

Increased telomere length and mtDNA copy number induced by
multi-walled carbon nanotube exposure in the workplace

Peer-reviewed author version

Ghosh, Manosij; Janssen, Lisa; MARTENS, Dries; Öner, Deniz; Vlaanderen, Jelle; Pronk, Anjoeka; Kuijpers, Eelco; Vermeulen, Roel; NAWROT, Tim; Godderis, Lode & Hoet, Peter (2020) Increased telomere length and mtDNA copy number induced by multi-walled carbon nanotube exposure in the workplace. In: JOURNAL OF HAZARDOUS MATERIALS, 394 (Art N° 122569).

DOI: 10.1016/j.jhazmat.2020.122569

Handle: <http://hdl.handle.net/1942/30993>

1 Increased telomere length and mtDNA copy number induced by multi-walled 2 carbon nanotube exposure in the workplace

3

4 Manosij Ghosh^{1*}, Lisa Janssen^{1*}, Dries S. Martens^{1,2*}, Deniz Öner¹, Jelle Vlaanderen³, Anjoeka
5 Pronk⁴, Eelco Kuijpers⁴, Roel Vermeulen³, Tim S. Nawrot^{1,2}, Lode Godderis^{1,5#}, Peter HM Hoet^{1#}

6

7 ¹ Department of Public Health and Primary Care, Centre Environment & Health, KU Leuven, Leuven, Belgium;

8 ² Centre for Environmental Sciences, Hasselt University, Hasselt, Belgium;

9 ³ Division of Environmental Epidemiology, Institute for Risk Assessment Sciences, Utrecht University, Utrecht,
10 The Netherlands;

11 ⁴ TNO, Netherlands Organisation for Applied Scientific Research, Zeist, The Netherlands

12 ⁵ External Service for Prevention and Protection at Work, Idewe, Heverlee, Belgium

13

14 * MG, LJ and DSM have equal contribution.

15 # Joint last authors, Corresponding authors

16 (Lode Godderis: lode.godderis@kuleuven.be; Peter Hoet: peter.hoet@kuleuven.be)

17 **Abstract:** Carbon nanotubes (CNTs) – except MWCNT-7 - have been classified as Group 3 [
18 “*Not classifiable as to its carcinogenicity to humans*”] by the IARC. Despite considerable
19 mechanistic evidence *in vitro/ in vivo*, the classification highlights a general lack of data,
20 especially among humans. In our previous study, we reported epigenetic changes in the
21 MWCNT exposed workers. Here, we evaluated whether MWCNT can also cause alterations in
22 aging related features including relative telomere length (TL) and/or mitochondrial copy
23 number (mtDNA_{cn}). Relative TL and mtDNA_{cn} were measured on extracted DNA from
24 peripheral blood from MWCNT exposed workers ($N = 24$) and non-exposed controls ($N = 43$)
25 using a qPCR method. A higher mtDNA_{cn} and longer TL were observed in MWCNT exposed
26 workers when compared to controls. Independent of age, sex, smoking behavior, alcohol
27 consumption and BMI, MWCNT-exposure was associated with an 18.30 % increase in blood
28 TL (95% CI: 7.15 to 30.62 %; $p = 0.001$) and 35.21 % increase in mtDNA_{cn} (95% CI: 19.12 to
29 53.46 %). Our results suggest that exposure to MWCNT can induce an increase in the mtDNA_{cn}
30 and TL; however, the mechanistic basis or consequence of such change requires further
31 experimental studies.

32 **Keywords:** nanotoxicology; carbon nanotubes; occupational exposure; telomere length;
33 mitochondrial DNA

34

1 **1. Introduction**

2 Carbon nanotubes (CNTs) with its specific electrical and thermal properties, and unique
3 mechanical properties, have a great potential for commercialization. On the other hand, there
4 are concerns about its effect on worker/consumer health from exposure during
5 manufacturing/handling and the use of consumer products. Studies thus far have established
6 the toxicity and potential carcinogenicity of several forms of CNTs, in *vitro* and in rodent
7 models. Since no human cancer data are available, the International Agency for Research on
8 Cancer (IARC) focused on these results assessing the mechanism of toxicity and
9 carcinogenicity of single-walled (SWCNT) and multi-walled (MWCNT) carbon nanotubes.
10 Based on these studies, a particular rigid MWCNT, namely Mitsui 7 (MWCNT-7), was classified
11 as *Group 2B (possibly carcinogenic to humans)* [1]. Other types of CNTs (MWCNT/ SWCNT)
12 were categorized into *Group 3*, which means they are not classifiable as to their
13 carcinogenicity to humans. The mechanistic data regarding end-points related to lung cancer
14 and mesothelioma, are too limited to draw conclusions [1].

15 Only recently, some epidemiological studies, mostly cross sectional in nature, have
16 started providing evidence on the early biological effects of CNT in humans. A cross-sectional
17 study by Beard et al. [2], conducted in the US in a large group of workers ($N = 108$), associated
18 elevated blood and sputum biomarkers, like IL-18, fibrinogen, endothelin-1 and different
19 metalloproteinases, with both exposure to CNTs and nanofibres. The conclusion was that
20 inhalable rather than respirable CNTs were more consistently associated with biomarkers of
21 fibrosis, inflammation, oxidative stress. Another study by Fatkhutdinova et al. [3], revealed an
22 increase in fibrotic markers and inflammatory cytokines, in biofluids of a small group ($N = 10$)
23 of MWCNT exposed workers compared to controls. In a subsequent study in the same
24 population, Shvedova et al. [4] showed significant changes in expression of several key
25 pathways, reflective of MWCNT-induced toxicity and their potential to trigger pulmonary,
26 cardiovascular, and carcinogenic outcomes in humans. Lee et al. [5] observed and increase in
27 oxidative stress markers in the exhaled breath condensate of exposed workers ($N = 9$). These
28 studies support the results from *in vitro* and animal studies, stating that CNT exposure can
29 induce oxidative stress and inflammation.

30 Vlaanderen et al. [6] conducted a cross-sectional study in a rather small ($N = 22$), but
31 well characterized group of MWCNT exposed workers compared to controls ($N = 39$). They

1 observed increase in immune markers including basic fibroblast growth factor, and soluble IL-
2 1 receptor II in MWCNT exposed workers. Based on the same set of workers Kuijpers et al., [7]
3 observed an increase in a cardiovascular biomarker (endothelial damage marker intercellular
4 adhesion molecule-1), associated with MWCNT exposure. Furthermore, Ghosh et al. [8] found
5 differences in gene-specific DNA methylation promotor CpGs for different genes, e.g. ATM
6 and HDAC4 in the same population.

