

Fluoroquinolone resistance in *Escherichia coli* isolates after exposure to non-fluoroquinolone antibiotics: a retrospective case–control study

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1 **Fluoroquinolone resistance in *Escherichia coli* isolates after exposure**  
2 **to non-fluoroquinolone antibiotics: a retrospective case-control**  
3 **study**

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19 **Running title:** Fluoroquinolone resistance in *E. coli* after exposure to other antibiotics

## 20 Synopsis

21 **Objectives:** To investigate whether prior exposure to non-fluoroquinolone antibiotics increases the risk  
22 of fluoroquinolone resistance in *Escherichia coli* (*E. coli*).

23 **Methods:** This was a secondary analysis of data collected retrospectively in a case-control study linking  
24 microbiological test results (isolated bacteria and their susceptibility) of urine samples routinely  
25 collected in primary, secondary and tertiary care patients in Belgium with information on prior  
26 antibiotic use at the patient level up to one year prior.

27 **Results:** In urine samples from 6125 patients, 7204 *E. coli* isolates were retrieved; 1949  
28 fluoroquinolone resistant isolates (cases) and 5255 fluoroquinolone susceptible isolates (controls).  
29 After adjusting for potential confounders (including fluoroquinolone use) and correcting for multiple  
30 testing, there was lower odds of fluoroquinolone resistance in *E. coli* isolates after exposure to  
31 cefazolin (OR = 0.65; 95% CI: 0.52 – 0.81;  $p = 0.00034$ ) and higher odds after exposure to  
32 trimethoprim/sulfamethoxazole (OR = 1.79; 95% CI, 1.38 - 2.31;  $p < 0.001$ ) and after exposure to  
33 nitrofurantoin (OR: 1.41, 95% CI: 1.14 - 1.76;  $p < 0.002$ ). A sensitivity analysis excluding samples with  
34 antibiotic use in the 6 months prior to the sampling date, confirmed the higher odds of fluoroquinolone  
35 resistance after exposure to trimethoprim/sulfamethoxazole and nitrofurantoin.

36 **Conclusions:** Assuming no residual confounding or other biases, this study suggests that exposure to  
37 non-fluoroquinolone antibiotics, i.e. trimethoprim/sulfamethoxazole and nitrofurantoin, might be  
38 causally related to fluoroquinolone resistance in *E. coli* isolates from urinary samples. Future  
39 prospective research is needed to confirm non-fluoroquinolone antibiotics as potential drivers of  
40 fluoroquinolone resistance.

## 41 Introduction

42 Fluoroquinolones, a major class of antibiotics, belong to the WHO list of highest priority antimicrobials  
43 for resistance surveillance in human medicine.<sup>1</sup> In various European countries, considerable increases  
44 in ciprofloxacin resistance in *Escherichia coli* (*E. coli*) causing uncomplicated urinary tract infections  
45 were found between 2000 and 2014.<sup>2</sup> Although a slight decrease in fluoroquinolone resistance has  
46 been reported in Belgium (from 26.7% in 2014 to 23.8% in 2017) it was still high compared to other  
47 northern European countries, such as the Netherlands (14.2% in 2017) and Denmark (12.8% in 2017).<sup>3</sup>

48 While fluoroquinolone use is likely the main driver of resistance to fluoroquinolones,<sup>4-11</sup> we need to  
49 consider other risk factors. Previous studies in *Enterococcus* species, *K. pneumoniae*, *P. aeruginosa* and  
50 *E. coli* found older age,<sup>4</sup> long term care facility residency,<sup>5</sup> recent hospitalization,<sup>9</sup> the use of urinary  
51 catheter,<sup>7</sup> and previous exposure to non-fluoroquinolone antibiotics,<sup>5, 10, 12-14</sup> as independent risk  
52 factors for fluoroquinolone resistance. The link between non-fluoroquinolones and fluoroquinolone  
53 resistance can be explained by the phenomenon of co-selection.<sup>15, 16</sup>

54 Even though investigators have identified several non-fluoroquinolone antibiotics as independent risk  
55 factors to fluoroquinolone resistance,<sup>4, 5, 12-14, 17</sup> most of these studies had no information on prior  
56 antibiotic use, i.e. before hospitalization, only included hospitalized patients and had small sample  
57 sizes, and in some case-control studies, controls were taken from the same hospital, with a high  
58 probability of selection bias and resulting in a persistent effect of a previous selection pressure of an  
59 antimicrobial.

60 Identification of a causal relationship between non-fluoroquinolone antibiotic use and fluoroquinolone  
61 resistance could guide treatment options to reduce fluoroquinolone resistance and enhance our  
62 understanding of co-selection. Therefore, we investigated the increased risk of fluoroquinolone  
63 resistance in urinary *E. coli* samples collected from primary, secondary and tertiary health care settings  
64 in Belgium by the use of non-fluoroquinolone antibiotics in the year prior to the sampling date. We  
65 hypothesized that we would find associations between non-fluoroquinolone antibiotic use and

66 fluoroquinolone resistance. Additionally, we assessed effect modification of this relation by exposure  
67 to fluoroquinolones.

