.
62. Van Der Speeten K, Stuart OA, Mahteme H, Sugarbaker PH. A pharmacologic analysis of intraoperative intracavitary cancer chemotherapy with doxorubicin. *Cancer Chemother Pharmacol* 2009;63:799–805.
63. Ozols RF, Young RC, Speyer JL, Sugarbaker PH, Greene R, Jenkins J, et al. Phase I and pharmacological studies of

- adriamycin administered intraperitoneally to patients with ovarian cancer. *Cancer Res* 1982;42:4265–9.
64. Ozols RF, Locker GY, Doroshow JH, Grotzinger KR, Myers CE, Young RC. Pharmacokinetics of adriamycin and tissue penetration in murine ovarian cancer. *Cancer Res* 1979;39:3209–14.
 65. Ozols RF, Locker GY, Doroshow JH, Grotzinger KR, Myers CE, Fisher RI, et al. Chemotherapy for murine ovarian cancer: A rationale for ip therapy with adriamycin. *Cancer Treat Rep* 1979;63:269–73.
 66. Nagai K, Nogami S, Egusa H, Konishi H. Pharmacokinetic evaluation of intraperitoneal doxorubicin in rats. *Pharmazie* 2014;69:125–7.
 67. Harrison LE, Bryan M, Pliner L, Saunders T. [Phase I trial of pegylated liposomal doxorubicin with hyperthermic intraperitoneal chemotherapy in patients undergoing cytoreduction for advanced intra-abdominal malignancy.](#) *Ann Surg Oncol* 2008;15:1407–13.
 68. Salvatorelli E, De Tursi M, Menna P, Carella C, Massari R, Colasante A, et al. [Pharmacokinetics of pegylated liposomal doxorubicin administered by intraoperative hyperthermic intraperitoneal chemotherapy to patients with advanced ovarian cancer and peritoneal carcinomatosis.](#) *Drug Metab Dispos* 2012;40:2365–73.
 69. Van Der Speeten K, Stuart OA, Mahteme H, Sugarbaker PH. Pharmacology of perioperative 5-fluorouracil. *J Surg Oncol* 2010;102:730–5.
 70. Glehen O, Kwiatkowski F, Sugarbaker PH, Elias D, Levine EA, De Simone M, et al. [Cytoreductive surgery combined with perioperative intraperitoneal chemotherapy for the management of peritoneal carcinomatosis from colorectal cancer: A multi-institutional study.](#) *J Clin Oncol* 2004;22:3284–92.
 71. Elias D, Benizri E, Di Pietrantonio D, Menegon P, Malka D, Raynard B. [Comparison of two kinds of intraperitoneal chemotherapy following complete cytoreductive surgery of colorectal peritoneal carcinomatosis.](#) *Ann Surg Oncol* 2007;14:509–14.
 72. McConnell YJ, Mack LA, Francis WP, Ho T, Temple WJ. [HIPEC + EPIC versus HIPEC-alone: Differences in major complications following cytoreduction surgery for peritoneal malignancy.](#) *J Surg Oncol* 2013;107:591–6.
 73. Tan GH, Ong WS, Chia CS, Tham CK, Soo KC, Teo MC. Does early post-operative intraperitoneal chemotherapy (EPIC) for patients treated with cytoreductive surgery and hyperthermic intraperitoneal chemotherapy (HIPEC) make a difference?. *Int J Hyperthermia* 2016;32:281–8.
 74. Heidelberger C, Chaudhuri NK, Danneberg P, Mooren D, Griesbach L, Duschinsky R, et al. [Fluorinated pyrimidines, a new class of tumour-inhibitory compounds.](#) *Nature* 1957;179:663–6.
 75. Muggia FM, Peters GJ, Landolph JR, Jr.. XIII International Charles Heidelberger Symposium and 50 years of fluoropyrimidines in cancer therapy held on September 6 to 8, 2007 at New York University Cancer Institute, Smilow Conference Center. *Mol Cancer Ther* 2009;8:992–9.
 76. Wagner PL, Jones D, Aronova A, Shia J, Weiser MR, Temple LK, et al. [Early postoperative intraperitoneal chemotherapy following cytoreductive surgery for appendiceal mucinous neoplasms with isolated peritoneal metastasis.](#) *Dis Colon Rectum* 2012;55:407–15.
 77. Yu W, Whang I, Chung HY, Averbach A, Sugarbaker PH. Indications for early postoperative intraperitoneal chemotherapy of advanced gastric cancer: Results of a prospective randomized trial. *World J Surg* 2001;25:985–90.
 78. Kwon OK, Chung HY, Yu W. [Early postoperative intraperitoneal chemotherapy for macroscopically serosa-invading gastric cancer patients.](#) *Cancer Res Treat* 2014;46:270–9.
 79. Passot G, Vaudoyer D, Villeneuve L, Kepenekian V, Beaujard AC, Bakrin N, et al. What made hyperthermic intraperitoneal chemotherapy an effective curative treatment for peritoneal surface malignancy: A 25-year experience with 1,125 procedures. *J Surg Oncol* 2016;113:796–803.
 80. Sugarbaker PH, Mora JT, Carmignani P, Stuart OA, Yoo D. [Update on chemotherapeutic agents utilized for perioperative intraperitoneal chemotherapy.](#) *Oncologist* 2005;10:112–22.
 81. Rohena CC, Mooberry SL. [Recent progress with microtubule stabilizers: New compounds, binding modes and cellular activities.](#) *Nat Prod Rep* 2014;31:335–55.
 82. De Bree E, Theodoropoulos PA, Rosing H, Michalakis J, Romanos J, Beijnen JH, et al. [Treatment of ovarian cancer using intraperitoneal chemotherapy with taxanes: From laboratory bench to bedside.](#) *Cancer Treat Rev* 2006;32:471–82.
 83. Mohamed F, Sugarbaker PH. [Intraperitoneal taxanes.](#) *Surg Oncol Clin N Am* 2003;12:825–33.
 84. Yonemura Y, Bandou E, Sawa T, Yoshimitsu Y, Endou Y, Sasaki T, et al. [Neoadjuvant treatment of gastric cancer with peritoneal dissemination.](#) *Eur J Surg Oncol* 2006;32:661–5.
 85. Sugarbaker PH. Treatment of peritoneal carcinomatosis from colon or appendiceal cancer with induction intraperitoneal chemotherapy. *Cancer Treat Res* 1996;82:317–25.
 86. Zylberberg B, Dormont D, Jankiewicz S, Darai E, Bretel JJ, Poncelet C, et al. Response to neo-adjuvant intraperitoneal and intravenous immunochemotherapy followed by interval secondary cytoreduction in stage IIIc ovarian cancer. *Eur J Gynaecol Oncol* 2001;22:40–5.
 87. Yonemura Y, Elnemr A, Endou Y, Ishibashi H, Mizumoto A, Miura M, et al. Effects of neoadjuvant intraperitoneal/systemic chemotherapy (bidirectional chemotherapy) for the treatment of patients with peritoneal metastasis from gastric cancer. *Int J Surg Oncol* 2012;2012:148420.
 88. Esquivel J, Vidal-Jove J, Steves MA, Sugarbaker PH. Morbidity and mortality of cytoreductive surgery and intraperitoneal chemotherapy. *Surgery* 1993;113:631–6.