7 Mitochondria, being a major source and a target of intracellular reactive oxygen
8 species, mitochondrial DNA (mtDNA) is particularly vulnerable. We estimated mtDNA copy
9 number by measuring the relative levels of unique mtDNA sequences of ND1 (mitochondrial
10 encoded NADH dehydrogenase 1) gene and hmito3 (129-bp fragment) compared to nuclear
11 human β -globin (HBG) gene. Increased mitochondrial biogenesis as an adaptive response to
12 oxidative stress, results in an increase in mtDNA copy number (mtDNAcn; mitochondrial to
13 nuclear genome ratio) [9]. Since ROS play a key role in the regulation of mtDNAcn [10], it
14 serves as a potential biomarker of ROS induced mitochondrial dysfunction [9,11,12]. In
15 addition to mtDNAcn, previous studies have also reported the influence of inflammation and
16 oxidative stress on relative telomere length (TL) [13–16]. Telomeres consist of tandem repeats
17 of DNA (5'-TTAGGG-3'), which play a critical role in chromosome stability and may be affected
18 by environmental and occupational chemicals [17]. Based on previously described evidence
19 that CNT exposure can result in elevated ROS-formation and inflammation, we hypothesize
20 that occupational exposure to MWCNT can influence both mitochondrial function and
21 telomeres, as reflected by the mtDNAcn and TL, respectively.

22 **2. Study Design**

23 The present study was designed to study the effect of MWCNT exposure at workplace
24 on telomere length (TL) and mitochondrial copy number (mtDNAcn) in DNA isolated from
25 peripheral whole blood. This section provides a brief overview of the study population and
26 methods used; which has been described elaborately in the Supplementary section “Materials
27 and Methods”. The study was approved by the Commission for Medical Ethics of UZ Leuven
28 (reference number S54607). Workers ($N= 24$) were recruited from a factory producing
29 MWCNTs commercially and compared to 43 control subjects (no history of MWCNT exposure).
30 Exposure assessment for MWCNT-exposed workers was performed earlier [18,19] and is
31 described in supplementary section “M.1. Study participants and exposure assessment”.

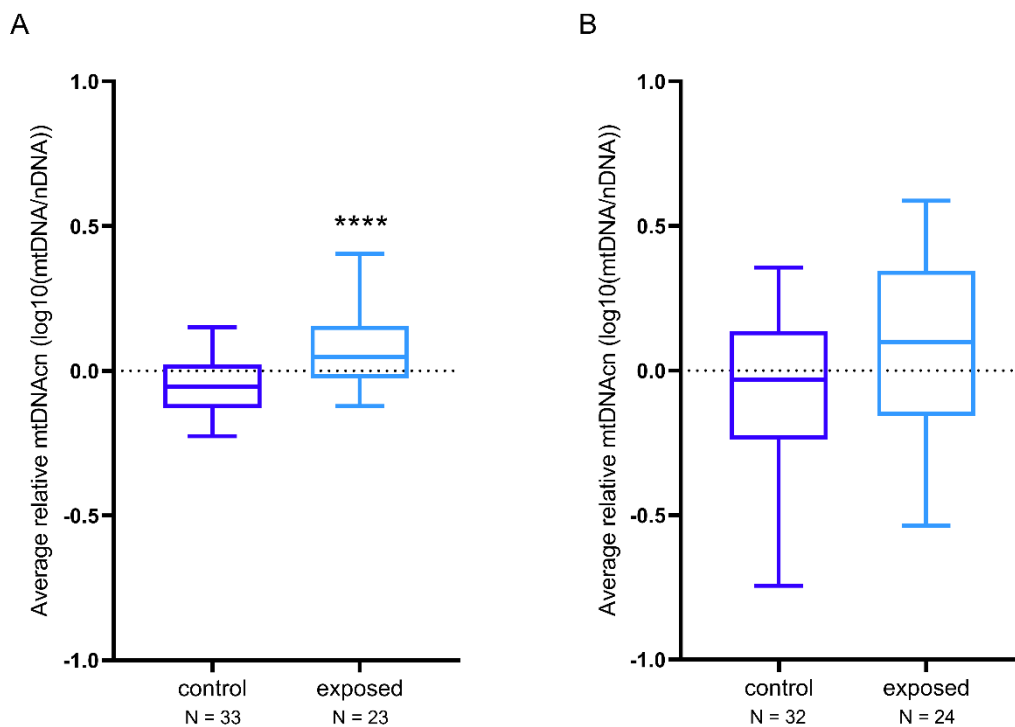
1 Biological sample collection was concluded in 2013, and is previously described by Vlaanderen
2 et al. [6]. The demographic characteristics of the study population are summarized in **Table 1**.
3 mtDNA content [11] and average relative TL [20,21] was measured using quantitative real-
4 time polymerase chain reaction (qPCR) assay, according to methods previously published and
5 is described in supplementary section “M.3. mtDNA copy number and TL assessment by
6 qPCR”.