68

## 69 **Methods**

### 70 Study design and setting

71 We performed a secondary analysis of data collected retrospectively from primary, secondary and  
72 tertiary care patients in Belgium during a case-control study. Microbiological test results (1<sup>st</sup> January  
73 2011 to 31<sup>st</sup> December 2012) were linked with data on prior antibiotic use at the patient level (1<sup>st</sup>  
74 January 2010 until 31<sup>st</sup> December 2012) up to one year before sample collection and susceptibility  
75 testing for fluoroquinolones.<sup>18</sup> More information on the data sources, linkage, data access and cleaning  
76 methods, is available as Supplementary material S1.

77

### 78 Study population

79 In the linked dataset, including 197,393 urine samples, in 21,569 (10.9%) samples *E. coli* was isolated.  
80 For 16,593 (76.9%) of these *E. coli* strains, ciprofloxacin susceptibility was tested, and in 1,743 (10.5%),  
81 106 (0.6%), 3,415 (20.6%), and 11,329 (68.3%) the test result was unknown, intermediate, resistant,  
82 and susceptible, respectively. After selecting only antibiotic use one year prior to sampling testing, we  
83 obtained a total of 8,400 samples. From this data, we excluded samples from the same patient within  
84 the next 30 days as this cut-off point is often used to differentiate between the same and new urinary  
85 tract infection episode<sup>16</sup>; resulting in a total of 7,204 samples. For the purpose of this study, *E. coli*  
86 strains with unknown susceptibility test results were excluded from the analysis, cases were defined  
87 as *E. coli* isolates with intermediate and resistant test results and controls were defined as *E. coli*  
88 isolates with susceptible test results.

89 Exposure and outcome assessment

90 The exposures of interest were the consumption of antibacterials for systematic use, i.e. substances  
91 with Anatomical Therapeutic Chemical (ATC) code J01.<sup>19</sup> These antibiotics were studied at the chemical  
92 substance level, e.g. doxycycline (ATC J01AA02), and at the chemical subgroup level, e.g. tetracyclines  
93 (ATC J01AA), separately. We use the terms *antibiotics alone* for the chemical substance level and *group*  
94 *of antibiotics* for the chemical sub-group level. Should a causal relation between exposure to an  
95 antibiotic and fluoroquinolone resistance exist, one would expect a similar relation for the group of  
96 antibiotics to which it belongs. We considered a patient's sample exposed to an antibiotic if that  
97 antibiotic was reimbursed to the patient up to one year prior to their fluoroquinolone susceptibility  
98 testing. The inputs for all exposures were categorical (Yes/No).

99 The primary outcome was fluoroquinolone resistance indicated as resistant or intermediate in the  
100 ciprofloxacin (fluoroquinolone) susceptibility test of *E. coli* isolates from urine samples. The  
101 susceptibility testing was mainly done by the Kirby Bauer disk diffusion technique according to CLSI  
102 guidelines. Modifications were present according to the manufacturer for deviations in disk charge or  
103 diameter. The majority of Belgian hospitals worked with Neosensitabs for producing these  
104 antibiograms.<sup>20</sup>

105

106 Covariates

107 We considered potential confounders to be any variable suspected to be linked to both  
108 fluoroquinolone resistance and non-fluoroquinolone antibiotic use. Confounding variables were  
109 extracted from the antibiotic use and socio-demographic data, and included prior or current use of  
110 fluoroquinolones (Yes/No), whether the urine sample was taken in a hospital (Yes/No), the total  
111 number of days that a patient stayed in the hospital the last 6 months before the urine sample was  
112 taken (zero if the patient was not hospitalized in the last 6 months), the most recent time between any  
113 antibiotic use and the susceptibility test for fluoroquinolones resistance (in days), age (in years), gender  
114 (female/male) and any other antibiotic use. We also assessed modification of the relationship between

115 non-fluoroquinolones and fluoroquinolone resistance by fluoroquinolone use for the secondary aim of  
116 the study.

117 Statistical methods

118 Categorical variables were described using proportions, while continuous variables were summarized  
119 using medians and IQRs. Significance of differences in median values (for continuous variables) or  
120 proportions (for categorical variables) among cases and controls were assessed using Wilcoxon rank  
121 sum or Fisher's exact tests, respectively. The (2x2) contingency table analyses included only antibiotics  
122 with a least 5 counts per cell.

123 To quantify the relation between fluoroquinolone resistance and the use of each of the non-  
124 fluoroquinolone antibiotics (both alone and as group), a generalized estimating equation (GEE)  
125 approach for a binary outcome was used to accommodate dependence in observations related to the  
126 same individual,<sup>21</sup> thereby producing crude ORs with their 95% CIs.

