



Draft Genome Sequence of *Bacillus pumilus* SCAL1, an Endophytic Heat-Tolerant Plant Growth-Promoting Bacterium

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ABSTRACT *Bacillus pumilus* strain SCAL1 is an endophytic, thermophilic plant that was isolated from the leaf of a plant, *Solanum lycopersicum* L., in Sindh, Pakistan. *B. pumilus* strain SCAL1 has usually exhibited high resistance to environmental stresses, with a growth temperature ranging from 30 to 60°C. An approximately 3.75-Mb draft genome was assembled into 68 contigs.

Bacillus pumilus forms endospores and naturally occurs in roots, leaves, seeds, stems, needles, twigs, and bark of various plant species (1–4). In this context, plant growth-promoting bacteria improve plant growth either (i) directly, through production of phytohormones, ammonia, siderophores, or enzymes (ACC deaminase and catalase), or by providing nitrogen, phosphate, or iron, or (ii) indirectly, by antagonistic mechanisms which protect plants from pathogens and insects. Certain endophytes generate induced systemic tolerance (IST), through which plants are protected from abiotic stressors and allowed to attenuate their negative effects (3–6). Bacterial spores show resistance to radiation, desiccation, and hydrogen peroxide treatment (7, 8).

After sample collection, surface sterilization was done (1). Strain taxonomy was based on 16S rRNA gene sequencing (2, 7, 9). Two milliliters of cell culture broth was taken and centrifuged at $12,000 \times g$ for 5 minutes. RNA-free genomic DNA was extracted with an Invitrogen PureLink genomic DNA minikit and quantified using a Qubit double-stranded DNA (dsDNA) HS assay kit (Life Technologies, Inc., Burlington, ON). Adaptor-ligated DNA was size selected to 480 bp on a 2% E-Gel SizeSelect agarose gel, and Agencourt MAPure XP beads (Beckman Coulter, Mississauga, ON) were used for purification. Library dilution factor was determined using an Ion Library quantitation kit prior to amplification and enrichment with an Ion PGM template OT2 400 kit on an Ion OneTouch 2 system. The enriched Ion Sphere particles were quantified using an Ion Sphere quality control kit prior to sequencing on a 316 version 2 chip with an Ion PGM 400 sequencing kit on an Ion Torrent PGM. Sequencing generated 590,303 reads (mean length, 256 bases) and 151 Mb of data (>127 million Q20 bases) in Torrent Suite 4.4.3. Further, assembly was performed using SPAdes 3.1.0 (10, 11) with trimming by custom script Trim_SPAdes into 68 contigs greater than 500 bp, giving a consensus length of 3,743,482 bp at $21\times$ coverage (largest contig, 371,528 bp; N_{50} , 112,210 bp) with a GC content of 41.4%. The Rapid Annotations using Subsystem Technology (RAST) server (8, 12, 13) was used to find closely related strains before using progressiveMauve version 2.3.1 (14) to reorder scaffolds with the use of *B. pumilus* SAFR-032 (NCBI reference

Received 21 March 2018 Accepted 24 March 2018 Published 3 May 2018

Citation Mukhtar T, Afridi MS, McArthur R, Van Hamme JD, Rineau F, Mahmood T, Amna, Sumaira, Zahid M, Salam A, Khan MN, Ali F, Mehmood S, Bangash N, Chaudhary HJ. 2018. Draft genome sequence of *Bacillus pumilus* SCAL1, an endophytic heat-tolerant plant growth-promoting bacterium. Genome Announc 6:e00306-18. <https://doi.org/10.1128/genomeA.00306-18>.

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sequence NC_009848) (15) as a reference. Open reading frame (ORF) prediction and gene annotation were done by using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) pipeline (16, 17). Gene search and annotation were performed for all contigs longer than 500 bp using the NCBI Prokaryotic Genomes Automatic Annotation Pipeline and the RAST server (8). The SEED viewer was used for assignment of predicted genes to functional categories (18). The RAST server predicted 3,327 coding sequences (CDS) with a total of approximately 3,802 genes, 400 pseudogenes, 57 tRNAs, and 1 noncoding RNA (ncRNA) gene and 17 coded rRNAs (5S, 16S, and 23S). Genome-wide phylogenetic analysis of various *Bacillus* species compared to the *B. pumilus* SCAL1 genome showed that it clusters closely to *B. pumilus* SAFR-032. Genome analysis revealed the genes for heat shock resistance, hydrocarbon metabolism, heavy metal tolerance, biofilm formation, and siderophore and IAA biosynthesis.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number [LFIZ00000000](https://doi.org/10.1093/bioinformatics/btt086) (version LFIZ01000000).