7 <Table 1>

8 3. Results

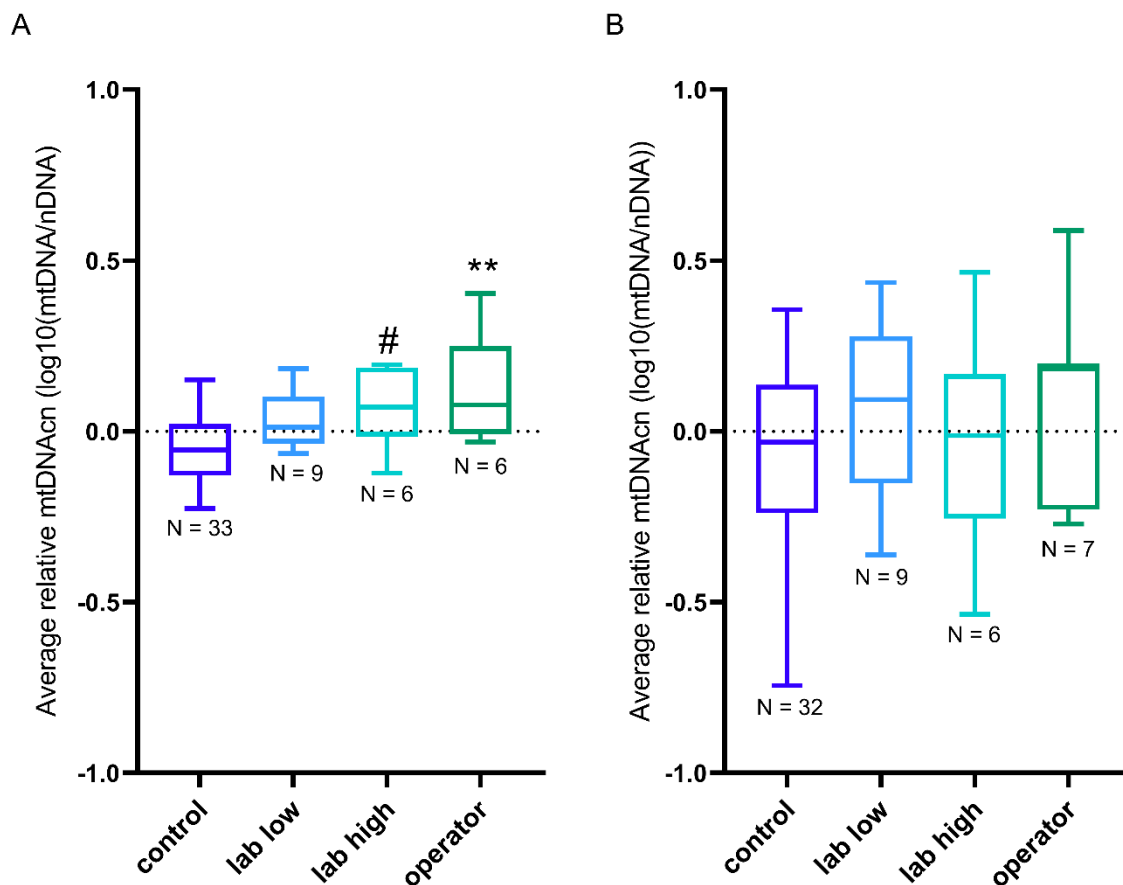
9 3.1. Mitochondrial DNA content

10 In unadjusted analysis a higher mtDNAcn was observed in MWCNT exposed workers
11 compared to controls (**Figure 1A and B**). After adjustment for age, sex, smoking behaviour,
12 alcohol consumption and BMI, a 35.2 % (95% CI: 19.1 to 53.5 %; $p < 0.0001$) higher mtDNAcn
13 was observed in exposed workers, using the ND1 mitochondrial gene (Table 2). When using
14 the mitochondrial hmito3 gene, exposed workers showed a 37.4 % (95% CI: -20.6 to 92.8%; p
15 = 0.068) higher mtDNAc compared to non-exposed individuals.



16
17 **Figure 1:** Box-plots showing the mtDNA content from MWCNT exposed and non-exposed controls, expressed as
18 the log₁₀ of (mtDNA/nDNA) for the two different mitochondrial genes, (A) ND1 and (B) hmito3; **** $p < 0.0001$.
19 P-values based on unpaired t-test between exposed and control.

1
 2 Besides comparing the exposed workers with the non-exposed workers, the association was
 3 examined in different groups of exposure (lab low, lab high and operator) compared with the
 4 non-exposed controls. In unadjusted (Figure 2A) and fully adjusted analysis (**Table 2**),
 5 mtDNA_{Acn}, evaluated using ND1, showed significant differences between all the groups and
 6 the non-exposed controls. A 26.5 % (95% CI: 6.4 to 50.7 %; p = 0.008) difference was found
 7 with the “lab low”-group, a 35.5 % (95% CI: 10.2 to 66.3 %; p = 0.004) difference with the “lab
 8 high”-group and a 40.6 % (95% CI: 13.8 to 73.4 %, p = 0.002) difference with the “operator”-
 9 group when compared to the non-exposed controls. For Hmito3 (**Figure 2B and Table 2**), no
 10 significant differences were found between the different groups and the non-exposed
 11 controls.



12
 13 **Figure 2:** Box-plots showing the average relative mtDNA content (log₁₀(mtDNA/nDNA)) from both mitochondrial
 14 genes, ND1 (A) and Hmito3 (B), for the three different MWCNT exposure groups [lab-low (1 µg/m³ EC), lab-high
 15 (7 µg/m³ EC), and operators (45 µg/m³ EC)] compared with the non-exposed controls. # p < 0.10, *p < 0.05, **p
 16 < 0.01. P-values based on one-way anova between different exposure groups and the control.

3.2. Telomere length

Compared to the non-exposed controls, the MWCNT exposed workers had consistently longer telomeres (Figure 3A and Table 2). Workers exposed to MWCNT had 18.3 % (95% CI: 7.2 to 30.6 %; $p = 0.001$) longer telomeres compared to the non-exposed controls (Table 2). When comparing the different exposure groups (Figure 3B and Table 2), 27.1 % (95% CI: 10.9 to 45.5 %; $p = 0.001$) significantly longer telomeres were observed in the “lab low”-group, compared to the non-exposed controls. In addition, longer telomeres were also observed in the “operator”-group when compared to the non-exposed controls (18.9%; 95% CI: 1.39 to 38.99 %; $p = 0.033$). In the “lab high”-group no significant difference in telomere length was observed when compared to the non-exposed controls.