127 In model 1, a multivariable GEE approach was used to adjust for confounding variables. Since  
128 multicollinearity was not an issue when adding all covariates in an initial full model (to check for  
129 multicollinearity we used (generalized) variance inflation factors, calculated using the car package in  
130 R, which had values smaller than 4) and our sample size was large enough, we conducted model  
131 building in a backward fashion by removing all non-significant ( $p < 0.15$ ) covariates. In model 2, we  
132 further examined effect modification of fluoroquinolone use only for non-fluoroquinolone  
133 antimicrobials that were found to be significant in model 1.

134 The Quasi-likelihood under the Independence model Criterion (QIC) was used to choose between an  
135 independence or exchangeable working correlation structure. A Bonferroni correction for multiple  
136 testing was applied to control for the overall Type I error. As a sensitivity analysis, we re-analyzed the  
137 available data excluding samples with antibiotic use in the 6 months prior to the sampling date to  
138 minimize the 'memory-like' correlations of resistance.<sup>22</sup>

139 All statistical analyses were performed using the statistical R software, version 3.5.1.<sup>23</sup> Unless stated  
140 otherwise (cf. in case of a Bonferroni correction), a 5% significance level was used for inference. The  
141 final data for this study consisted of complete cases for which no missing information regarding the  
142 determinants of the outcome under study was present.

143 Ethical statement

144 The study protocol were approved by the Sectorial committee Social Security and Health of the former

145 Privacy Commission (now Data Protection Authority; SCSZG/13/274) as well as by the ethics committee

146 of Antwerp University Hospital (reference: 14/4/27).<sup>24</sup> This manuscript adheres to the STROBE and the

147 RECORD-PE reporting guidelines.<sup>25 26</sup>

## 148 Results

149 The final data used for this study provides information on the fluoroquinolone resistance status for  
150 7,204 *E. coli* isolates identified in urine samples of 6,125 patients. Most patients were female (77%).  
151 The number of isolates per patient widely varied, where the minimum number was one and the  
152 maximum was 10. Our analysis included 1,949 case samples from 1,601 patients and 5,255 control  
153 samples from 4,710 patients.

154 Significant differences in characteristics between cases and controls were observed (Table 1). Cases  
155 were older than controls, with a difference of 10 years in median values. The median length of stay at  
156 the hospital was longer in cases than controls, with a difference of 12 days in medians. The median  
157 time between any antibiotic use and a susceptibility test for fluoroquinolones was longer in controls  
158 (77 days) than cases (38 days), with a difference of 39 days in median values (Figure 1). The proportion  
159 of samples from females was lower in cases than in controls (66.70% and 80.36%, respectively). The  
160 proportion of urine samples taken in the hospital was slightly lower in cases than in controls (12.98%  
161 and 13.17%, respectively). Moreover, the proportion of fluoroquinolone use was higher in cases than  
162 in controls (73.01% and 32.33%, respectively). Among the different types of fluoroquinolones used,  
163 ciprofloxacin was the most frequently used and its use was higher in cases than in controls (52.03%  
164 and 19.12%, respectively).

165  
166 Univariable analysis

167 Exposures: Table 2 shows the crude ORs with 95% CIs. For antibiotics alone, only the following  
168 antibiotics showed a significant association with fluoroquinolone resistance in *E. coli* isolates:  
169 tetracycline, ampicillin, amoxicillin, temocillin, benzylpenicillin, cefadroxil, cefuroxime, ceftazidime,  
170 ceftriaxone, meropenem, trimethoprim/sulfamethoxazole, amikacin, nitrofurantoin, nifurtinol, and  
171 fosfomycin. For group of antibiotics, only penicillin with extended spectrum, penicillin &  $\beta$ -lactamase

172 inhibitors, second and third generation cephalosporins, lincosamides, other aminoglycosides,  
173 glycopeptides, nitrofurantoin derivatives and other antibacterials did.

174

175 **Multivariable** analysis

176 **Model 1:** For *antibiotics alone*, after adjusting for the predefined confounders and correcting for  
177 multiple testing (the number of tests performed was 14 based on the covariates finally included in the  
178 multivariable model; Bonferroni correction:  $0.05/14 = 0.004$ ), only cefazolin,  
179 trimethoprim/sulfamethoxazole and nitrofurantoin showed a significant association with  
180 fluoroquinolone resistance. Next to these three antibiotics, the final model also adjusted for  
181 fluoroquinolone use, age, time of length of stay at the hospital, most recent time between any  
182 antibiotic use and the susceptibility test, gender, the use of vancomycin, temocillin, metronidazole,  
183 roxithromycin, cefadroxil and cefuroxime. After exposure to cefazolin, there was lower odds of  
184 fluoroquinolone resistance in *E. coli* isolates (OR = 0.65; 95% CI: 0.52 – 0.81;  $p = 0.00034$ ) as compared  
185 to no exposure to this antibiotic. There was a higher odds of fluoroquinolone resistance in *E. coli*  
186 isolates after exposure to trimethoprim/sulfamethoxazole (OR = 1.79; 95% CI: 1.38 - 2.31;  $p = 0.00023$ )  
187 and after exposure to nitrofurantoin (OR: 1.41; 95% CI: 1.14 - 1.76;  $p = 0.00013$ ) compared to no  
188 exposure to these antibiotics, respectively (Table 2).

