# Electrochemical sensor platform for MIP incorporated microfluidic paper-based analytical devices

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**Abstract:** A biomimetic sensor platform capable of performing electrochemical measurements for the detection of molecules was developed by incorporating molecularly imprinted polymers (MIPs) into microfluidic paper-based analytical devices( $\mu$ PADs). The usage of  $\mu$ PADs in combination with electrochemical measurements vastly improves several parameters compared to lab-sized electrochemical setups. Interdigitated electrodes were utilized, this way no top electrode is needed. The amplitude of the impedance was monitored and a complete impedance spectrum was measured continuously. The combination of low-cost, disposable paper-based microfluidics and electrochemical read-out, make an ideal point-of-care application.

**Keywords:** impedimetric, electrochemical, microfluidics, microfluidic paper-based analytical device(µPAD), molecularly imprinted polymer(MIP)

# Introduction

Thoelen R. et al. showed that platforms capable of performing electrochemical measurements are suitable for the detection of molecules by applying molecularly imprinted polymers[1]. Impedimetric setups have since than been employed for different kinds of molecules in different kinds of fluids[2].

The use of paper-based microfluidics further miniaturizes the electrochemical read-out platform. Hereby some parameters (i.e. temperature[2]) are no longer controlled while others are vastly improved.

Firstly, paper-based microfluidics do not require actively driven pumps. Indeed, capillary forces of the paper fibres cause fluid to flow. In addition, paper-based microfluidics use less fluid which in turn leads to less stabilization time. This severely decreases the duration of a measurement. Paperbased microfluidics also enable sample pretreatment because the pore size distribution filters out the unwanted larger molecules.

# **Materials & Methods**

Whatman nr.1 was used as paper. Its design was cut-out with a  $CO_2$ -laser. It has an addition zone, a running zone which contains the non-imprinted polymers(NIP) and MIP, and a pumping zone which provides a sustained flow over time[3]. The paper is sealed on one side with Kapton tape. The edges of this tape are exploited to adhere the  $\mu$ PAD to the interdigitated electrodes. A schematic side view of the  $\mu$ PAD can be seen in Figure 1.

The amplitude of the impedance is observed for a complete spectrum. Since the NIP provides a negative control no non-spiked solution step is needed.

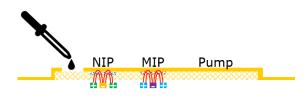


Figure 1: Schematic side view of the µPAD

# **Results and Discussion**

 $|Z_{MIP}|$ - $|Z_{NIP}|$  is trending to the positive. It should be noted that only molecules that have past the MIP region will give rise to a higher differential signal. Therefore increasing the pump capacity could potentially boost the signal.

# Conclusion

Although  $\mu$ PADs give rise to a lower signal it is yet the smallest biomimetic sensor platform based on imprinted polymers and electrochemical read-out. Therefore it is the most promising step towards a point-of-care device.

### References

[1] Thoelen R. et al., Biosensors and Bioelectronics, 2008, Vol. 23, pp 913-918

[2] Peeters M. et al., Sensors and Actuators B, 2012, Vol. 171-172, pp 302-610

[3] Mendez S. et al., LANGMUIR, 2010, Vol. 26, pp 1380-1385

### Acknowledgements

This work is funded by the BIOMAT project which is carried out under Interreg V-A grensregio Vlaanderen - Nederland and is supported by the European Union and The European Regional Development Fund and with financial support of province of Limburg - Belgium.