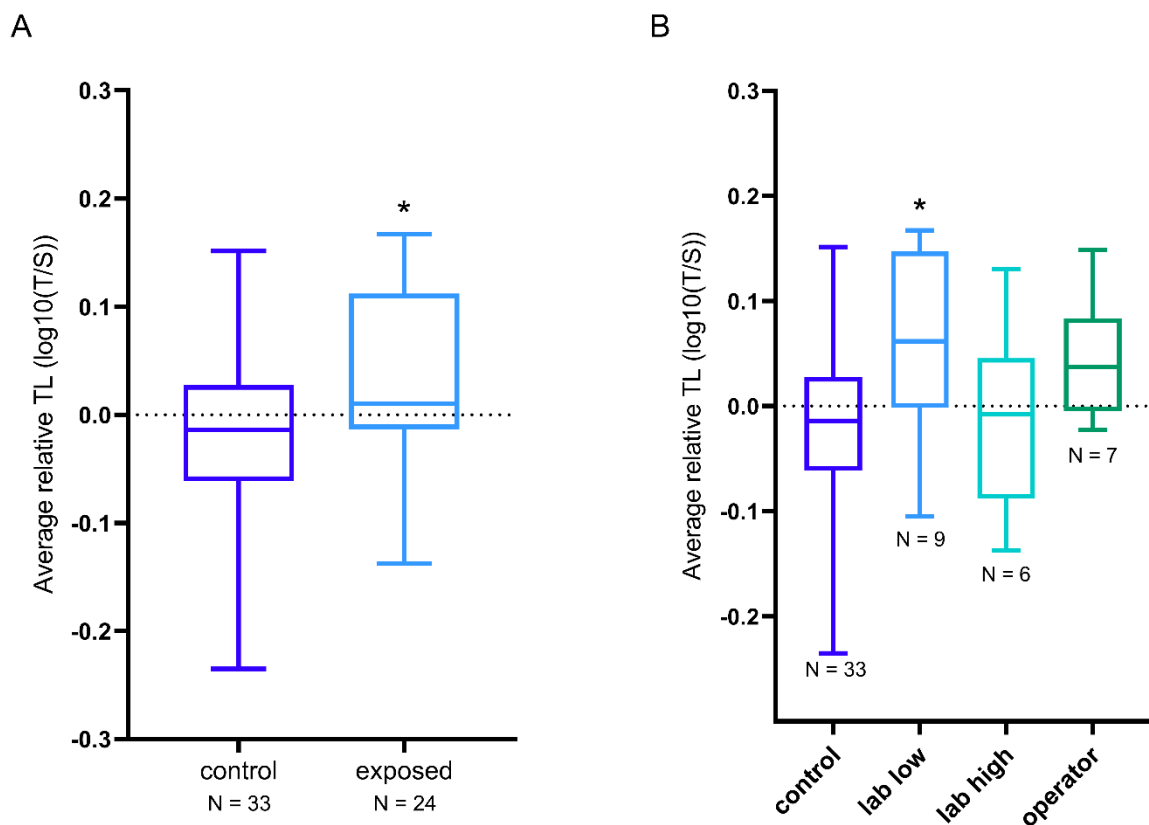


Figure 3: (A) Box-plot showing the log of the average relative TL of MWCNT exposed workers compared to non-exposed controls (B) Box-plot showing the log of the average TL for the three different groups of exposed workers [lab-low ($1 \mu\text{g}/\text{m}^3$ EC), lab-high ($7 \mu\text{g}/\text{m}^3$ EC), and operators ($45 \mu\text{g}/\text{m}^3$ EC)] compared to non-exposed controls; * $p < 0.05$, ** $p < 0.01$, *** < 0.001 . P-values based on unpaired t-test (A) and one-way anova (B) between the different exposure groups and control.

1 **4. Discussion**

2 *4.1. MWCNT exposure is associated with an increase in mtDNAcn*

3 The first key finding of the present study is that MWCNT exposure is significantly
4 associated with an increase in mtDNAcn when compared to non-exposed controls. This is in
5 line with the observed increase in oxidative stress and inflammatory response in several
6 studies [3,4], including the ones reported on the present population [7,18]. In our study, a
7 positive association between MWCNT exposure levels and mtDNAcn was observed, when
8 comparing three different exposure groups (lab low, lab high and operator) with the non-
9 exposed controls. These associations were independent of the effect of age, sex, smoking
10 behaviour, alcohol consumption and BMI.

11 Although the biological mechanism by which environmental exposure can induce an
12 increase in mtDNAcn is still to be revealed, a hypothesis is proposed by Lee et al. [22]. Several
13 studies have shown a close association between an increase in mtDNAcn and DNA damage by
14 ROS and reduced respiratory chain function as a result of oxidative damage [23–26]. As
15 mentioned before, the increase in mtDNAcn is thought to be a compensation for oxidative
16 damage to mtDNA [9]. Although, whether mtDNAcn has a direct role in carcinogenesis and
17 other pathologies/diseases is still under investigation. Concern regarding elevated mtDNAcn
18 has been raised by several studies showing an association between an increased mtDNAcn
19 and several cancers, e.g. head and neck cancer [27], lung cancer [28] and breast cancer [29].
20 Besides that, a decrease in mtDNAcn has been associated with Alzheimer's disease [30].

21 While there are no studies reporting mtDNAcn variation for CNT exposure, mtDNAcn
22 variations have been observed for other exposures. A cross-sectional study by Hou et al. [12]
23 reported higher mtDNAcn, associated with occupational exposure to PM₁, coarse particles
24 (PM_{2.5-10}) and PM₁₀. Another study, conducted by Masayeva et al. [31] has shown an
25 association between cigarette smoking and an increase in mtDNAcn in salivary cells. A study
26 by Tan et al. [32] supports these results. Moreover, Pavanello et al. [33] reported an increase
27 in mtDNAcn as a result of exposure to polycyclic aromatic hydrocarbons (PAHs). Lee et al. [34]
28 has also shown this association between tobacco smoke and mtDNAcn increase in adjacent
29 lung tissues of patients with cancer. However, several other studies reported contrary
30 findings, e.g. a study by Janssen et al. [11] reported an inverse association between air
31 pollution exposure and mtDNAcn in placental tissue and Pieters et al. [35] reported a decrease

1 in mtDNAcn associated with exposure to PAHs. Mitochondria respond dynamically to
2 environmental insults as reflected by these studies, and depending on the exposure, exposure
3 levels, timing, duration of exposure and study design both positive and negative associations
4 are found. These inconsistencies potentially reflect different phases in the mitochondrial
5 response to environmental exposures, in which both damaging and compensating
6 mechanisms are present. Therefore, alternations in mtDNAcn, may present a biological
7 mechanism by which CNTs, or more specific MWCNTs, affect exposed individuals.

8 *4.2. MWCNT exposure is associated with longer telomere length*

9 The second key finding is that humans exposed to MWCNT have significantly longer
10 telomeres, when compared to non-exposed controls. While this is the first study on TL and
11 CNT exposure, some studies observed longer telomeres in relation to other environmental
12 exposures. Studies have also observed rapid increase in blood TL in response to ambient PM
13 [36,37]. In addition, some studies report contradictory results. For example, “The Normative
14 Aging Study” found an association between shorter telomeres in and long-term exposures to
15 airborne particles rich in black carbon [38]. A study on the association between arsenic
16 exposure in drinking water and TL showed longer telomeres in peripheral blood after exposure
17 to arsenic acid [39]. They also reported a positive association between urine arsenic levels and
18 telomerase reverse transcriptase gene (TERT) expression.

19 These findings suggest that carcinogenicity of arsenic among other compounds can be
20 explained by extending the lifespan of possible malignant cells by elongation of the telomeres
21 [39]. A similar observation was made in a Chinese female population, where an association
22 was observed between CLPTM1L-TERT polymorphism, longer TL (measured in peripheral
23 blood) and the risk of lung cancer [40]. A study by Jones et al. [41] also found an association
24 between the telomerase RNA component (TERC) polymorphisms and both longer telomeres
25 and susceptibility to colorectal cancer. This suggests that SNPs close to TERC can have
26 functional effects on TERC expression and thus on TL. We acknowledge that only TL was
27 evaluated but other important TL regulating factors, including telomerase activity, epigenetic
28 factors may further explain our findings. In the context of other diseases and carcinogenesis,
29 both positive and negative associations with TL are observed. In a study by Haycock et al. [42],
30 associations between TL and the risk of cancer and non-neoplastic diseases were studied using
31 a Mendelian randomization study. This study has shown that genetically increased TL is

1 associated with an increased risk of several cancers, e.g. melanoma, lung cancer, chronic
2 lymphocytic leukaemia, which has been confirmed by several other prospective observational
3 or Mendelian randomization studies [43,44]. It is important to know that these results should
4 be interpreted as a reflection of the average association at the population level. However,
5 these findings are contradictory to those based on retrospective studies, tending to report an
6 association between shorter telomeres and an increased risk for cancer [45,46]. A plausible
7 explanation, proposed by Aviv et al., [47], for the relation between increased TL and cancer,
8 is the potential accumulation of mutation, due to telomere lengthening associated stem-like
9 properties, similar to the findings of Haycock et al. [42]. Although it is not possible to connect
10 our findings with an increased risk in cancer, it must be said that some studies show an
11 association between long somatic telomeres and some forms of cancer.