189 As for *group of antibiotics*, after adjusting for confounders and correcting for multiple testing (Nine  
190 covariates were finally included in the multivariable model; Bonferroni correction:  $0.05/9 = 0.006$ ), only  
191 first generation cephalosporins and nitrofurantoin derivatives were significantly associated with  
192 fluoroquinolone resistance in *E. coli* isolates in urine samples. Next to these two groups of antibiotics,  
193 the final model also adjusted for fluoroquinolones use, age, time of length of stay at the hospital, most  
194 recent time between any antibiotic use and the susceptibility test, gender, whether the sample was  
195 taken at hospital, the use of trimethoprim/sulfamethoxazole and imidazole. The odds of  
196 fluoroquinolone resistance in *E. coli* isolates was lower after exposure to first generation

197 cephalosporins (OR = 0.65; 95% CI: 0.53 – 0.81;  $p < 0.0001$ ) and higher after exposure to nitrofurantoin  
198 derivatives (OR = 1.38; 95% CI: 1.17 - 1.64;  $p = 0.00015$ ) (Table 2).

199 Table 3 shows the results of fitting model 2 to the data and stratifying by fluoroquinolone use. Only  
200 the interaction term between the use of fluoroquinolones and trimethoprim/sulfamethoxazole  
201 exposure was significant ( $p = 0.004$ ). In those using fluoroquinolones, the association between  
202 exposure to trimethoprim/sulfamethoxazole and fluoroquinolone resistance was found to be non-  
203 significant (OR = 1.13; 95% CI: 0.87 - 1.46), whereas the association was significant in those that did  
204 not use fluoroquinolones (OR = 2.75; 95% CI: 1.87 - 4.05). The estimated exposure effects of non-  
205 fluoroquinolone antibiotics on fluoroquinolone resistance were always smaller in those who used  
206 fluoroquinolones than in those who did not use fluoroquinolones. However, fluoroquinolone  
207 resistance was always higher in those exposed to fluoroquinolones as compared to those not exposed  
208 to fluoroquinolones regardless of the use of non-fluoroquinolone antibiotics.

209  
210 Sensitivity analysis

211 In 1445 cases and 3821 controls there were no samples with antibiotic use in the 6 months prior to the  
212 sampling date. Only trimethoprim/sulfamethoxazole and nitrofurantoin were consistently associated  
213 with fluoroquinolone resistance and no substantial changes were found in the association between  
214 fluoroquinolone resistance and non-fluoroquinolone antibiotic use, after adjusting for confounders  
215 and correcting for multiple testing (the number of tests performed in the final model was 14; corrected  
216  $p$ -value = 0.004): trimethoprim/sulfamethoxazole: OR = 1.63; 95% CI; 1.19 - 2.23;  $p = 0.0023$ ;  
217 nitrofurantoin: OR = 1.51; 95% CI, 1.18 - 1.93;  $p = 0.0011$ .

## 218 Discussion

219 Our study suggests a potentially causal relation between non-fluoroquinolone antibiotics use and  
220 fluoroquinolone resistance in *E. coli* isolates identified in urine samples which were routinely collected  
221 in primary, secondary and tertiary care patients in Belgium. Particularly, exposure to trimethoprim-  
222 sulfamethoxazole and nitrofurantoin in the year prior to the sample collection and susceptibility testing  
223 was significantly associated with fluoroquinolone resistance. These relations were confirmed when  
224 excluding samples with antibiotic use in the 6 months prior to sampling and susceptibility testing.  
225 Regarding the effect of fluoroquinolone use, the effect of exposure to non-fluoroquinolone antibiotics  
226 to fluoroquinolone resistance was smaller in those who also used fluoroquinolones than in those who  
227 did not use fluoroquinolones. Fluoroquinolone resistance was always higher in those who used  
228 fluoroquinolones as in those who did not.

229

### 230 Comparison with other studies

231 Previous studies found associations between use of several non-fluoroquinolone antibiotics and  
232 fluoroquinolone resistance. The use of ceftriaxone was correlated with a decreased susceptibility of *E.*  
233 *coli* isolates to ciprofloxacin in in-patients;<sup>12</sup> the use of aminoglycosides in the last 30 days increased  
234 the risk of ciprofloxacin more than fivefold,<sup>5</sup> whereas treatment with aminoglycosides in a year prior  
235 to sampling increased the risk of infection with levofloxacin resistance by ten-fold<sup>13</sup> in nosocomial  
236 bacteremia due to extended-spectrum  $\beta$ -lactamase-producing *E.coli* and *Klebsiella pneumoniae* (*K.*  
237 *pneumoniae*). In another study, aminoglycoside use in the previous 30 days was found to increase  
238 levofloxacin resistance by over eight-fold in nosocomial *E. coli* and *K. pneumoniae* infections.<sup>4</sup>  
239 Aminoglycosides and ceftriaxone were less frequently used compared to other antibiotics in our study  
240 population, which might explain why we were not able to show an association with fluoroquinolone  
241 resistance.