ACKNOWLEDGMENTS

This research was supported by Department of Biological Sciences, Thompson Rivers University, Canada.

We thank Jonathan D. Van Hamme, Department of Biological Sciences, Thompson Rivers University, for sequencing *B. pumilus* SCAL1 and assisting with data analysis. We thank Rachel Backer, Department of Plant Science, McGill University, for assistance with English language revisions.

REFERENCES

- Haruna E, Zin NM, Kerfahi D, Adams JM. 2018. Extensive overlap of tropical rainforest bacterial endophytes between soil, plant parts, and plant species. *Microb Ecol* 75:88–103. <https://doi.org/10.1007/s00248-017-1002-2>.
- Zachow C, Pirker H, Westendorf C, Tilcher R, Berg G. 2009. The *Caenorhabditis elegans* assay: a tool to evaluate the pathogenic potential of bacterial biocontrol agents. *Eur J Plant Pathol* 125:367–376. <https://doi.org/10.1007/s10658-009-9486-3>.
- Stone JK, Bacon CW, White JF. 2000. An overview of endophytic microbes: endophytism defined, p 29–33. *In* *Microbial endophytes*. Marcel Dekker, New York, NY.
- Sturz AV, Christie BR, Nowak J. 2000. Bacterial endophytes: potential role in developing sustainable systems of crop production. *Crit Rev Plant Sci* 19:1–30. <https://doi.org/10.1080/07352680091139169>.
- Jacobs MJ, Bugbee WM, Gabrielson DA. 1985. Enumeration, location, and characterization of endophytic bacteria within sugar beet roots. *Can J Bot* 63:1262–1265. <https://doi.org/10.1139/b85-174>.
- Larran S, Monaco C, Alippi HE. 2000. Endophytic fungi in beet (*Beta vulgaris* var. *esculenta* L.) leaves. *Adv Hort Sci* 14:193–196. <http://www.jstor.org/stable/42883275>.
- Lottmann J, Heuer H, Smalla K, Berg G. 1999. Influence of transgenic T4-lysozyme-producing potato plants on potentially beneficial plant-associated bacteria. *FEMS Microbiol Ecol* 29:365–377. <https://doi.org/10.1111/j.1574-6941.1999.tb00627.x>.
- Iyer R, Damania A, Iken B. 2017. Whole genome sequencing of *Microbacterium* sp. AISO3 from polluted San Jacinto river sediment reveals high bacterial mobility, metabolic versatility and heavy metal resistance. *Genom Data* 14:0–13.
- Loper JE, Hassan KA, Mavrodi DV, Davis IIEW, Lim CK, Shaffer BT, Elbourne LD, Stockwell VO, Hartney SL, Breakwell K, Henkels MD. 2012. Comparative genomics of plant-associated *Pseudomonas* spp.: insights into diversity and inheritance of traits involved in multitrophic interactions. *PLoS Genet* 8:1002784. <https://doi.org/10.1371/journal.pgen.1002784>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formisano K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci Rep* 5:8365. <https://doi.org/10.1038/srep08365>.
- Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. *PLoS One* 5:11147. <https://doi.org/10.1371/journal.pone.0011147>.
- Tirumalai MR, Fox GE. 2013. An ICEBs1-like element may be associated with the extreme radiation and desiccation resistance of *Bacillus pumilus* SAFR-032 spores. *Extremophiles* 17:767–774. <https://doi.org/10.1007/s00792-013-0559-z>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity GM, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of standard operating procedures (SOPs) for (meta) genomic annotation. *OMICS* 12:137–141. <https://doi.org/10.1089/omi.2008.0017>.
- Kadnikov VV, Mardanov AV, Poltarau AB, Sokolova DS, Semenova EM, Ravin NV, Tourova TP, Nazina TN. 2017. Genome sequencing and annotation of *Geobacillus* sp. 1017, a hydrocarbon-oxidizing thermophilic bacterium isolated from a heavy oil reservoir (China). *Genom Data* 11:95–97. <https://doi.org/10.1016/j.gdata.2016.12.011>.