12 We also would like to recognize some strengths and limitations of our study. First, we
13 have a small-sized study and our results need to be confirmed in a larger independent
14 investigation. Nevertheless, we observed robust associations independent of the effect of age,
15 sex, smoking behaviour, alcohol consumption, BMI. We have a well characterized exposure
16 group. Finally, the measurements are conducted only at one time point and therefore a follow-
17 up of the same subject would provide a better interpretation of the present findings.

18 **5. Conclusion**

19 Overall, a higher mtDNAcn and longer telomeres were observed in MWCNT exposed
20 workers when compared to non-exposed controls, independent of age, sex, smoking behavior,
21 alcohol consumption and BMI. When comparing the three groups of exposure (lab low, lab
22 high and operator), significant exposure-associated differences were observed between the
23 groups. While we observe significant change in mtDNAcn and TL in the MWCNT exposed
24 workers, no association can be made regarding possible disease outcome at the moment.

25

26 **Acknowledgement**

27 We like to acknowledge FWO post-doctoral fellowship for Manosij Ghosh (12W7718N) and
28 Dries S. Martens (12X9620N).

29 **References**

30 [1] IARC working group on the Evaluation of Carcinogenic Risks to Humans, Some

- 1 Nanomaterials and Some Fibres To Humans Some Nanomaterials and Some Fibres,
2 2017.
- 3 [2] J.D. Beard, A. Erdely, M.M. Dahm, M.A. de Perio, M.E. Birch, D.E. Evans, J.E. Fernback,
4 T. Eye, V. Kodali, R.R. Mercer, S.J. Bertke, M.K. Schubauer-Berigan, Carbon nanotube
5 and nanofiber exposure and sputum and blood biomarkers of early effect among U.S.
6 workers, *Environ. Int.* (2018). <https://doi.org/10.1016/j.envint.2018.04.004>.
- 7 [3] L.M. Fatkhutdinova, T.O. Khaliullin, O.L. Vasil'yeva, R.R. Zalyalov, I.G. Mustafin, E.R.
8 Kisin, M.E. Birch, N. Yanamala, A.A. Shvedova, Fibrosis biomarkers in workers exposed
9 to MWCNTs., *Toxicol. Appl. Pharmacol.* 299 (2016) 125–31.
10 <https://doi.org/10.1016/j.taap.2016.02.016>.
- 11 [4] A.A. Shvedova, N. Yanamala, E.R. Kisin, T.O. Khailullin, M.E. Birch, L.M. Fatkhutdinova,
12 T. Nurkiewicz, Integrated Analysis of Dysregulated ncRNA and mRNA Expression Profiles
13 in Humans Exposed to Carbon Nanotubes, *PLoS One.* 11 (2016) e0150628.
14 <https://doi.org/10.1371/journal.pone.0150628>.
- 15 [5] J.S. Lee, Y.C. Choi, J.H. Shin, J.H. Lee, Y. Lee, S.Y. Park, J.E. Baek, J.D. Park, K. Ahn, I.J. Yu,
16 Health surveillance study of workers who manufacture multi-walled carbon nanotubes,
17 *Nanotoxicology.* (2015). <https://doi.org/10.3109/17435390.2014.978404>.
- 18 [6] J. Vlaanderen, A. Pronk, N. Rothman, A. Hildesheim, D. Silverman, H.D. Hosgood, S.
19 Spaan, E. Kuijpers, L. Godderis, P. Hoet, Q. Lan, R. Vermeulen, A cross-sectional study
20 of changes in markers of immunological effects and lung health due to exposure to
21 multi-walled carbon nanotubes, *Nanotoxicology.* 11 (2017) 395–404.
22 <https://doi.org/10.1080/17435390.2017.1308031>.
- 23 [7] E. Kuijpers, A. Pronk, R. Kleemann, J. Vlaanderen, Q. Lan, N. Rothman, D. Silverman, P.
24 Hoet, L. Godderis, R. Vermeulen, Cardiovascular effects among workers exposed to
25 multiwalled carbon nanotubes, *Occup. Environ. Med.* (2018) oemed-2017-104796.
26 <https://doi.org/10.1136/oemed-2017-104796>.
- 27 [8] M. Ghosh, D. Öner, K. Poels, A.M. Tabish, J. Vlaanderen, A. Pronk, E. Kuijpers, Q. Lan, R.
28 Vermeulen, B. Bekaert, P.H. Hoet, L. Godderis, Changes in DNA methylation induced by
29 multi-walled carbon nanotube exposure in the workplace, *Nanotoxicology.* (2017) 1–
30 16. <https://doi.org/10.1080/17435390.2017.1406169>.
- 31 [9] A.N. Malik, A. Czajka, Is mitochondrial DNA content a potential biomarker of
32 mitochondrial dysfunction?, *Mitochondrion.* 13 (2013) 481–492.

- 1 <https://doi.org/10.1016/j.mito.2012.10.011>.
- 2 [10] A. Hori, M. Yoshida, T. Shibata, F. Ling, Reactive oxygen species regulate DNA copy
3 number in isolated yeast mitochondria by triggering recombination-mediated
4 replication, *Nucleic Acids Res.* 37 (2009) 749–761. <https://doi.org/10.1093/nar/gkn993>.
- 5 [11] B.G. Janssen, E. Munters, N. Pieters, K. Smeets, B. Cox, A. Cuypers, F. Fierens, J. Penders,
6 J. Vangronsveld, W. Gyselaers, T.S. Nawrot, Placental mitochondrial DNA content and
7 particulate air pollution during in utero life, *Environ. Health Perspect.* (2012).
8 <https://doi.org/10.1289/ehp.1104458>.
- 9 [12] L. Hou, Z.Z. Zhu, X. Zhang, F. Nordio, M. Bonzini, J. Schwartz, M. Hoxha, L. Dioni, B.
10 Marinelli, V. Pegoraro, P. Apostoli, P.A. Bertazzi, A. Baccarelli, Airborne particulate
11 matter and mitochondrial damage: A cross-sectional study, *Environ. Heal. A Glob.*
12 *Access Sci. Source.* (2010). <https://doi.org/10.1186/1476-069X-9-48>.
- 13 [13] H.C. Lee, Y.H. Wei, Mitochondrial role in life and death of the cell, *J. Biomed. Sci.* (2000).
14 <https://doi.org/10.1159/000025424>.
- 15 [14] T. von Zglinicki, Oxidative stress shortens telomeres., *Trends Biochem. Sci.* (2002).
- 16 [15] T. Richter, T. von Zglinicki, A continuous correlation between oxidative stress and
17 telomere shortening in fibroblasts, *Exp. Gerontol.* (2007).
18 <https://doi.org/10.1016/j.exger.2007.08.005>.
- 19 [16] D. Jurk, C. Wilson, J.F. Passos, F. Oakley, C. Correia-Melo, L. Greaves, G. Saretzki, C. Fox,
20 C. Lawless, R. Anderson, G. Hewitt, S.L. Pender, N. Fullard, G. Nelson, J. Mann, B. van de
21 Sluis, D.A. Mann, T. von Zglinicki, Chronic inflammation induces telomere dysfunction
22 and accelerates ageing in mice, *Nat. Commun.* 5 (2014) 4172.
23 <https://doi.org/10.1038/ncomms5172>.
- 24 [17] D.S. Martens, T.S. Nawrot, Ageing at the level of telomeres in association to residential
25 landscape and air pollution at home and work: a review of the current evidence, *Toxicol.*
26 *Lett.* 298 (2018) 42–52. <https://doi.org/10.1016/J.TOXLET.2018.06.1213>.
- 27 [18] J. Vlaanderen, A. Pronk, N. Rothman, A. Hildesheim, D. Silverman, H.D. Hosgood, S.
28 Spaan, E. Kuijpers, L. Godderis, P. Hoet, Q. Lan, R. Vermeulen, A cross-sectional study
29 of changes in markers of immunological effects and lung health due to exposure to
30 multi-walled carbon nanotubes, *Nanotoxicology.* (2017).
31 <https://doi.org/10.1080/17435390.2017.1308031>.
- 32 [19] E. Kuijpers, C. Bekker, W. Fransman, D. Brouwer, P. Tromp, J. Vlaanderen, L. Godderis,