242 A three-fold risk of levofloxacin resistance with the use of “other types of antibiotics” was found in  
243 *Enterococcus faecalis* isolates from in-patients with urinary tract infection (UTI).<sup>10</sup> Another study

244 showed that the use of  $\beta$ -lactamase inhibitors and extended spectrum cephalosporins and clindamycin,  
245 in the previous 30 days, increased levofloxacin resistance by over nine-fold in hospitalized patients  
246 with UTI caused by *Enterococcus* species.<sup>14</sup> Meanwhile, we did not observe associations between  
247 fluoroquinolone resistance and any of these antibiotics, despite having a large sample size.

248 In line with our findings, a recent large study that analyzed data at a higher geographical level and  
249 explored whether higher levels of non-fluoroquinolone antibiotics were associated with higher  
250 fluoroquinolone resistance in *E. coli* isolates, found that the use of trimethoprim and derivatives was  
251 associated with increased ciprofloxacin resistance, i.e. use over one year, three and one month before  
252 an antibiotic susceptibility test (AST), but not for the use of nitrofurantoin.<sup>16</sup> Another recent large study  
253 at the patient level observed that the purchase of trimethoprim was associated with ciprofloxacin  
254 resistance which is also consistent with our findings.<sup>22</sup> The association between the use of  
255 sulfamethoxazole/trimethoprim and fluoroquinolone resistance might be explained by the selection  
256 of resistance genes that code for resistance against both fluoroquinolones and  
257 sulfamethoxazole/trimethoprim by the use sulfamethoxazole/trimethoprim, as for example found in  
258 *E. coli* sequence type ST131 isolates.<sup>27</sup> On the other hand, and given that the strains were not available  
259 for further investigations, we are not able to provide an biological mechanisms for the higher odds of  
260 fluoroquinolone resistance after exposure to nitrofurantoin found in our study since, to our  
261 knowledge, no previous studies found such associations and neither found co-selection resistant genes  
262 in urinary pathogens coding for both nitrofurantoin and fluoroquinolone resistance.

263

#### 264 Strengths and limitations

265 We used data at patient level for each antibiotic use and for each corresponding susceptibility test in  
266 a large linked dataset from primary, secondary and tertiary healthcare settings. Our study findings fit  
267 with previous studies,<sup>4, 5, 10, 12-14</sup> although not exactly the same non-fluoroquinolones antibiotics were  
268 identified. In turn, we attempted to minimized reverse causality by excluding samples with antibiotic  
269 use in the 6 months prior to a susceptibility test.

270 Furthermore, we used a longer patient history of antibiotic use in a larger sample of patients. This  
271 enabled a more granular disaggregation of the effects of specific antibiotic classes in different periods  
272 before a formal susceptibility test was performed and using hospitalized and non-hospitalized patients  
273 from which a urinary sample was taken and where *E. coli* was identified, and hence, helps to narrow  
274 the possible effect of exposure to antibiotics on resistance to fluoroquinolones. Another strength is  
275 that we used a GEE approach which considers clustering of samples from the same patient, while other  
276 approaches such as logistic regression would have required selection of one sample per patient, by  
277 doing this, we would have lost very valuable and from an evolutionary microbiological perspective  
278 relevant information.

279 Even though one can argue that the potential for confounding by indication, contraindication or  
280 disease severity could mislead our results, statistical control for age and gender to some extent covers  
281 these possible confounders, for instance, co-morbidities require most often antibiotic treatment which  
282 is linked to age; regarding morbidity such as UTIs, older and female patients in the database are more  
283 likely to have more than one urinary sample tested, and women are also more vulnerable to UTIs.

284 Finally, because there is a concern when using multiple samples from patients as prior resistance is one  
285 of the main predictors of future resistant samples,<sup>22</sup> we have conducted an additional sensitivity  
286 analysis using only the first sample per patient, which provided similar results as well (data not shown).  
287 Moreover, as a main sensitivity analysis we excluded samples with antibiotic use in the 6 months prior  
288 to a sampling date to reduce potential memory-like correlations of resistance,<sup>22</sup> considering that the  
289 choice of antibiotic for treatment may be influenced by prior AST results. Yet, given the nature of our  
290 data reverse causality cannot be completely excluded, despite using individual-level data

291  
292 Regarding potential limitations, our study is a retrospective observational study. Hence, we missed  
293 other potential confounders, such as the presence of a urinary catheter.<sup>7</sup> Even though we adjusted for  
294 the setting in which the urinary sample was taken, though more likely, not all hospital samples are  
295 catheter specimens of urine, and not all outpatient samples are midstream specimens. Additionally,

296 we treated patients as independent individuals without considering a broader range of important  
297 factors such as household membership,<sup>28</sup> food intake, socioeconomic  
298 factors,<sup>29</sup> and variation in prescribing among practices and areas,<sup>30, 31</sup> thus limiting the chances of  
299 meeting the Stable Unit Treatment Value Assumption (SUTVA) that underlies standard regression  
300 models.<sup>16</sup> Nevertheless, it is a strength that the previous study on data grouped at higher levels<sup>16</sup> and  
301 ours on data at the individual level, each having their strengths and weaknesses, agreed that  
302 trimethoprim and derivatives (trimethoprim/sulfamethoxazole) use may select for fluoroquinolone  
303 resistance.

304

305 We were not able to assess the effect of all non-fluoroquinolone antibiotics present in the dataset due  
306 to limited use of these antibiotics in our study sample and not being able to cope with small cell counts  
307 by using statistical techniques, e.g. Firth correction.<sup>16</sup> Misclassification bias of the measurement of the  
308 exposures is possible as we only have information on the reimbursement of purchased antibiotics and  
309 not on actual consumption and antibiotics can be purchased and consumed without being reimbursed.  
310 However, we believe overall compliance to antibiotic treatment in Belgium is high, particularly starting  
311 a course of purchased antibiotics, and patients do not need to end the course for antibiotics to select  
312 for resistant bacteria. Hence, misclassification bias of the exposures is less likely to affect our findings.  
313 Since our study population consisted of samples that are more likely to be taken and sent for a  
314 resistance test, e.g. in case of treatment failure,<sup>16, 32</sup> this might have confounded our estimates.

315 Another relative issue is that we did not measure the exposure using DDD, which would rather have  
316 included the volume of use in the model enabling the demonstration of a dose-response effect, which  
317 consequently could have led to more convincing evidence in favour of a causal relation as described in  
318 one of the Bradford-Hill criteria.<sup>33</sup> Finally, the generalizability of our findings might not be applicable  
319 to other European countries since co-resistance and antibiotic use trends vary across countries.

320 Implications for future studies

321 Future studies should be conducted prospectively, collecting information on all potential confounders,  
322 not only at the hospital level but also at the community level or **geographical level**, and ideally measure  
323 actual exposure. The fact that microorganisms can share resistance genes even without the trigger of  
324 exposure to antibiotics, requires assessment of a broader range of factors that could lead to increase  
325 resistance, (i.e. household membership,<sup>28</sup> food intake, socioeconomic factors,<sup>29</sup> and **variation in**  
326 **prescribing among practices and areas.**<sup>30, 31</sup>). Another consideration is that after exposure the time to  
327 selection of resistant bacteria will be more similar across antibiotics than the persistence of  
328 resistance,<sup>34, 35</sup> while the latter might also differ by pathogen. Regarding the assessment of a dose-  
329 response relationship, both the number of DDD per treatment and the number of treatments should  
330 be assessed. **Finally, further studies should be performed in patients that were not tested for antibiotic**  
331 **resistance for at least two years prior to a first AST result to assess fluoroquinolone resistance in UTIs**  
332 **due to *E. coli*,<sup>22</sup> to prevent confounding of the relation between non-fluoroquinolone use and**  
333 **fluoroquinolone resistance due to 'reverse causality'.**

334

335 In conclusion, our study suggests a (causal) association between exposure to non-fluoroquinolone  
336 antibiotics, specifically to trimethoprim/sulfamethoxazole and nitrofurantoin, and fluoroquinolone  
337 resistance in *E. coli* isolates from urinary samples. **Assuming no residual confounding or other biases**  
338 **previously discussed**, this study implies that co-selection could drive fluoroquinolone resistance after  
339 exposure to non-fluoroquinolone antibiotics especially in those not exposed to fluoroquinolones.  
340 Further prospective evidence however is quintessential to confirm which non-fluoroquinolone  
341 antibiotics increase the risk of fluoroquinolone resistance.

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## 355 **Transparency to declare**

356 The funders had an advisory role in the design and data collection of the overall study but were not

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## 358 **Author contributions**

359 L.C-P., S.C, R.B., S.A., B.C., designed the study and analysis. B.C., K.L. contributed data. L.C-P., R.B.,

360 B.C., K.L., S.A, H.G., and S.C, participated in the interpretation of the results and writing the

361 manuscript. All authors read and approved the final manuscript.

## 362 **Data accessibility**

363 Original databases at patient level are no longer available in accordance with restriction from the

364 Sector Committee. However, final data used for this study can be requested for further analyses

365 upon formal request and under strict supervision of Sciensano.