- 1 P. Hoet, Q. Lan, D. Silverman, R. Vermeulen, A. Pronk, Occupational Exposure to Multi-
2 Walled Carbon Nanotubes during Commercial Production Synthesis and Handling, *Ann.*
3 *Occup. Hyg.* (2016). <https://doi.org/10.1093/annhyg/mev082>.
- 4 [20] D.S. Martens, M. Plusquin, W. Gyselaers, I. De Vivo, T.S. Nawrot, Maternal pre-
5 pregnancy body mass index and newborn telomere length, *BMC Med.* 14 (2016) 148.
6 <https://doi.org/10.1186/s12916-016-0689-0>.
- 7 [21] R.M. Cawthon, Telomere length measurement by a novel monochrome multiplex
8 quantitative PCR method, *Nucleic Acids Res.* (2009).
9 <https://doi.org/10.1093/nar/gkn1027>.
- 10 [22] H.C. Lee, P.H. Yin, C.W. Chi, Y.H. Wei, Increase in mitochondrial mass in human
11 fibroblasts under oxidative stress and during replicative cell senescence, *J. Biomed. Sci.*
12 (2002). <https://doi.org/10.1007/BF02254978>.
- 13 [23] P.R. Smith, J.M. Cooper, G.G. Govan, A.E. Harding, A.H.V. Schapira, Smoking and
14 mitochondrial function: a model for environmental toxins, *QJM.* (2012).
15 <https://doi.org/10.1093/qjmed/86.10.657>.
- 16 [24] H.C. Lee, C.Y. Lu, H.J. Fahn, Y.H. Wei, Aging- and smoking-associated alteration in the
17 relative content of mitochondrial DNA in human lung, *FEBS Lett.* (1998).
18 [https://doi.org/10.1016/S0014-5793\(98\)01564-6](https://doi.org/10.1016/S0014-5793(98)01564-6).
- 19 [25] H.J. Fahn, L.S. Wang, S.H. Kao, S.C. Chang, M.H. Huang, Y.H. Wei, Smoking-associated
20 mitochondrial DNA mutations and lipid peroxidation in human lung tissues, *Am. J.*
21 *Respir. Cell Mol. Biol.* (1998). <https://doi.org/10.1165/ajrcmb.19.6.3130>.
- 22 [26] A.M. James, M.P. Murphy, How Mitochondrial Damage Affects Cell Function, *J. Biomed.*
23 *Sci.* (2003). <https://doi.org/10.1159/000064721>.
- 24 [27] W.W. Jiang, B. Masayeva, M. Zahurak, A.L. Carvalho, E. Rosenbaum, E. Mambo, S.
25 Zhou, K. Minhas, N. Benoit, W.H. Westra, A. Alberg, D. Sidransky, W. Koch, J. Califano,
26 Increased mitochondrial DNA content in saliva associated with head and neck cancer,
27 *Clin. Cancer Res.* (2005). <https://doi.org/10.1158/1078-0432.CCR-04-2147>.
- 28 [28] H.D. Hosgood, C.S. Liu, N. Rothman, S.J. Weinstein, M.R. Bonner, M. Shen, U. Lim, J.
29 Virtamo, W. ling Cheng, D. Albanes, Q. Lan, Mitochondrial DNA copy number and lung
30 cancer risk in a prospective cohort study, *Carcinogenesis.* (2010).
31 <https://doi.org/10.1093/carcin/bgq045>.
- 32 [29] B. Thyagarajan, R. Wang, H. Nelson, H. Barcelo, W.P. Koh, J.M. Yuan, Mitochondrial DNA

- 1 Copy Number Is Associated with Breast Cancer Risk, *PLoS One*. (2013).
2 <https://doi.org/10.1371/journal.pone.0065968>.
- 3 [30] A.C. Rice, P.M. Keeney, N.K. Algarzae, A.C. Ladd, R.R. Thomas, J.P. Bennett,
4 Mitochondrial DNA copy numbers in pyramidal neurons are decreased and
5 mitochondrial biogenesis transcriptome signaling is disrupted in Alzheimer's disease
6 hippocampi, *J. Alzheimer's Dis.* (2014). <https://doi.org/10.3233/JAD-131715>.
- 7 [31] B.G. Masayeva, E. Mambo, R.J. Taylor, O.G. Goloubeva, S. Zhou, Y. Cohen, K. Minhas,
8 W. Koch, J. Sciubba, A.J. Alberg, D. Sidransky, J. Califano, Mitochondrial DNA content
9 increase in response to cigarette smoking, *Cancer Epidemiol. Biomarkers Prev.* (2006).
10 <https://doi.org/10.1158/1055-9965.EPI-05-0210>.
- 11 [32] D. Tan, D.S. Goerlitz, R.G. Dumitrescu, D. Han, F. Seillier-Moisewitsch, S.M. Spernak,
12 R.A. Orden, J. Chen, R. Goldman, P.G. Shields, Associations between cigarette smoking
13 and mitochondrial DNA abnormalities in buccal cells, *Carcinogenesis*. (2008).
14 <https://doi.org/10.1093/carcin/bgn034>.
- 15 [33] S. Pavanello, L. Dioni, M. Hoxha, U. Fedeli, D. Mielzynska-Švach, A.A. Baccarelli,
16 Mitochondrial dna copy number and exposure to polycyclic aromatic hydrocarbons,
17 *Cancer Epidemiol. Biomarkers Prev.* (2013). <https://doi.org/10.1158/1055-9965.EPI-13-0118>.
- 18
- 19 [34] H.C. Lee, P.H. Yin, C.Y. Lu, C.W. Chi, Y.H. Wei, Increase of mitochondria and
20 mitochondrial DNA in response to oxidative stress in human cells., *Biochem. J.* (2000).
- 21 [35] N. Pieters, G. Koppen, K. Smeets, D. Napierska, M. Plusquin, S. De Prins, H. Van De
22 Weghe, V. Nelen, B. Cox, A. Cuypers, P. Hoet, G. Schoeters, T.S. Nawrot, Decreased
23 Mitochondrial DNA Content in Association with Exposure to Polycyclic Aromatic
24 Hydrocarbons in House Dust during Wintertime: From a Population Enquiry to Cell
25 Culture, *PLoS One*. (2013). <https://doi.org/10.1371/journal.pone.0063208>.
- 26 [36] L. Dioni, M. Hoxha, F. Nordio, M. Bonzini, L. Tarantini, B. Albeti, A. Savarese, J.
27 Schwartz, P.A. Bertazzi, P. Apostoli, L. Hou, A. Baccarelli, Effects of short-term exposure
28 to inhalable particulate matter on telomere length, telomerase expression, and
29 telomerase methylation in steel workers, *Environ. Health Perspect.* (2011).
30 <https://doi.org/10.1289/ehp.1002486>.
- 31 [37] L. Hou, S. Wang, C. Dou, X. Zhang, Y. Yu, Y. Zheng, U. Avula, M. Hoxha, A. Díaz, J.
32 McCracken, F. Barretta, B. Marinelli, P.A. Bertazzi, J. Schwartz, A.A. Baccarelli, Air