366 **Code availability**

367 Code used for data analysis is available upon request.

368

369 **Supplementary material**

370 More information on the data sources, linkage, data access and cleaning methods is available in

371 Supplementary material S1.

372 The RECORD statement for pharmacoepidemiology (RECORD-PE) checklist, extended from the

373 STROBE and RECORD statements is reported as Supplementary material S2.

374

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**Table 1. Characteristics of 7204 *E. coli* isolates identified in urine samples of 6125 patients in primary, secondary, and tertiary healthcare settings in Belgium, 2011-2012. Controls are susceptible to fluoroquinolones according to applied disk diffusion test, cases are not (intermediate + resistant).**

<b>Exposure one year prior to susceptibility test</b>	<b>Cases (N= 1949), n (%)</b>	<b>Controls (N= 5255), n (%)</b>	<b>P</b>
Median age (IQR), years	76 (63 - 83)	66 (37 - 81)	< 0.0001 <sup>a</sup>
Gender, female	1300 (66.70)	4223 (80.36)	< 0.0001
Urine sample taken in the hospital	253 (12.98)	692 (13.17)	0.88
Median length of stay at the hospital (IQR), days	21 (5 - 54)	9 (2 -34)	< 0.0001 <sup>a</sup>
Median of time between any antibiotic use and the susceptibility test (IQR), days	38 (14 - 104)	77 (25 – 175)	< 0.0001 <sup>a</sup>
Fluoroquinolones (J01MA) use	1423 (73.01)	1699 (32.33)	< 0.0001
Ofloxacin	85 (4.36)	92 (1.75)	< 0.0001
Ciprofloxacin	1014 (52.03)	1005 (19.12)	< 0.0001
Norfloxacin	152 (7.80)	219 (4.17)	< 0.0001
Levofloxacin	205 (10.52)	189 (3.60)	< 0.0001
Moxifloxacin	400 (20.52)	426 (8.11)	< 0.0001

<sup>a</sup> Wilcoxon sum rank test; all other comparisons were made by Fisher's exact test.

**Table 2. Exposure to non-fluoroquinolone antibiotics, controls are susceptible to fluoroquinolones, cases are not. Univariable and multivariable analysis of non-fluoroquinolone antibiotic use one year prior to a susceptibility test for fluoroquinolones in 7204 *E. coli* isolates identified in urine samples of 6125 patients in primary, secondary and tertiary healthcare setting in Belgium (2011-2012).**

Exposure one year prior to susceptibility test	Cases (N= 1949), n (%)	Controls (N= 5255), n (%)	Crude OR (95% CI)	Adjusted OR (95% CI)
<b>Antibiotics alone</b>				
Doxycycline	45 (2.31)	114 (2.17)	1.07 (0.73 - 1.55)	-
Tetracycline	8 (0.41)	40 (0.76)	0.47 (0.24 - 0.95)	-
Minocycline	19 (0.97)	57 (1.08)	1.15 (0.67 - 1.96)	-
Ampicillin	19 (0.97)	91 (1.73)	0.96 (0.57 - 1.60)	-
Amoxicillin	434 (22.27)	1552 (29.53)	0.68 (0.60 - 0.78)	-
Temocillin	45 (2.31)	42 (0.80)	2.93 (1.85 - 4.64)	1.83 (1.07 - 3.14)
Benzylpenicillin	7 (0.36)	10 (0.19)	2.91 (1.08 - 7.81)	-
Oxacillin	6 (0.31)	17 (0.32)	0.95 (0.40 - 2.29)	-
Flucloxacillin	102 (5.23)	219 (4.17)	1.27 (0.97 - 1.66)	-
Amoxicillin - clavulanic acid	766 (39.30)	1940 (36.92)	1.11 (0.98 - 1.25)	-
Piperacillin - clavulanic acid	60 (3.07)	90 (1.71)	1.38 (0.96 - 1.99)	-
Cefazolin	200 (10.26)	534 (10.16)	1.01 (0.83 - 1.23)	0.65 (0.52 - 0.81) <sup>a</sup>
Cefadroxil	9 (0.46)	78 (1.48)	0.31 (0.15 - 0.62)	0.43 (0.18 - 1.01)
Cefuroxime	296 (15.19)	601 (11.44)	1.39 (1.16 - 1.65)	1.24 (1.01 - 1.53)
Ceftazidime	48 (2.46)	57 (1.08)	1.65 (1.04 - 2.60)	-
Ceftriaxone	45 (2.31)	68 (1.29)	1.80 (1.20 - 2.71)	-
Meropenem	28 (1.44)	39 (0.74)	2.31 (1.40 - 3.82)	-
Trimethoprim/sulfamethoxazole	266 (13.65)	368 (7.00)	2.10 (1.73 - 2.54)	1.79 (1.38 - 2.31) <sup>b</sup>
Erythromycin	16 (0.82)	42 (0.80)	1.18 (0.68 - 2.04)	-
Roxithromycin	10 (0.51)	18 (0.34)	1.50 (0.62 - 3.63)	2.07 (0.84 - 5.12)
Clarithromycin	113 (5.80)	288 (5.48)	1.06 (0.83 - 1.35)	-
Azithromycin	106 (5.44)	333 (6.34)	0.85 (0.67 - 1.09)	-
Clindamycin	117 (6.00)	248 (4.72)	1.27 (0.97 - 1.66)	-
Lincomycin	12 (0.62)	20 (0.38)	1.62 (0.77 - 3.42)	-
Gentamicin	11 (0.56)	21 (0.40)	1.41 (0.65 - 3.09)	-
Amikacin	30 (1.54)	44 (0.84)	1.70 (1.03 - 2.80)	-
Vancomycin	45 (2.31)	76 (1.45)	1.61 (1.04 - 2.49)	0.61 (0.37 - 1.01)
Metronidazole	40 (2.05)	94 (1.79)	1.30 (0.90 - 1.90)	0.62 (0.38 - 1.01)
Nitrofurantoin	296 (15.19)	480 (9.13)	1.78 (1.49 - 2.13)	1.41 (1.14 - 1.76) <sup>c</sup>
Nifurtinol	180 (9.24)	285 (5.42)	1.77 (1.44 - 2.19)	-
Fosfomycin	348 (17.86)	825 (15.70)	1.17 (1.00 - 1.36)	-
<b>Group of antibiotics</b>				
Tetracyclines	73 (3.75)	213 (4.05)	0.92 (0.69 - 1.23)	-
Penicillin with extended spectrum	475 (24.37)	1629 (31.00)	0.72 (0.63 - 0.82)	-
β-lactamase sensitive penicillin	10 (0.51)	30 (0.57)	1.54 (0.51 - 2.27)	-
β-lactamase resistant penicillin	106 (5.44)	232 (4.41)	1.25 (0.96 - 1.61)	-
Penicillin & β-lactamase inhibitors	790 (40.53)	1969 (37.47)	1.14 (1.01 - 1.28)	-
First generation cephalosporins	208 (10.67)	603 (11.47)	0.92 (0.76 - 1.11)	0.65 (0.53 - 0.81) <sup>d</sup>