- 1 pollution exposure and telomere length in highly exposed subjects in Beijing, China: A
2 repeated-measure study, *Environ. Int.* (2012).
3 <https://doi.org/10.1016/j.envint.2012.06.020>.
- 4 [38] J. Mccracken, A. Baccarelli, M. Hoxha, L. Dioni, S. Melly, B. Coull, H. Suh, P. Vokonas, J.
5 Schwartz, Annual ambient black carbon associated with shorter telomeres in elderly
6 men: Veterans affairs normative aging study, *Environ. Health Perspect.* (2010).
7 <https://doi.org/10.1289/ehp.0901831>.
- 8 [39] H. Li, K. Engström, M. Vahter, K. Broberg, Arsenic exposure through drinking water is
9 associated with longer telomeres in peripheral blood, *Chem. Res. Toxicol.* (2012).
10 <https://doi.org/10.1021/tx300222t>.
- 11 [40] Q. Lan, R. Cawthon, Y. Gao, W. Hu, H.D. Hosgood, F. Barone-Adesi, B.-T. Ji, B. Bassig, W.-
12 H. Chow, X. Shu, Q. Cai, Y. Xiang, S. Berndt, C. Kim, S. Chanock, W. Zheng, N. Rothman,
13 Longer Telomere Length in Peripheral White Blood Cells Is Associated with Risk of Lung
14 Cancer and the rs2736100 (CLPTM1L-TERT) Polymorphism in a Prospective Cohort
15 Study among Women in China, *PLoS One.* 8 (2013) e59230.
16 <https://doi.org/10.1371/journal.pone.0059230>.
- 17 [41] A.M. Jones, A.D. Beggs, L. Carvajal-Carmona, S. Farrington, A. Tenesa, M. Walker, K.
18 Howarth, S. Ballereau, S. V. Hodgson, A. Zauber, M. Bertagnolli, R. Midgley, H. Campbell,
19 D. Kerr, M.G. Dunlop, I.P.M. Tomlinson, TERC polymorphisms are associated both with
20 susceptibility to colorectal cancer and with longer telomeres, *Gut.* (2012).
21 <https://doi.org/10.1136/gut.2011.239772>.
- 22 [42] P.C. Haycock, S. Burgess, A. Nounu, J. Zheng, G.N. Okoli, J. Bowden, K.H. Wade, N.J.
23 Timpson, D.M. Evans, P. Willeit, A. Aviv, T.R. Gaunt, G. Hemani, M. Mangino, H.P. Ellis,
24 K.M. Kurian, K.A. Pooley, R.A. Eeles, J.E. Lee, S. Fang, W. V. Chen, M.H. Law, L.M.
25 Bowdler, M.M. Iles, Q. Yang, B.B. Worrall, H.S. Markus, R.J. Hung, C.I. Amos, A.B.
26 Spurdle, D.J. Thompson, T.A. O'Mara, B. Wolpin, L. Amundadottir, R. Stolzenberg-
27 Solomon, A. Trichopoulou, N.C. Onland-Moret, E. Lund, E.J. Duell, F. Canzian, G. Severi,
28 K. Overvad, M.J. Gunter, R. Tumino, U. Svenson, A. Van Rij, A.F. Baas, M.J. Bown, N.J.
29 Samani, F.N.G. Van t'Hof, G. Tromp, G.T. Jones, H. Kuivaniemi, J.R. Elmore, M.
30 Johansson, J. Mckay, G. Scelo, R. Carreras-Torres, V. Gaborieau, P. Brennan, P.M. Bracci,
31 R.E. Neale, S.H. Olson, S. Gallinger, D. Li, G.M. Petersen, H.A. Risch, A.P. Klein, J. Han,
32 C.C. Abnet, N.D. Freedman, P.R. Taylor, J.M. Maris, K.K. Aben, L.A. Kiemeny, S.H.

1 Vermeulen, J.K. Wiencke, K.M. Walsh, M. Wrensch, T. Rice, C. Turnbull, K. Litchfield, L.
2 Paternoster, M. Standl, G.R. Abecasis, J.P. SanGiovanni, Y. Li, V. Mijatovic, Y. Sapkota,
3 S.K. Low, K.T. Zondervan, G.W. Montgomery, D.R. Nyholt, D.A. Van Heel, K. Hunt, D.E.
4 Arking, F.N. Ashar, N. Sotoodehnia, D. Woo, J. Rosand, M.E. Comeau, W.M. Brown, E.K.
5 Silverman, J.E. Hokanson, M.H. Cho, J. Hui, M.A. Ferreira, P.J. Thompson, A.C. Morrison,
6 J.F. Felix, N.L. Smith, A.M. Christiano, L. Petukhova, R.C. Betz, X. Fan, X. Zhang, C. Zhu,
7 C.D. Langefeld, S.D. Thompson, F. Wang, X. Lin, D.A. Schwartz, T. Fingerlin, J.I. Rotter,
8 M.F. Cotch, R.A. Jensen, M. Munz, H. Dommisch, A.S. Schaefer, F. Han, H.M. Ollila, R.P.
9 Hillary, O. Albagha, S.H. Ralston, C. Zeng, W. Zheng, X.O. Shu, A. Reis, S. Uebe, U.
10 Hüffmeier, Y. Kawamura, T. Otowa, T. Sasaki, M.L. Hibberd, S. Davila, G. Xie, K.
11 Siminovitch, J.X. Bei, Y.X. Zeng, A. Försti, B. Chen, S. Landi, A. Franke, A. Fischer, D.
12 Ellinghaus, C. Flores, I. Noth, S.F. Ma, J.N. Foo, J. Liu, J.W. Kim, D.G. Cox, O. Delattre, O.
13 Mirabeau, C.F. Skibola, C.S. Tang, M. Garcia-Barcelo, K.P. Chang, W.H. Su, Y.S. Chang,
14 N.G. Martin, S. Gordon, T.D. Wade, C. Lee, M. Kubo, P.C. Cha, Y. Nakamura, D. Levy, M.
15 Kimura, S.J. Hwang, S. Hunt, T. Spector, N. Soranzo, A.W. Manichaikul, R.G. Barr, B.
16 Kahali, E. Speliotes, L.M. Yerges-Armstrong, C.Y. Cheng, J.B. Jonas, T.Y. Wong, I. Fogh,
17 K. Lin, J.F. Powell, K. Rice, C.L. Relton, R.M. Martin, G. Davey Smith, Association between
18 telomere length and risk of cancer and non-neoplastic diseases a mendelian
19 randomization study, JAMA Oncol. (2017).
20 <https://doi.org/10.1001/jamaoncol.2016.5945>.

21 [43] C. Zhang, J.A. Doherty, S. Burgess, R.J. Hung, S. Lindström, P. Kraft, J. Gong, C.I. Amos,
22 T.A. Sellers, A.N.A. Monteiro, G. Chenevix-Trench, H. Bickeböller, A. Risch, P. Brennan,
23 J.D. McKay, R.S. Houlston, M.T. Landi, M.N. Timofeeva, Y. Wang, J. Heinrich, Z. Kote-
24 Jarai, R.A. Eeles, K. Muir, F. Wiklund, H. Grönberg, S.I. Berndt, S.J. Chanock, F.
25 Schumacher, C.A. Haiman, B.E. Henderson, A.A. Al Olama, I.L. Andrulis, J.L. Hopper, J.
26 Chang-Claude, E.M. John, K.E. Malone, M.D. Gammon, G. Ursin, A.S. Whittemore, D.J.
27 Hunter, S.B. Gruber, J.A. Knight, L. Hou, L. Le Marchand, P.A. Newcomb, T.J. Hudson,
28 A.T. Chan, L. Li, M.O. Woods, H. Ahsan, B.L. Pierce, Genetic determinants of telomere
29 length and risk of common cancers: A Mendelian randomization study, Hum. Mol.
30 Genet. (2015). <https://doi.org/10.1093/hmg/ddv252>.

31 [44] K.M. Walsh, V. Codd, T. Rice, C.P. Nelson, I. V. Smirnov, L.S. McCoy, H.M. Hansen, E.
32 Elhauge, J. Ojha, S.S. Francis, N.R. Madsen, P.M. Bracci, A.R. Pico, A.M. Molinaro, T.

- 1 Tihan, M.S. Berger, S.M. Chang, M.D. Prados, R.B. Jenkins, J.L. Wiemels, E.C.T. Group,
2 N.J. Samani, J.K. Wiencke, M.R. Wrensch, Longer genotypically-estimated leukocyte
3 telomere length is associated with increased adult glioma risk, *Oncotarget*. (2015).
4 <https://doi.org/10.18632/oncotarget.6468>.
- 5 [45] H. Ma, Z. Zhou, S. Wei, Z. Liu, K.A. Pooley, A.M. Dunning, U. Svenson, G. Roos, H.D.
6 Hosgood, M. Shen, Q. Wei, Shortened Telomere length is associated with increased risk
7 of cancer: A meta-analysis, *PLoS One*. (2011).
8 <https://doi.org/10.1371/journal.pone.0020466>.
- 9 [46] I.M. Wentzensen, L. Mirabello, R.M. Pfeiffer, S.A. Savage, The association of telomere
10 length and cancer: A meta-analysis, *Cancer Epidemiol. Biomarkers Prev.* (2011).
11 <https://doi.org/10.1158/1055-9965.EPI-11-0005>.
- 12 [47] A. Aviv, J.J. Anderson, J.W. Shay, Mutations, Cancer and the Telomere Length Paradox,
13 *Trends in Cancer*. (2017). <https://doi.org/10.1016/j.trecan.2017.02.005>.
14
15

1 **Table 1:** Demographic characteristics of the study population reported as N, (%) and mean \pm
 2 SD.

Variables		Control (N = 43)	Exposed (N = 24)	P-value
Sex	Male	32 (74.4 %)	20 (83.3 %)	0.409
	Female	11 (25.6 %)	4 (16.7 %)	
Age (years)		34.6 \pm 8.57	35.9 \pm 6.90	0.729
BMI (kg/m²)		24.88 \pm 4.71	27.19 \pm 4.99	0.064
Smoking	Never smoker	24 (55.8 %)	13 (54.2 %)	0.805
	Former smoker	7 (16.3 %)	6 (25.0 %)	
	Current smoker	12 (27.9 %)	5 (20.8 %)	
Alcohol consumption	Yes/No	35/8	15/9	0.091
	Glasses/day ^b	1.1 \pm 0.91	0.9 \pm 0.77	
Previous history of exposure to chemicals^a		9 (20.9 %)	8 (33.3 %)	
Duration of exposure to nanoparticle at current job (years)		/	4.25 \pm 2.40	

3 ^a as reported by the study subjects.

4 ^b Alcoholic drinks consumed on average per day over the past 4 weeks.

5

6

7

8 **Table 2:** Association of mtDNA and TL with CNT exposure.

	mtDNA (ND1)			mtDNA (hmito3)			TL		
	n	% difference (95%CI)	P-value	n	% difference (95%CI)	P-value	n	% difference (95%CI)	P-value
MWCNT exposure									
Non-exposed controls	33	Ref		32	Ref		33	Ref	
MWCNT exposed	23	35.2 (19.1, 53.5)	<0.0001	24	37.4 (-20.6, 92.8)	0.068	24	18.3 (7.2, 30.6)	0.001
Detailed exposure groups									
Non-exposed controls	33	Ref		32	Ref		33	Ref	
Lab low (1 µg/m ³ EC)	9	26.5 (6.4, 50.7)	0.008	9	23.0 (-22.6, 95.9)	0.38	9	27.1 (10.9, 45.6)	0.001
Lab high (7 µg/m ³ EC)	6	35.5 (10.2, 66.3)	0.004	6	8.9 (-37.2, 88.8)	0.76	6	8.4 (-7.7, 27.4)	0.33
Operator (45 µg/m ³ EC)	6	40.6 (13.8, 73.4)	0.002	7	61.4 (-5.6, 176.7)	0.081	7	18.9 (1.4, 39.0)	0.033

9 ^aEstimates from the linear regression models provided as a % difference (95%CI) in outcome compared with the non-exposed group (Ref); MWCNT- Multi-walled carbon nanotube; relative
10 telomere length (TL), mitochondrial copy number (mtDNAcn); ND1 (mitochondrial encoded NADH dehydrogenase 1); Hmito3 (Human mitochondrial genome); Models adjusted for age, sex,
11 smoking behaviour, alcohol consumption and BMI

12