Second generation cephalosporins	296 (15.19)	604 (11.49)	1.38 (1.16 - 1.64)	-
Third generation cephalosporins	83 (4.26)	122 (2.32)	1.87 (1.36 - 2.67)	-
Macrolides	243 (12.47)	694 (13.21)	0.94 (0.79 - 1.11)	-
Lincosamides	125 (6.41)	266 (5.06)	1.29 (1.00 - 1.65)	-
Other aminoglycosides	42 (2.15)	61 (1.16)	1.68 (1.11 - 2.54)	-
Glycopeptides	48 (2.46)	76 (1.45)	1.72 (1.12 - 2.64)	-
Imidazoles derivatives	43 (2.21)	97 (1.85)	1.43 (0.98 - 2.10)	-
Nitrofurans derivatives	447 (22.93)	738 (14.04)	1.82 (1.57 - 2.11)	1.38 (1.17 - 1.64) <sup>e</sup>
Other antibacterials	352 (18.06)	828 (15.76)	1.18 (1.01 - 1.37)	-

<sup>a</sup> p-value: 0.00016; <sup>b</sup> p-value: 0.0000098; <sup>c</sup> p-value: 0.00166; <sup>d</sup> p-value: 0.000083, <sup>e</sup> p-value: 0.00015

**Table 3. Effect modification by fluoroquinolone antibiotic use of the relation between fluoroquinolone resistance and non-fluoroquinolone antibiotic use in 7204 *E. coli* isolates identified in urine samples of 6125 patients in primary, secondary, and tertiary health care settings in Belgium (2011-2012).**

Exposure one year prior to susceptibility test	Not use of fluoroquinolones = 0	Use of fluoroquinolone = 1	<i>P</i> <sup>a</sup>
	Adjusted OR (95%CI)	Adjusted OR (95%CI)	
<b>Antibiotic alone</b>			
Cefazolin	0.65 (0.46 – 0.92)	0.74 (0.56 – 0.99)	0.15
Trimethoprim/sulfamethoxazole	2.75 (1.87 – 4.05)	1.13 (0.87 – 1.46)	0.004
Nitrofurantoin	1.49 (1.05 – 2.10)	1.44 (1.13 – 1.84)	0.96
<b>Group of antibiotics</b>			
First generation cephalosporins	0.61 (0.44 – 0.86)	0.71 (0.54 – 0.94)	0.15
Nitrofurans derivatives	1.28 (0.94 – 1.73)	1.40 (1.15 – 1.72)	0.44

<sup>a</sup> *p* value of the interaction term

**Figure 1. Histogram for the most recent time between any antimicrobial use and a susceptibility test for fluoroquinolones (one year prior) (days), with a median line stratified by cases and controls in *E. coli* isolates from urine samples retrieved from 6125 patients in primary, secondary, and tertiary healthcare centers in Belgium (2011-2012).**

